

## RESEARCH ARTICLE

# Potential Therapeutic Efficacy of Curcumin in Liver Cancer

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### Abstract

**Purpose:** Liver cancer, one of the most common cancers in China, is reported to feature relatively high morbidity and mortality. Curcumin (Cum) is considered as a drug possessing anti-angiogenic, anti-inflammation and anti-oxidation effect. Previous research has demonstrated antitumor effects in a series of cancers. **Materials and Methods:** In this study the *in vitro* cytotoxicity of Cum was measured by MTT assay and pro-apoptotic effects were assessed by DAPI staining and measurement of caspase-3 activity. *In vivo* anti-hepatoma efficacy of Cum was assessed with HepG2 xenografts. **Results:** It is found that Cum dose-dependently inhibited cell growth in HepG2 cells with activation of apoptosis. Moreover, Cum delayed the growth of liver cancer in a dose-dependent manner in nude mice. **Conclusions:** Cum might be a promising phytochemistry in cancer therapy and further efforts are needed to explore this therapeutic strategy.

**Keywords:** Curcumin - apoptosis - liver cancer - *in vitro* - *in vivo*

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### Introduction

Liver cancer, one of the most common cancers in China, is reported with relatively high morbidity and mortality. Due to its resistance to chemotherapy, the response and survival of patients with liver cancer remains poor (Di et al., 2012; Kudo et al., 2012; Li et al., 2012; Ni et al., 2013). Recent studies focus on the potential of Traditional Chinese Medicine (TCM) in treating different kinds of cancers (Yin et al., 2011; Li et al., 2012).

Curcumin (Cum) is a phenolic pigment isolated from the root of Turmeric (*Curcuma Longa*) (Link et al., 2013; Yin et al., 2013). It is formerly considered as a drug possessing anti-angiogenic, anti-inflammation and anti-oxidation effect (Kwon et al., 2005; Suckow et al., 2006; EI-Azab et al., 2011; Wu et al., 2013). Previous researches in 2013 have demonstrated the antitumor effect of Cum in a series of cancers (Bayet-Robert et al., 2013; Du et al., 2013; Kumaravel et al., 2013; Masuelli et al., 2013; Ono et al., 2013; Sun et al., 2013; Wei et al., 2013; Yallapu et al., 2013; Zhang et al., 2013).

In the current study, we systematically evaluated the *in vitro* and *in vivo* anticancer efficiency of Cum in liver cancer. The cell inhibition effect of Cum was measured by MTT assay. Apoptosis of liver cancer cells was measured by DAPI and the activity of Caspase-3. The growth curve of tumor volume and bodyweight of the mice were measured every two days.

### Materials and Methods

#### Materials

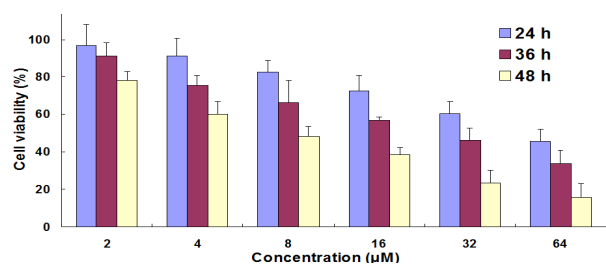
Curcumin, was purchased from Sigma Chem.Co., (St. Louis, USA). All other chemicals were of analytical grade and used without further purification. Human liver cancer cell line HepG2 was obtained from Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China).

Male and female nude mice (nu/nu; 6–8 weeks old and weighing 18–22 g) were purchased from Model Animal Research Center of Nanjing University (Nanjing, China). The mice were housed and maintained in the animal facility of the Animal Center of Nanjing Medical University. The animal protocol was reviewed and approved by the Institutional Animal Care and Use Committee of Nanjing Medical University.

#### *In vitro* cytotoxicity

The half maximal inhibitory concentrations (IC<sub>50</sub>s) of HepG2 cells were determined by the MTT assay. Briefly, cells were seeded in 96-well plates (1×10<sup>4</sup> cells per well) 24 h prior to the assay. Then cells were exposed to a series of doses of Cum. After 24, 36 and 48 hrs of incubation, 20 μl of 5 mg/mL MTT solution was added to each well and the plate was incubated for 4 h. Then, the media were removed and dimethylsulfoxide (DMSO) (150 μL) was added to each well. The optical density (OD) of each well

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**Figure 1. Effects of Cum on HepG2 Cell Proliferation.** (A) HepG2 cells were treated with Cum at 2, 4, 8, 16, 32 and 64 µM for 24, 36 and 48 hours

was measured using a microplate reader at 560 nm (Bio-Rad, Hercules, USA).

