RESEARCH ARTICLE

Expressional Correlation of Human Epidermal Growth Factor Receptor 2, Estrogen/Progesterone Receptor and Protein 53 in Breast Cancer

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Abstract

<u>Background</u>: This study aimed to show the localization of estrogen / progesterone receptors, human epidermal growth factor receptor 2 (Her-2) and protein 53 (p53) by immunohistochemistry in a series of consecutive breast cancer patients. <u>Materials and Methods</u>: The study covered invasive breast cancers from 299 patients presenting at the Oncogenetic Clinic and Pathology Centers of Ahwaz Jondishapour University of Medical Sciences Hospital in Iran during the time period from 2009 to 2011. The Scarff-Bloom Richardson scoring method was used. <u>Results</u>: Of the 299, 27% (80/299) were <40, 33% (100/299) were 41-50, and the remaining 40% (119/299) were>50 years old. The highest incidence of breast cancer in this study population was in the group of more than 50 year age, and the most common histological type of breast cancer was the invasive ductal carcinoma, which accounted for 68% (203/299) of the cases. Out of possible total of 207, 6% (13/207), 41% (85/207), and 53% (109/207) were scored as grade I, II, III, respectively. <u>Conclusion</u>: Our findings demonstrated a lack of association between labeling for the markers studied and tumor size and age of the patients. We confirmed an association between ER labeling and nuclear grade of breast cancer. The conflicting results obtained compared with the literature be because of differences in the immunohistochemical techniques applied in the various studies and to the scoring systems used.

Keywords: Breast cancer - estrogen receptor (ER) - progesterone receptor (PR) - p53

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Introduction

Breast cancer composes a remarkably diverse group of diseases regarding presentation, morphology, molecular profile and response to therapy. The risks of both breast cancer and death because of breast cancer are clearly increasing worldwide. Some 45% of the more than 1 million new cases of breast cancer diagnosed each year, and more than 55% of breast-cancer-related deaths, occur in low- and middle-income countries (Matsuda et al., 2013). Breast cancer affects Iranian women at least one decade younger than their counterparts in developed countries (Mousavi et al., 2007). For instance, in Iran it has been shown that, even after adjusting for age, young women are at higher risk for developing breast cancer than are their Western counterparts (Harirchi et al., 2010). The mortality rate of breast cancer was 5.8 per 100,000 women in Tehran in 1998 (Mohagheghi et al., 2010), 2.5 per 100,000 for female population, and 7762 years life lost in the 18 provinces of Iran in 2001 (Khosravi et al., 2007).

Breast cancer is not a single disease but a group of

several important tumors subtypes, each with a different natural history and each requiring a different treatment. Key factors such as tumor size, histological grade, vascular invasion, and nodal status are helpful, but increasing attention is being paid to the molecular features of the tumor (Schonborn et al., 1994). Many investigations have been performed about these regulators considering the role of steroid hormone and growth factor receptors, to growth and differentiation of both normal and malignant human breast cells. The discovery of the biomarkers opened a new view to diagnosis and treatment of these diseases.

Many studies of gene expression have identified expression profiles and gene sets that are prognostic, predictive, or both for patients with breast cancer (van de Vijver et al., 2002; Sorlie et al., 2003; Chang et al., 2004; 2005; Ma et al., 2004; Paik et al., 2004; Bertucci et al., 2005; Wang et al., 2005; Habashy et al., 2011; Patsialou et al., 2012; Tian et al., 2013). Prognostic and predictive biomarkers for breast cancer commonly used in clinical practice include estrogen receptor (ER) and progesterone receptor (PR) over-expression, oncogene over-expression c-erbB2, human epidermal growth factor receptor 2 (Her-

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2), protein 53 (p53), protein 21 (p21) and etc. Her-2 is a normal cellular gene that encodes a membrane protein 185 (P185) and its amplification plays an important role in the pathogenesis of breast cancer (Esteva-Lorenzo et al., 1998; Szoke and Udvarhelyi, 2012).

Her-2/neu over-expression tumors were shown to increase disease recurrence and metastasis and shorten survival (Cho et al., 2008; Szoke and Udvarhelyi, 2012; Dairkee et al., 2013). p53 is a tumor suppressor gene and an important component of breast cancer pathophysiology (Cho et al., 2008). The intensity of estrogene receptor (ER) expression in normal epithelium risk is a risk factor for breast cancer conferring a 3-fold increase in risk (Dairkee et al., 2013). Similarly to ER, PR has been found elevated very early in pre-malignant breast lesions at the hyperplasic enlarged lobular unit (Lagiou et al., 2009). In regarding to follow up difficulties cases in developing countries that lack screening programmes (as in Iran), we have studied localization of ER/PR, Her-2 and p53 by Immuno-histochemistry in consecutive breast cancer specimens submitted to pathology centers, and compared the labeling for these markers with histological type and grade of the cancers as well as type of breast cancer, age and size of tumor of these patients.

