

RESEARCH COMMUNICATION

Hepatocyte Growth Factor in Egyptian Females with Breast Benign Lumps and Cancers

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Abstract

Background: Hepatocyte growth factor (HGF) also known as scatter factor (SF) and its receptor c-met play important roles in mammary differentiation and have been implicated in mammary carcinogenesis. **Objective:** Estimation of the plasma level of HGF in females with benign breast lumps or breast carcinomas and correlating levels with important prognostic parameters. **Subjects:** Sixty eight adult premenopausal females were divided into control group of fifteen healthy volunteers and fifty-three patients subdivided into fifteen with benign breast lumps and thirty-eight with breast carcinomas. **Methods:** A thorough clinical examination, plain chest x-rays, ultrasonography of the abdomen and pelvis, pre-operative fine needle aspiration cytology, estimation of fasting serum glucose, urea, creatinine and uric acid levels, alanine aminotransferase activities, C-reactive protein, HGF level and histopathological examination of the breast masses were performed. **Results:** Significant increase in HGF levels was found in patients with benign breast lumps and in breast cancer patients when each was compared to controls and when cancer patients were compared to the benign breast lumps group. **Conclusion:** The serum level of HGF is an independent prognostic indicator for breast cancer.

Keywords: Breast lumps - hepatocyte growth factor - metastasis

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Introduction

Most breast cancers evolve over a long latent period by several processes which require cell growth (mitogenicity), migration (motogenicity) and differentiation which remain mostly undetermined (Cao et al., 2006). Growth factors (GFs) are polypeptide molecules that bind to receptors on cell surface (Favoni et al., 2000). Hepatocyte GF, also known as scatter factor, Hepatopoietin A, lung fibroblast derived mitogen and Met-ligand, was originally discovered with liver regeneration (Liu et al., 2006).

Hepatocyte growth factor is synthesized as biologically inactive chain precursor (728 amino acids called pro-HGF) (Parr et al., 2004) which requires proteolytic processing, by various serine proteases to generate active two-chain form (HGF) in vitro (Ohnishi et al., 2006). It is primarily expressed in stromal cells and weakly in epithelial and endothelial cells (Peruzzi et al., 2006). Activator inhibitor type -1 and type -2 are two serine protease inhibitors with the ability to bind HGF activator and thus block its HGF-activating properties (Cao et al., 2006).

Hepatocyte GF promotes mammary ductal morphogenesis and in maintaining ductal spacing (Pollard et al., 2001) and is also expressed in "benign breast disease" detected as incidental microscopic findings (Guray et al., 2006).

Fibrocystic changes are the most frequent benign disorder of the breast (Guray et al., 2006). Cysts contain a variety of biologically active substances including steroids, cytokines, growth factors as well as proteins (Celis et al., 2006).

Previous workers found that expression of HGF and its receptor c-Met is correlated with breast carcinoma progression (Beviglia et al., 1999).

The study was done to estimate the serum level of HGF in females with benign breast lumps and females with breast carcinoma and correlating its level with the prognostic parameters.

Materials and Methods

The study involved sixty eight adult premenopausal females divided into three groups: Group 1; fifteen normal healthy volunteer females, Group 2; included fifteen females with benign breast lumps and Group 3 included thirty eight females with breast carcinoma. Pregnancy or intake of contraceptive hormonal therapy at the time of the study were excluded.

To all studied subjects, thorough clinical examination was done, plain x-ray chest, ultrasonography of the abdomen and pelvis and laboratory investigations included fasting serum (Burtis et al., 2006) glucose, urea, creatinine, uric acid, alanine aminotransferase and alkaline

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phosphatase activities. Serum HGF was quantitatively determined using a solid phase sandwich ELISA from Biosource human (HU) Kit (Cat number KAC 2212/ KAC 2211) USA (Sheen et al., 2005). Histopathological examination: Preoperative as fine needle aspiration cytology and post operative for breast mass after surgery were performed.

Data were analysed using SPSS soft ware package, version 11.5 (SPSS, Chicago, IL, USA) for statistical analysis. Presentation was in form of median and range for such non parametric data .

Results

A significant increase in age was found in breast cancer patients group when it was compared with its value in patients with benign breast lumps group .

As regard subjects with benign breast lumps (n=15), nine patients (60%) had fibroadenoma , four patients (26.6%) had fibrocystic disease of the breast and two

(13.4%) patients had chronic non specific inflammation.

All breast cancer patients (n=38); (100%) had infiltrating ductal carcinoma .

Thirty six patients (94.73%) had IDC grade II, two (5.26%) grade III.

Nine patients (23.7%) had tumor size ≤ 2cm in its biggest diameter (T1), twenty two patients (57.9%) had tumor size >2cm and ≤ 5cm (T2) and seven patients (18.4%) had tumor size > 5cm in the biggest diameter (T3) . According to tumor size and lymph node status , the patients were divided into three stages, five patients (13.2%) were stage I , eleven patients (28.9%) were stage II and twenty two patients (57.9%) were stage III.