Cell viability was determined by following formula:

$$\text{Cell viability (\%)} = \text{OD (test well)} / \text{OD (reference well)} \times 100\% \quad (1)$$

All the results obtained from MTT assays were confirmed by repeating the experiment on at least three independent occasions and testing in triplicate each time. DAPI staining

The cells were treated with Cum at three doses (4, 8 and 16 µM) for 48 hrs, and then washed once in PBS followed by fixation in cold methanol : acetone (1:1) for 5 min. After washing thrice in PBS for 5 min, these cells were treated with 4 µg/ml DAPI for 10 min at routine temperature. The cells treated with the combination of the agents showed morphological changes of apoptosis, including a condensed and fragmented nuclear structure and decreased cell size (original magnification 200×).

#### Caspase-3 activity analysis

HepG2 cells were treated with a series of doses of Cum for 48h. Determination of caspase-3 activity was performed by the caspase colorimetric protease assay kit (Keygen Biotech, Nanjing, China) by following the manufacturer's instruction. The optical density was measured at 405 nm. The obtained values were expressed as folds of controls.

#### In vivo antitumor efficacy

Nude mice implanted with HepG2 cell line were used to qualify the antitumor efficacy of Cum through intravenous administration. The mice were raised under specific pathogen-free (SPF) circumstances and all of the animal experiments were performed in full compliance with guidelines approved by the Animal Care Committee of Nanjing Medical University. The mice were subcutaneously injected at the left axillary space with 0.1 ml of cell suspension containing 4–6×10<sup>6</sup> HepG2 cells. Treatments were started after 7–8 days of implantation. The mice whose tumor reached a tumor volume of 100 mm<sup>3</sup> were selected and this day was designated as "Day 0".

On Day 0, the mice were randomly divided into four groups, with each group being composed of 6 mice. The mice were treated intravenously with saline and a series of doses of Cum, respectively. Cum was administered at an equivalent dose of 20, 40, and 60 mg/kg. All mice were tagged, and tumors were measured every other day with calipers during the period of study. The tumor volume was calculated by the formula  $(W^2 \times L) / 2$ , where W is the

**Table 1. IC50s of Cum Against HepG2 Cells at Different Incubation Time**

IC50s (µM)	24 h	48 h	72 h
Cum	45.7±3.2	23.9±1.7	8.0±0.5

tumor measurement at the widest point, and L is the tumor dimension at the longest point.

Each animal was weighed at the time of treatment so that dosages could be adjusted to achieve the mg/kg amounts reported. Animals also were weighed every other day throughout the experiments. After 15 days of injections, the mice were sacrificed for the detection of peripheral blood parameters as well as liver and kidney functions.

#### Statistical analysis and research experience

Results were presented as Mean±SD. Statistical comparisons were made by t test or ANOVA analysis. The accepted level of significance was *P* value < 0.05.

We have enough experience in conducting medical researches, including clinical researches, and have published some results elsewhere (Huang et al., 2004; Zhou et al., 2009; Jiang et al., 2010; Yan et al., 2010; Gao et al., 2011; Huang et al., 2011; Li et al., 2011; Li et al., 2011; Li et al., 2011; Xu et al., 2011; Xu et al., 2011; Xu et al., 2011; Yan et al., 2011; Zhang et al., 2011; Gong et al., 2012; Gong et al., 2012; Gu et al., 2012; Li et al., 2012; Yu et al., 2012; Zhan et al., 2012; Zhan et al., 2012; Deng et al., 2013; Huang et al., 2013; Liu et al., 2013; Liu et al., 2013; Lu et al., 2013; Wu et al., 2013; Yin et al., 2013; Yin et al., 2013).