Materials and Methods

Patients

Patients with breast cancer and who had a family history of breast or ovarian cancer, or both, that was compatible with a dominant mode of inheritance were selected. We evaluated invasive breast cancers from 299 patients during time period from 2010 to 2013. These patients were asked to provide a blood sample and to sign an informed-consent form authorizing an analysis for analysis of molecular biomarkers. All procedures were approved by international guidelines and by the Institute Research Ethics and Use Committee of Ahwaz Jondishapour University of Medical Sciences (AJUMS). The samples were fixed in 10% formaldehyde solution. The tissue were processed routinely for embedding in paraffin wax and 5μ m thick sections were cut and placed on glass slides coated with 3-Aminopropyl Triethoxy Silane (APES) to enhance adhesion of sections to the slides for immuno-histochemistry. One slide of each tissue was stained with Hematoxylin and Eosin (H&E) to determine the histological type and grade of tumor.

Immunohistochemistry (IHC)

Briefly, 5-micron sections were cut, deparaffinized in xylene, rehydrated in a series of graded alcohols and placed in a tris buffer bath. Endogenous peroxidase activity was quenched using 3% hydrogen peroxide. Slides were rinsed with deionized water and placed in a tris buffer bath. After incubation, 1% preimmune goat serum was used to block nonspecific staining, and sections were stained with primary antibodies, respectively. Biotinylated link antibodies were added using the Labeled Streptavidin-biotin (LSAB) Kit (Dako), according to manufacturer's instructions. Detection was achieved using Diaminobenzidine (DAB) and H₂O₂ as substrates. All used antibodies were purchased from Dako. The following list includes all antibodies: c-erbB-2, rabbit antihuman c-erbB-2 oncoprotein; p53, mouse monoclonal antihuman p53; ER, mouse monoclonal antihuman ER; and PR, mouse monoclonal antihuman PR. Expression of ER/PR and p53 was graded as weakly positive (<10% the cells are stained), and positive when more than 10% tumor cell's nucleus stained, whereas absences of staining were considered as negative.

We have used the Scarff-Bloom Richardson scoring method (Eleston and Ellis, 2002). In short, the Scoring criteria for Her-2/neu were as follows: 1, Zero score defined tumors with no staining; 2, 1+ score refers to membrane staining (not continually) in less than 10% of tumor cells; 3, 2+ score which is characterized by weak to moderate complete membrane staining in more than 10% of the tumor cells; 4, 3+ score is defined as strong complete membrane staining in more than 10% of the tumor cells (high intensity). If the tumor was 0 or 1+, it was considered Her-2 negative and if the tumor was 2+ or 3+ Her-2 was positive. Tumor size was categorized macroscopically in to three classes, <2, 2-5 and >5 cm, respectively.

Statistical analysis

Statistical analysis was performed using SPSS version 13.0. Relationships between tumor markers and other parameters were studied using the test. Differences at p<0.05 were considered as statistically significant.

Results

Two hundred ninety nine (299) breast cancer women were included in this study; all of the samples had embedded in paraffin blocks. Of these 299, 27% (80/299) were <40, 33% (100/299) were 41-50, and the rest 40% (119/299) had >50 years old. The highest incidence of breast cancer in this study population was in the group of more than 50-year age and the most common histological type of breast cancer was the invasive ductal carcinoma, which accounted for 68% (203/299) of the cases. The tumor grades were performed only in 207 patients and for the rest grading was out of rule. Out of total 207, 6% (13/207), 41% (85/207), and 53% (109/207) were scored as grade I, II, III, respectively. IHC staining of four biomarkers (Her-2, ER, PR and p53) were performed for all patients. Relationship between different marker labeling and various known prognostic markers are