Thirty two patients (84.2%) were (ER) positive and only six patients (15.8%) were (ER) negative . Twenty five patients (65.8%) were (PR) positive and thirteen patients (34.2%) were (PR) negative. Twenty patients (52.61%) were having prominent angiogenesis (>30/10 HPFS) and eighteen patients(47.4%) were having angiogenesis less than thirty (<30/10HPFS).

Table 1. Serum Levels of Fasting Glucose (FSG) , Urea (Ur) , Creatinine (Cr) , Uric acid (UA), Alanine aminotransferase (ALT) and Alkaline Phosphatase (AP) Activities and Hepatocyte Growth Factor (HGF) (pg/ml) in the Studied Groups

Group/Item		FSG (mg/dl)	Ur (mg/dl)	Cr (mg/dl)	UA (mg/dl)	ALT (U/L)	AP (U/L)	HGF (pg/ml)
Control group (group1) (n=15)	Median	90	25	0.9	4.4	12	82	311
	Min-max	82-96	18-30	0.6-1.3	4-5.7	8-17	47-92	177-814
Benign breast lumps group (group2) (n=15)	Median	88	18	0.8	4.6	16	74	554*
	Min-max	80-100	11-32	0.7-1.0	4.1-5.7	5-30	49-95	279-1220
Breast cancer patients group (group3) (n=38)	p							0.002
	Median	95*	26*	0.9	4.45	13.5	81	1073.5*3
	Min-max	82-100	16-40	0.6-1.3	4-6	5-40	39-260	56-2352
	P	0.017	NS					0.000
	P ₁	0.009	0.005					0.000

*P=Statistical difference from group 1 (control subjects).; *P₁=Statistical difference from group 2 (subjects with benign breast lumps).; NS=No significant difference.

Table 2. Median (and Range) of HGF for 38 Cases of Cancer Breast Presented According to Age, Tumor size, Grade, Lymph Nodes Involvement, Stage, ER, PR and Angiogenesis

	Classification	No	Median	Range	P	P ₁	P ₂
Age	≤50	32	1121.5	356-2352	0.186		
	>50	6	948.5	638-1244			
Tumor size	T1	9	1006	706-1570	0.728		
	T2	22	1141.5	356-2244			
	T3	7	1222	744-2352			
Grade	Grade II	36	1073.5	356-2352	0.472	0.223	0.359
	Grade III	2	872.5	491-1254			
Lymph node	LN=N0	11	1078	491-2532	0.159		
	LN ≤3	6	841	732-1286			
	LN>3	21	1118	356-2244			
Stage	Stage I	5	1021	891-1570	0.955		
	StageII	11	1165	491-2352			
	StageIII	22	1093.5	356-2244			
ER	positive	32	1045	356-2352	0.262		
	negative	6	1249	908-2244			
PR	positive	25	1078	356-2352	0.890		
	negative	13	1069	638-2244			
Angiogenesis	>30/10HPFs	20	966	356-2244	0.483		
	<30/10HPFs	18	1141.5	491-2352			

P= Statistical difference between first and second groups.; P₁= Statistical difference between first and third groups.; P₂= Statistical difference between second and third groups.

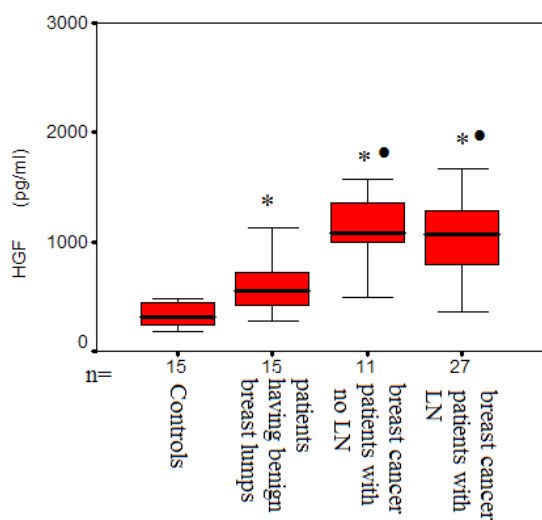


Figure 1. Box Plot Presentation of Serum Level of Hepatocyte Growth Factor (HGF) (pg/ml)

Table 1 shows serum level of fasting glucose (FSG), urea (Ur), creatinine (Cr), uric acid (UA), and serum alanine aminotransferase (ALT) and serum alkaline phosphatase (AP) activities and HGF (pg/ml) in the studied groups.

Significant increase in HGF level was found in patients with benign breast lumps group and in breast cancer patients group when each was compared with the control group.