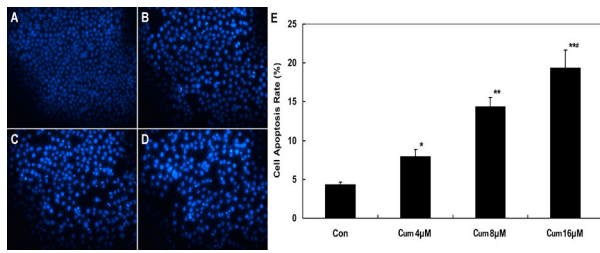
## Results and Discussion

#### In vitro cytotoxicity of Cum against HepG2 cells

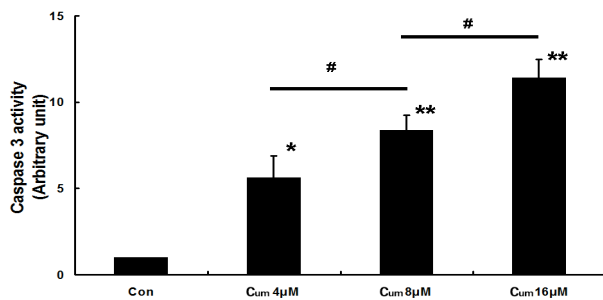
It is shown in Figure 1 that Cum inhibits the proliferation of HepG2 cells in a dose and time dependant manner. The cell viability decreases steadily when the dose of Cum increases as well as the incubation time prolongs. As indicated in Table 1, the IC50 value of Cum against HepG2 cells for 24h is 45.7±3.2 µM, while the IC50 of Cum for 36h is nearly a half (23.9±1.7 µM). Moreover, the longest incubation time of 48h produces the strongest inhibition effect of Cum with an IC50 value of 8.0±0.5 µM.

#### Apoptotic staining

It is indicated in Figure 2 that escalated doses of Cum leads to the increase of the apoptosis of HepG2 cells. As shown in Figure 2A–D, cells in control group have rounded and intact nuclei with diffused 4', 6-diamidino-2'-phenylindole dihydrochloride (DAPI) staining, while cells exposed to Cum have smaller and brighter stained nuclei with condensed chromatin forming crescent-shaped profiles around the periphery of the nucleus or separate globular structures (apoptotic bodies). Quantitative analysis demonstrates the dose-dependent pro-apoptotic effect in HepG2 cells (Figure 2E). Each dose of Cum led to significantly higher cellular apoptosis rate than that in control group (*p*<0.05). In detail, there was more



**Figure 2. Apoptosis of HepG2 Cells Detected by DAPI Staining.** (A) The non-treated cells. (B) Cells were treated with 4µM Cum. (C) Cells were treated with 8 µM Cum. (D) Cells were treated with 16 µM Cum. E: Quantitative analysis of apoptotic rate of cells exposed to different agents. Values represents Mean±SD (n = 3). \*means  $p < 0.05$  vs control group. \*\*means  $p < 0.01$  vs control group. #means  $p < 0.05$  vs 8 µM Cum



**Figure 3. Analysis of caspase-3 activity in Cells Exposed to a Series of Doses of Cum.** Values represents Mean ± SD. \* represents  $p < 0.05$  vs control, \*\*represents  $p < 0.01$  vs control, #represents  $p < 0.05$

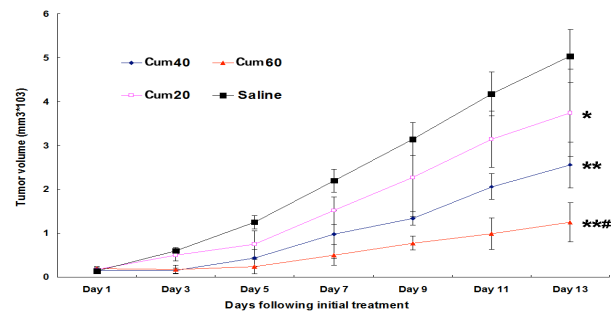
than 8% cells exhibiting apoptosis in cells exposed to the low dose of Cum (4 µM), while less than 5% cells underwent apoptosis in control group. Most importantly, there is significant increase in apoptosis rate among groups exposed to different doses of Cum ( $p < 0.05$ ).