Table 1. Relationships between Marker Labeling andTape of Breast Cancer

Cancer Type (No. of cases)	Prognostic Markers (No. (%))					
	G1	G2	G3	G4		
Invasive ductal carcinoma (203)	76 (37.50)	104 (51.23)	100 (49.3)	101 (49.3)		
Inflamatory ductal carcinoma (45)	28 (62.20)	29 (64.50)	23 (51.1)	26 (57.8)		
Insitu ductal carcinoma (38)	15 (39.47)	17 (44.70)	19 (50.0)	16 (41.1)		
Medulary carcinoma (13)	10 (76.92)	8 (61.53)	12 (92.3)	11 (84.6)		
Total (299)	137 (45.81)	158 (52.84)	159 (53.2)	140 (46.8)		
P-value	0.006	0.022	0.034	0.042		

*Abbreviation: G1) ER-positive and/or PR-positive and HER2-negative; G**200.0** HER2-positive and ER-positive: G3) HER2-positive and ER-negative; and G4) Triple negative (negative for ER, PR and HER2)

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Grade (No. of cases)	Markers					
G1		G2	G3	G4		
	+ -	+ -	+ -	+ -		
I (13)	11(84.6) 2(15.4)	9(69.2) 4(30.8)	4(30.8) 9(69.2)	6(46.2) 7(53.8)		
II (80)	47(58.8) 33(41.2)	43(53.8) 37(46.2)	39(48.8) 41(51.2)	43(53.8) 37(46.2)		
III (109)	65(60.0) 44(40.0)	54(50.4) 55(50.6)	57(52.0) 52(48.0)	52(48.0) 57(52.0)		
Total (202) P-value	123(60.9) 79(39.1) 0.006	106(52.5) 96(47.5) 0.022	100(49.5) 102(51.5) 0.034	101(50.0)101(50.0) 0.042		

*Abbreviation: G1) ER-positive and/or PR-positive and HER2-negative; G2) HER2-positive and ER-positive: G3) HER2-positive and ER-negative; and G4) Triple negative (negative for ER, PR and HER2)

Table 3. Relationships between marker Labeling and Size of Breast Cancer

Size (No. of cases)	Markers					
	G1	G2	G3	G4		
	+ -	+ -	+ -	+ -		
Less than 2 cm (34)	21(61.8) 13(38.2)	21(61.8) 13(38.2)	13(38.2) 21(61.8)	20(58.8) 14(41.2)		
2-5 cm (169)	104(61.5) 65(38.5)	86(50.9) 83(49.1)	90(53.3) 79(46.7)	84(49.7) 85(50.3)		
More than 5 cm (10)	6(60.0) 4(40.0)	5(50.0) 5(50.0)	5(50.0) 5(50.0)	3(30.0) 7(70.0)		
Total (213)	131(61.5) 82(38.5)	112(52.6) 101(47.4)	108(50.7) 105(49.3)	107(50.2) 106(49.8)		
P-value	0.995	0.264	0.504	0.279		

*Abbreviation: G1) ER-positive and/or PR-positive and HER2-negative; G2) HER2-positive and ER-positive: G3) HER2-positive and ER-negative; and G4) Triple negative (negative for ER, PR and HER2)

Table 4. Relationships between Marker Labeling and Age

Age Groups (No. of cases)			Markers						
	(G1		G2		G3		G4	
	+	-	+	-	+	-	+	-	
<40 (67)	32(47.8)	35(52.2)	30(44.8)	37(55.2)	40(59.7)	27(40.3)	36(53.80)	31(46.2)	
40-50 (113)	77(68.2)	36(31.8)	71(62.8)	42(37.2)	55(48.7)	58(51.3)	55 (48)	58(52.0)	
>50 (119)	71(59.7)	48(40.3)	58(48.7)	61(51.3)	62(52.1)	57(47.9)	61(52.11)	58(47.9)	
Total (299)	180(60.2)	119(39.8)	159(53.2)	140(46.8)	157(52.5)	142(47.5)	152(50.80)	147(49.1)	
P-value	0.0	25	0.0)29	0	.356	0	.801	

*Abbreviation: G1) ER-positive and/or PR-positive and HER2-negative; G2) HER2-positive and ER-positive: G3) HER2-positive and ER-negative; and G4) Triple negative (negative for ER, PR and HER2)

Grade (No. of cases)	Markers				
	G1	G2	G3	G4	
	+ -	+ -	+ -	+ -	
I (17)	17(100) 0(0.00)	15(88.2) 2(11.8)	7(41.2) 10(58.8)	11(64.7) 6(35.3)	
II (138)	81(58.7) 57(41.3)	68(49.3) 70(50.7)	66(47.8) 72(52.2)	68(49.3) 70(50.7)	
III (46)	21(45.7) 25(54.3)	19(41.3) 27(58.7)	29(63.0) 17(37.0)	26(56.5) 20(43.5)	
Total (201)	119(59.2) 82(40.8)	102(50.7) 99(49.3)	102(50.7) 99(49.3)	105(52.2) 96(47.8)	
P-value	0.00049	0.003	0.144	0.395	

*Abbreviation: G1) ER-positive and/or PR-positive and HER2-negative; G2) HER2-positive and ER-positive: G3) HER2-positive and ER-negative; and G4) Triple negative (negative for ER, PR and HER2)

summarized in Tables 1-5.