A significant increase was also found in breast cancer patients group when compared to patients with benign breast lumps groups.

No difference was found between patients having breast cancer without lymph node metastasis and those with lymph node metastasis groups (Figure 1).

In breast cancer patients, no significant differences in serum level of HGF with any of the prognostic parameters (Table 2).

Figure 1 shows box plot presentation of serum level of HGF (pg/ml) in control group, patients with benign breast lumps and in patients having breast cancer without and with lymph node metastasis.

Discussion

Benign breast lumps and carcinoma of the breast are biologically different diseases. Several polypeptide hormones and growth factors have been shown to play an active role in breast epithelial cell growth and development (Hondermarck et al., 2003). Genetic aberration in growth factor signaling pathways are strongly connected with lumps (Cao et al., 2006).

Hepatocyte growth factor which belongs to the plasminogen prothrombin gene superfamily (Cao et al., 2006), plays a role in mammary gland development (Pollard et al., 2001).

Previous workers found that in breast cancer deregulated HGF and c-Met signaling may be found (Bonine-Summers et al., 2007).

In benign breast lumps group, the median level

of HGF was 554pg/ml (279-1220pg/ml) vs 311pg/ml (177-814pg/ml) which was significantly higher than its level in control group ($p=0.002$) (Table 1). This was not comparable with previous work (Sheen-Chen et al., 2005) that showed lower values.

In the present study, HGF level in breast cancer patients group was 1073.5pg/ml (356-2352pg/ml) (Table 1) which was comparable with results of previous workers (Taniguchi et al., 1995). However, Sheen-Chen SM et al., (2005) reported lower values. A significant increase in HGF level in whole breast cancer patients group when compared with control group and patients with benign breast lumps group ($p=0.000$ for both) (Table 1). This finding is in accordance with the results of other workers (Taniguchi et al., 1995; Tuck et al., 1996; Jin et al., 1997; Cianfrocca et al., 2004; Parr et al., 2004; Sheen-Chen et al., 2005).

By immunohistochemistry, HGF concentration was significantly higher in breast tumor tissues than benign lumps and normal tissues. Previous workers found that HGF is higher in tumor tissues than in adjacent normal tissues (Yamashita et al., 1994; Nagy et al., 1996; Rahimi et al., 1998; Favoni et al., 2000; Parr et al., 2004).

The increase in HGF, could be explained by: over stimulation of the cells that secrete HGF, or mutation and aberrant regulation of the HGF/Met signaling pathway or an imbalance between HGF activators and inhibitors.

In the present work, no significant difference was found in serum HGF level between patients having breast cancer with and without LN metastasis (Figure 1). As HGF median value was 1069 pg/ml (356-2244pg/ml) in patients having breast cancer with LN metastasis group and was 1078pg/ml (491-2352pg/ml) in patients without LN metastasis group.

In breast cancer patients, no significant difference was found in HGF and age. Similar findings are found by Sheen-Chen et al., (2005) and others (Yamashita et al., 1994; Wei-Kai et al., 2007). Also, no significant difference was found between HGF level and tumor size like others (Taniguchi et al., 1995; Sung et al., 2003) and tumor grade as only two patients were grade III and the rest were grade II.

Higher serum levels of HGF in patients with high grade tumor which does not agree with our study were found (Taniguchi et al., 1995; Sheen-Chen et al., 2005).

No significant difference in HGF level and LN metastasis which was found by Sung et al., (2003) (Sung et al., 2003).

Serum level of HGF in the current work, showed no significant difference and the stage of cancer. However, previous workers found that HGF levels were higher with high-stage disease (Sheen-Chen et al., 2005) and in patients with liver metastasis and in patients with multiple metastasis sites (Taniguchi et al., 1995; Eichbaum et al., 2007) compared with benign breast disease. This was not found as our patients were seeking medical advice before this very late stage.

In the present study, no significant associations in HGF level and hormone receptors (ER and PR) were found which goes in agreement with other work (Sung et al., 2003) also between tissue level of HGF in breast

cancer tissue and hormone receptors (Yamashita et al., 1944; Wei-Kai et al., 2003).

In the present study, serum level of HGF did not show significant difference with angiogenesis. Previous study (Yamashita et al., 1944) found no significant associations. However, other workers (Taniguchi et al., 1995; Sheen-Chen et al., 2005) found that HGF stimulates tumor invasion and neovascularization. So it may play important roles in tumor growth and metastasis.

In conclusion, serum level of HGF in benign breast lumps and breast cancer patients showed significant increase when compared with controls. A significant increase in its level was found in breast cancer patients as compared to benign breast lumps groups. HGF is an independent prognostic marker in breast cancer. More studies are needed on HGF receptors and of HGF activators and inhibitors.