#### Caspase-3 activation

It is reported that the Caspase family is crucial in apoptosis. Caspase-3, the key member of Caspase family, is a frequently activated death protease, catalyzing the specific cleavage of many key cellular proteins (Szymczyk et al., 2006). Therefore, caspase-3 is essential for certain processes associated with the dismantling of the cell and the formation of apoptotic bodies. In the current study, the same doses of Cum produced similar tendency as in DAPI staining (Figure 3). Increasing the dose of Cum caused the elevation of Caspase-3 activity with a remarkable difference, which was more significant than in control group. Results from DAPI staining and Caspase-3 activity demonstrate the potential of Cum in promoting the apoptosis of HepG2 cells.

#### In vivo antitumor evaluation of Cum in HepG2 xenograft mice

The therapeutic efficiency of Cum was evaluated in HepG2 human liver cancer xenografts in nude mice. As shown in Figure 4, Cum dose-dependently delayed tumor growth in HepG2 xenografts. All the three doses of Cum significantly inhibited the growth of liver cancer since Day 5 ( $p < 0.05$  vs control). Moreover, the group that received 60 mg/kg Cum was observed to maintain the greatest amount



**Figure 4. Antitumor Effect of Cum in HepG2 Xenograft Models.** (A) Tumor volume of established HepG2 xenografts in nude mice during therapy under different treatments. Mice were treated with different protocols on Day 0 as showed in the figure. Saline: vehicle; Cum was administered at the doses of 20, 40 and 60 mg/kg. Different agents were delivered through intravenous pathway when tumor volume measured 100 mm<sup>3</sup>. Data are presented as mean±SD (n = 6). The difference between tumor volumes in the group of saline and Cum is significant (\*means  $P < 0.05$ , \* means  $P < 0.01$ ). Significant difference (#means  $P < 0.05$ ) also is observed between the group receiving 60 mg/kg Cum and 40 mg/kg Cum

of anti-tumor activity among all the four groups (Figure 4). In detail, The tumor volumes of the group received low dose of Cum (20 mg/kg) is nearly 3740 mm<sup>3</sup> at the end of treatment, while that of the group received high dose of Cum (60 mg/kg) is around 1300 mm<sup>3</sup>, which is the lowest among all the groups indicating the strongest tumor inhibition. Statistical analysis reveals the significant differences between the group receiving Cum and control group. It is also noted that the high dose of Cum inhibited the growth of tumor more significantly than the other two doses of Cum.

In conclusion, the current study demonstrates the antitumor effect of Cum in the treatment of liver cancer. In vitro cytotoxicity evaluation indicates that Cum possesses a dose-dependent cell inhibition effect against HepG2 cells with the activation of Caspase-3. In vivo evaluation shows that Cum effectively inhibits the growth of liver cancer in a dose-dependent manner in nude mice. Therefore, Cum might be a promising phytomedicine in cancer therapy and further efforts are needed to explore this therapeutic strategy.

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#### References

Bayet-Robert M, Morvan D (2013). Metabolomics reveals metabolic targets and biphasic responses in breast cancer cells treated by curcumin alone and in association with docetaxel. *PLoS One*, **8**, e57971.