A significant relationship (p<0.25) was demonstrated between the used biomarkers and the type of breast cancer. The results also showed that there is a significant relationship (p<0.006) among ER labeling and invasive and inflammatory ductal carcinoma. None of the biomarkers were demonstrated a significant relationship with Insitu ductal carcinoma but they have lonely a significant relationship (p<0.006) with medulary carcinoma (Table 1). All labeling markers were showed no significant correlation with any of grades (Table 2). All four biomarkers didn't have any significant relationship with tumor size and age of patients except ER and PR labeling had a significant correlations with 41-50 years (p<0.073) old and >50 old of ages (p<0.096)

Discussion

Nowadays molecular markers seem to have the potential improvement on our capacity to taking care of patients with, or at risk for breast cancer. In this study, we have compared labeling of four markers with some of known prognostic factors including histological type of cancer, histological grade, and nuclear grade, size of tumor and age of patients. In regarding to results each of biomarkers lonely has a significant association with the type of breast cancer. ER is expressed in about 70% of invasive breast cancer (Lee et al., 2007) and in our study ER labeling showed a significant relationship with invasive ductal carcinoma and medullary carcinoma. Inflammatory breast cancer is rare type of invasive

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breast cancer accounts for about 1% of all breast cancers; therefore, differentiated duct carcinomas of breast express ER and are generally, responsive to hormone therapy (Yenidunya et al., 2011; Fasching et al., 2012). PR expression commonly parallels that of ER expression, which is confirmed by the strong correlation between the labeling of the two receptors in the present study and other studies (Deblois and Giguère, 2013).

A number of clinical studies have documented an association between HER-2 amplification/overexpression and negative steroid hormone receptors (HR) status in breast tumors (Rosenthal et al., 2002; Muller et al., 2003; Jehoram et al., 2005; Ellis et al., 2006; Rody et al., 2009). In general, the higher the level of HER-2 overexpression and gene amplification will show the lower corresponding ER level. Our data also demonstrate an inverse correlation between HER-2 protein/gene levels and PR levels that most likely occurs because suppression of ER expression leads to reduced expression of PR. The negative correlation between HER-2 labeling and PR labeling supports the findings of others that HER-2 expression is a marker of poor prognosis.

The results of many studies suggest the p53 expression, HER-2 expression, and coexpressions of HER-2 and p53 have prognostic significance in breast cancer. Overexpression of HER-2 correlated strongly with poor patient survival (Ouyang et al., 2001; Yang et al., 2012) but contrasts with studies that suggest HER-2 over-expression has no (Erdem et al., 2005) or only limited prognostic value (Barnes et al., 1988). Coexpression of HER-2 and p53 has been reported in several studies, with frequency of coexpression as high as 42% (Rudas et al., 1997; Thor et al., 1998; Umekita et al., 2000; Kazkayasi et al., 2001; Bull et al., 2004; Skálová et al., 2009). Patients whose breast cancer tissues express HER-2 and p53 have been found to have a poor prognosis in several studies (Tsuda, 2009).

In conclusion, these conflicting results may be because of difference in the immunohistochemical techniques applied in the various studies and to the scoring systems used. Our findings also confirm lack of association between labeling for the markers studied and tumor size and age of the patients. We have confirmed the association between ER labeling and nuclear grade of breast cancer.

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References

- Barnes DM, Lammie GA, Millis RR, et al (1988). An immunohistochemical evaluation of c-erbB-2 expression in human breast carcinoma. *Br J Cancer.* 58, 448-52.
- Bertucci F, Finetti P, Rougemont J, et al (2005). Gene expression profiling identifies molecular subtypes of inflammatory breast cancer. *Cancer Res.* **65**, 2170-8.
- Bull SB, Ozcelik H, Pinnaduwage D, et al (2004). The combination of p53 mutation and neu/erbB-2 amplification is associated with poor survival in node-negative breast cancer. J Clin Oncol. 22, 86-96.