References

- Beviglia L, Kramer RH (1999). HGF induces FAK activation and integrin - mediated adhesion in MLTn3 breast carcinoma cells. *Int J Cancer*, **83**, 640-9.
- Bonine-Summers AR, Aakre ME, Brown KA, et al (2007). Epidermal growth factor receptor plays a significant role in hepatocyte growth factor mediated biological responses in mammary epithelial cells. *Cancer Biol Ther*, **6**, 561-70.
- Burtis CA, Ashwood ER, Bruns DE (2006). Tietz Textbook of Clinical Chemistry and Molecular Diagnostics. 4 thEd . Elsevier Saunders Company. St Louis, 870-1, 802, 798, 807-8, 604-7, 609-10.
- Cao R, Bjorndahl MA, Gallego MI, et al (2006). Hepatocyte growth factor is a novel lymphangiogenic factor with an indirect mechanism of action. *Am Soc Hematol*, **19**, 2538-66.
- Celis JE, Gromov P, Moreira JMA, et al (2006). Apocrine cysts of the breast. Biomarkers, origin, enlargement and relation with cancer phenotype. *Mol Cell Proteomics*, **5**, 462-83.
- Cianfrocca M , Goldstein LJ (2004). Prognostic and predictive factors in early-stage breast cancer. *Oncologist*, **9**, 606-16.
- Eichbaum MH, de Rossi TM, Kaul S, et al (2007). Serum levels of hepatocyte growth factor / scatter factor in patients with liver metastasis from breast cancer. *Tumor Biol*, **28**, 36-44.
- Favoni RE, De Cupis A (2000). The role of polypeptide growth factors in human carcinomas: New targets for a novel pharmacological approach. *Pharmacol Rev*, **52**, 179-206.
- Guray M, Sahin AA (2006). Benign Breast Diseases: Classification, Diagnosis and Management. *Oncologist*, **11**, 435-49.
- Hondermarck H (2003). Breast cancer when proteomics challenge biological complexity. *Mol Cell Proteomics*, **2**, 281-91.
- Jin L, Fuchs A, Schnett SJ, et al (1997). Expression of scatter factor and c - met receptor in benign and malignant breast tissue. *Cancer*, **79**, 749-60.
- Liu Y (2006). Hepatocyte growth factor in kidney fibrosis: therapeutic potential and mechanisms of action. *Kidney Int*, **69**, 213-7.
- Nagy J, Curry GW, Hillan KJ, et al (1996). Hepatocyte growth factor /Scatter factor expression and c- met in primary breast cancer. *Surg Oncol*, **5**, 15- 21.
- Ohnishi T, Kakimoto K, Bandow K, et al (2006). Mature Hepatocyte growth factor /Scatter factor on the surface of human granulocytes is released by a mechanism involving activated Factor Xa. *J Immunol*, **176**, 6945-53.
- Parr C, Watkins G, Mansel RE, et al (2004). The hepatocyte growth factor regulatory factor, in human breast cancer. *Clin Cancer Res*, **10**, 202-11.
- Peruzzi B, Bottaro DP (2006). Targeting the c-Met signaling pathway in cancer. *Clin Cancer Res*, **12**, 3657-60.
- Pollard JW (2001). Tumor - stromal interactions: Transforming growth factor-beta isoforms and hepatocyte growth factor/scatter factor in mammary gland ductal morphogenesis. *Breast Cancer Res*, **3**, 230-7.
- Rahimi-Hung W, Tremblay E, Saulnier R, et al (1998). C-Src kinase activity is required for hepatocyte growth factor-induced motility and anchorage-independent growth of mammary carcinoma cells. *J Biol Chem*, **273**, 33714-21.
- Sheen-Chen SM, Liu YW, Eng HL, et al (2005). Serum levels of Hepatocyte Growth Factor in patients with breast cancer. *Cancer Epidemiol Biomarkers Prev*. **14**, 715-7.
- Sung HJ, Kim SY, Kim CK, et al (2003). Clinical usefulness of circulating hepatocyte growth factor (HGF) in breast cancer. *Korean J Lab Med*, **23**, 309-14.
- Taniguchi T, Toi M, Inada K, et al (1995). Serum concentrations of hepatocyte growth factor in breast cancer patients. *Clin Cancer Res*. **1**, 1031-4.
- Tuck AB, Park M, Sterns EE, et al (1996). Coexpression of hepatocyte growth factor and receptor (Met) in human breast carcinoma. *Am J Pathol*. **148**, 225-32.
- Wei-Kai H, Yu-xiu, Ting Y, et al (2007). Adipocytokines and breast cancer risk. *Chin Med J*, **120**, 1592-6.
- Yamashita JI, Ogawa M, Yamashita SI, et al (1994). Immunoreactive Hepatocyte Growth Factor is a strong and independent predictor of recurrence and Survival in Human Breast Cancer. *Cancer Res*. **54**, 1630.