- Deng QQ, Huang XE, Ye LH, et al (2013). Phase II trial of Loubo® (Lobaplatin) and pemetrexed for patients with metastatic breast cancer not responding to anthracycline or taxanes. *Asian Pac J Cancer Prev*, **14**, 413-7.
- Di Carlo I, Toro A(2013). Future challenges for the treatment of liver tumors. *Future Oncol*, **9**, 481-3.
- Du Q, Hu B, An HM, et al (2013). Synergistic anticancer effects of curcumin and resveratrol in Hepa1-6 hepatocellular carcinoma cells. *Oncol Rep*, **29**, 1851-8.
- El-Azab M, Hishe H, Moustafa Y, et al (2011). Anti-angiogenic effect of resveratrol or curcumin in Ehrlich ascites carcinoma-bearing mice. *Eur J Pharmacol*, **10**, 7-14.
- Gao LL, Huang XE, Zhang Q, et al (2011). A Cisplatin and vinorelbine (NP) regimen as a postoperative adjuvant chemotherapy for completely resected breast cancers in China: final results of a phase II clinical trial. *Asian Pac J Cancer Prev*, **12**, 77-80.
- Gong P, Huang XE, Chen CY, et al (2012). Comparison on complications of peripherally inserted central catheters by ultrasound guide or conventional method in cancer patients. *Asian Pac J Cancer Prev*, **13**, 1873-5.
- Gu M, Li SY, Huang XE, et al (2013). A phase II study on continuous infusion paclitaxel and 5-Fu as first-line chemotherapy for patients with advanced esophageal cancer. *Asian Pac J Cancer Prev*, **13**, 5587-91.
- Huang XE, Li CG, Li Y, et al (2011). Weekly TP regimen as a postoperative adjuvant chemotherapy for completely resected breast cancer in China: final result of a phase II trial. *Asian Pac J Cancer Prev*, **12**, 2797-800.
- Huang XE, Wei GL, Huo JG, et al (2013). Intrapleural or intraperitoneal lobaplatin for treatment of patients with malignant pleural effusion or ascites. *Asian Pac J Cancer Prev*, **14**, 2611-4.
- Jiang Y, Huang XE, Yan PW, et al (2010). Validation of treatment efficacy of a computer-assisted program for breast cancer patients receiving postoperative adjuvant chemotherapy. *Asian Pac J Cancer Prev*, **11**, 1059-62.
- Kudo M (2012). Targeted therapy for liver cancer: updated review in 2012. *Curr Cancer Drug Targets*, **12**, 1062-72.
- Kumaravel M, Sankar P, Latha P, et al (2013). Antiproliferative effects of an analog of curcumin in Hep-2 cells: a comparative study with curcumin. *Nat Prod Commun*, **8**, 183-6.
- Kwon YK, Jun JM, Shin SW, et al (2005). Curcumin decreases cell proliferation rates through BTG2-mediated cyclin D1 down-regulation in U937 cells. *Int J Oncol*, **26**, 1597-603.
- Li CG, Huang XE, Li Y, et al (2011). Clinical observations on safety and efficacy of OxyContin® administered by rectal route in treating cancer related pain. *Asian Pac J Cancer Prev*, **12**, 2477-8.
- Li CG, Huang XE, Li Y, et al (2011). Phase II trial of irinotecan plus nedaplatin (INP) in treating patients with extensive stage small cell lung cancer. *Asian Pac J Cancer Prev*, **12**, 487-90.
- Li CG, Huang XE, Xu L, et al (2012). Clinical application of serum tumor associated material (TAM) from non-small cell lung cancer patients. *Asian Pac J Cancer Prev*, **13**, 301-4.
- Li X, Lu X, Xu H, et al (2012). Paclitaxel/tetrandrine coloaded nanoparticles effectively promote the apoptosis of gastric cancer cells based on "oxidation therapy". *Mol Pharm*, **9**, 222-9.
- Li X, Xu H, Dai X, et al (2012). Enhanced in vitro and in vivo therapeutic efficacy of codrug-loaded nanoparticles against liver cancer. *Int J Nanomedicine*, **7**, 5183-90.
- Li Y, Yan PW, Huang XE, et al (2011). MDR1 gene C3435T polymorphism is associated with clinical outcomes in gastric cancer patients treated with postoperative adjuvant chemotherapy. *Asian Pac J Cancer Prev*, **12**, 2405-9.
- Link A, Balaguer F, Shen Y, et al (2013). Curcumin modulates DNA methylation in colorectal cancer cells. *PLoS One*, **8**, e57709.
- Liu J, Huang XE, Tian GY, et al (2013). Phase II Study on Safety and Efficacy of Yadanzi® (Javanica oil emulsion injection) Combined with Chemotherapy for Patients with Gastric Cancer. *Asian Pac J Cancer Prev*, **14**, 2009-12.