- Chang HY, Nuyten DS, Sneddon JB, et al (2005). Robustness, scalability, and integration of a wound-response gene expression signature in predicting breast cancer survival. *Proc Natl Acad Sci USA*, **102**, 3738-43.
- Chang HY, Sneddon JB, Alizadeh AA, et al (2004). Gene expression signature of fibroblast serum response predicts human cancer progression: similarities between tumors and wounds. *PLoS Biol*, **2**, 206-14.
- Cho EY, Choi YL, Han JJ, Kim KM, Oh YL (2008). Expression and amplification of Her2, EGFR and cyclin D1 in breast cancer: immunohistochemistry and chromogenic in situ hybridization. *Pathol Int*, 58, 17-25.
- Dairkee SH, Luciani-Torres MG, Moore DH, Goodson WH (2013). Bisphenol-A-induced inactivation of the p53 axis underlying deregulation of proliferation kinetics, and cell death in non-malignant human breast epithelial cells. *Carcinogenesis*, **34**, 703-12.
- Deblois G, Giguère V (2013). Oestrogen-related receptors in breast cancer: control of cellular metabolism and beyond. *Nat Rev Cancer*, 13, 27-36.
- Ellis MJ, Tao Y, Young O, et al (2006). Estrogen-independent proliferation is present in estrogen-receptor HER2-positive primary breast cancer after neoadjuvant letrozole. J Clin Oncol. 24, 3019-25.
- Elston CW, Ellis IO (2002). Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term followup. *Histopathology*, **41**, 154-61.
- Erdem O, Dursun A, Coskun U, Gunel N (2005). The prognostic value of p53 and c-erbB-2 expression, proliferative activity and angiogenesis in node-negative breast carcinoma. *Tumori*, **91**, 46-52.
- Esteva-Lorenzo FJ, Sastry L, King CR (1998). The erbB-2 gene: from research to application. In: Dickson RB, Salomon DS., editor. In Hormones and Growth Factors in Development and Neoplasia. New York: John Wiley & Sons. 421-44.
- Fasching PA, Pharoah PD, Cox A, et al (2012). The role of genetic breast cancer susceptibility variants as prognostic factors. *Hum Mol Genet.* 21, 3926-39.
- Habashy HO, Powe DG, Glaab E, et al (2011). RERG (Ras-like, oestrogen-regulated, growth-inhibitor). expression in breast cancer: a marker of ER-positive luminal-like subtype. *Breast Cancer Res Treat*, **128**, 315-26.
- Harirchi I, Karbakhsh M, Montazeri A, et al (2010). Decreasing trend of tumor size and downstaging in breast cancer in Iran: results of a 15-year study. *Eur J Cancer Prev*, **19**, 126-30.
- Jehoram TA, Bency J, Sitara A, et al (2005). Relationship between the expression of various markers and prognostic factors in breast cancer. *Acta Histochimica*, **107**, 87-93.
- Kazkayasi M, Hücümenoğlu S, Siriner GI, Hücümenoğlu M (2001). Over-expression of p53 and c-erbB-2 oncoproteins in laryngeal carcinoma. *Eur Arch Otorhinolaryngol*, 258, 329-35.
- Khosravi A, Taylor R, Naghavi M, Lopez AD (2007). Differential mortality in Iran. *Popul Hlth Metr*, **28**, 5-7.
- Lagiou P, Georgila C, Samoli E, et al (2009). Estrogen alpha and progesterone receptor expression in the normal mammary epithelium in relation to breast cancer risk. *Int J Cancer*, **124**, 440-2.
- Lee S, Mohsin SK, Mao S, et al (2006). Hormones, receptors, and growth in hyperplastic enlarged lobular units: early potential precursors of breast cancer. *Breast Cancer Res*, **8**, 1-9.
- Ma XJ, Wang Z, Ryan P, et al (2004). A two gene expression ratio predicts clinical outcome in breast cancer patients treated with tamoxifen. *Cancer Cell*, **5**, 607-16.
- Matsuda T, Matsuda A (2013). Burden of cancer incidence below the age of 40 in Asia 2002 extrapolated from the

Cancer Incidence in Five Continents Vol. IX. Jpn J Clin Oncol, 43, 449-50.

- Mohagheghi MA, Mosavi-Jarrahi A (2010). Review of cancer registration and cancer data in Iran, a historical prospect. *Asian Pac J Cancer Prev*, **11**, 1155-7.
- Mousavi SM, Montazeri A, Mohagheghi MA, et al (2007). Breast cancer in Iran: an epidemiological review. *Breast* J, 13, 383-91.
- Müller V, Thomssen C, Karakas C, et al (2003). Quantitative assessment of HER-2/neu protein concentration in breast cancer by enzyme-linked immunosorbent assay. *Int J Biol Markers*, **18**, 13-20.
- Ouyang X, Gulliford T, Zhang H, et al (2001). Association of ErbB2 Ser1113 phosphorylation with epidermal growth factor receptor co-expression and poor prognosis in human breast cancer. *Mol Cell Biochem*, **218**, 47-54.
- Paik S, Shak S, Tang G, et al (2004). A multigene assay to predict recurrence of tamoxifen- treated node-negative breast cancer. *N Engl J Med*, **351**, 2817-26.
- Patsialou A, Wang Y, Lin J, et al (2012). Selective geneexpression profiling of migratory tumor cells in vivo predicts clinical outcome in breast cancer patients. *Breast Cancer Res*, 14, 139.
- Rody A, Holtrich U, Pusztai L, et al (2009). T-cell metagene predicts a favorable prognosis in estrogen receptor-negative and HER2-positive breast cancers. *Breast Cancer Res*, **11**, 15.
- Rosenthal SI, Depowski PL, Sheehan CE, Ross JS (2002). Comparison of HER-2/neu oncogene amplification detected by fluorescence in situ hybridization in lobular and ductal breast cancer. *Appl Immunohistochem Mol Morphol*, **10**, 40-6.
- Rudas M, Neumayer R, Gnant MFX, et al (1997). p53 protein expression, cell proliferation and steroid hormone receptors in ductal and lobular in situ carcinomas of the breast. *Eur J Cancer*, **33**, 39-44.
- Schonborn I, Zschieseche W, Spitzer E, et al (1994). C-erb-2 overexpression in primary breast cancer: independent prognostic factor in patients at high risk. *Breast Cancer Res Treat*, 29, 287-95.
- Skálová A, Sima R, Vanecek T, et al (2009). Acinic cell carcinoma with high-grade transformation: a report of 9 cases with immunohistochemical study and analysis of TP53 and HER-2/neu genes. Am J Surg Pathol, 33, 1137-45.
- Sorlie T, Tibshirani R, Parker J, et al (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proc Natl Acad Sci U S A* **100**, 8418-23.
- Sotiriou C, Neo SY, McShane LM, et al (2003). Breast cancer classification and prognosis based on gene expression profiles from a population-based study. *Proc Natl Acad Sci* USA, 100, 10393-8.
- Szoke J, Udvarhelyi N (2012). Modern pathologic diagnostics in breast cancer. *Orv Hetil*. **153**, 22-30.
- Thor AD, Berry DA, Budman DR, et al (1998). erbB-2, p53, and efficacy of adjuvant therapy in lymph node-positive breast cancer. *J Natl Cancer Inst*, **90**, 1346-60.
- Tian Y, Chen B, Guan P, Kang Y, Lu Z (2013). A Prognosis Classifier for Breast Cancer Based on Conserved Gene Regulation between Mammary Gland Development and Tumorigenesis: A Multiscale Statistical Model. *PLoS One*, 8, 60131.
- Tsuda H (2009). Gene and chromosomal alterations in sporadic breast cancer: correlation with histopathological features and implications for genesis and progression. *Breast Cancer*, 16, 186-201.
- Umekita Y, Ohi Y, Sagara Y, Yoshida H (2000). Co-expression of epidermal growth factor receptor and transforming growth

- Expressional Correlation of Receptor and p53 Markers in Breast CancerI Clinfactor-alpha predicts worse prognosis in breast-cancerpatients. Int J Cancer, 89, 484-7.
 - van de Vijver MJ, He YD, van 't Veer LJ, et al (2002). A geneexpression signature as a predictor of survival in breast cancer. *N Engl J Med*, **347**, 1999-2009.
 - Wang Y, Klijn JG, Zhang Y et al (2005). Gene expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. *Lancet*, **365**, 671-9.
 - Yang SX, Loo WT, Chow LW, et al (2012). Decreased expression of C-erbB-2 and CXCR4 in breast cancer after primary chemotherapy. *J Transl Med*, **10**, 1-3.
 - Yenidunya S, Bayrak R, Haltas H (2011). Predictive value of pathological and immunohistochemical parameters for axillary lymph node metastasis in breast carcinoma. *Diagn Pathol.* 13, 6-18.