- Liu W, Li SY, Huang XE, et al (2012). Inhibition of tumor growth in vitro by a combination of extracts from *rosa roxburghii* tratt and *fagopyrum cymosum*. *Asian Pac J Cancer Prev*, **13**, 2409-14.
- Liu YC, Zhou SB, Gao F, et al (2013). Chemotherapy and late course three dimensional conformal radiotherapy for treatment of patients with stage III non-small cell lung cancer. *Asian Pac J Cancer Prev*, **14**, 2663-5.
- Lu YY, Huang XE, Xu L, et al (2013). Potential predictors of sensitivity to pemetrexed as first-line chemotherapy for patients with advanced non-squamous NSCLCs. *Asian Pac J Cancer Prev*, **14**, 2005-8.
- Masulli L, Benvenuto M, Fantini M, et al (2013). Curcumin induces apoptosis in breast cancer cell lines and delays the growth of mammary tumors in neu transgenic mice. *J Biol Regul Homeost Agents*, **27**, 105-19.
- Ni CY, Yang Y, Chang YQ, et al (2013). Fast-track surgery improves postoperative recovery in patients undergoing partial hepatectomy for primary liver cancer: A prospective randomized controlled trial. *Eur J Surg Oncol*, **39**, 542-7.
- Ono M, Higuchi T, Takeshima M, et al (2013). Antiproliferative and apoptosis-inducing activity of curcumin against human gallbladder adenocarcinoma cells. *Anticancer Res*, **33**, 1861-6.
- Shu J, Li CG, Liu YC, et al (2012). Comparison of serum tumor associated material (TAM) with conventional biomarkers in cancer patients. *Asian Pac J Cancer Prev*, **13**, 2399-403.
- Suckow BK, Suckow MA (2006). Lifespan extension by the antioxidant curcumin in *Drosophila melanogaster*. *Int J Biomed Sci*, **2**, 402-5.
- Sun MQ, Meng AF, Huang XE, et al (2013). Comparison of psychological influence on breast cancer patients between breast-conserving surgery and modified radical mastectomy. *Asian Pac J Cancer Prev*, **14**, 149-52.
- Sun XD, Liu XE, Huang DS (2013). Curcumin reverses the epithelial-mesenchymal transition of pancreatic cancer cells by inhibiting the Hedgehog signaling pathway. *Oncol Rep*, **29**, 2401-7.
- Szymczyk KH, Freeman TA, Adams CS, et al (2006). Active caspase-3 is required for osteoclast differentiation. *J Cell Physiol*, **209**, 836-44.
- Wei X, Zhou D, Wang H, et al (2013). Effects of pyridine analogs of curcumin on growth, apoptosis and NF- $\kappa$ B activity in prostate cancer PC-3 cells. *Anticancer Res*, **33**, 1343-50.
- Wu B, Yao H, Wang S, et al (2013). DAPK1 modulates a curcumin-induced G2/M arrest and apoptosis by regulating STAT3, NF- $\kappa$ B, and caspase-3 activation. *Biochem Biophys Res Commun*, **434**, 75-80.
- Wu XY, Huang XE, You SX, et al (2013). Phase II study of pemetrexed as second or third line combined chemotherapy in patients with colorectal cancer. *Asian Pac J Cancer Prev*, **14**, 2019-22.
- Xu HX, Huang XE, Li Y, et al (2011). A clinical study on safety and efficacy of Aidi injection combined with chemotherapy. *Asian Pac J Cancer Prev*, **12**, 2233-6.
- Xu HX, Huang XE, Qian ZY, et al (2011). Clinical observation of Endostar® combined with chemotherapy in advanced colorectal cancer patients. *Asian Pac J Cancer Prev*, **12**, 3087-90.
- Xu JW, Li CG, Huang XE, et al (2011). Ubenimex capsule improves general performance and chemotherapy related

- toxicity in advanced gastric cancer cases. *Asian Pac J Cancer Prev*, **12**, 985-7.
- Xu T, Xu ZC, Zou Q, Yu B, Huang XE (2012). P53 Arg72Pro polymorphism and bladder cancer risk--meta-analysis evidence for a link in Asians but not Caucasians. *Asian Pac J Cancer Prev*, **13**, 2349-54.
- Xu X, Wang L, Xu HQ, et al (2013). Clinical comparison between Paclitaxel Liposome (Lipusu®) and Paclitaxel for treatment of patients with metastatic gastric cancer. *Asian Pac J Cancer Prev*, **14**, 2591-4.
- Yallapu MM, Ebeling MC, Jaggi M, et al (2013). Plasma proteins interaction with curcumin nanoparticles: implications in cancer therapeutics. *Curr Drug Metab*, **14**, 504-15.
- Yan PW, Huang XE, Yan F, et al (2011). Influence of MDR1 gene codon 3435 polymorphisms on outcome of platinum-based chemotherapy for advanced non small cell lung cancer. *Asian Pac J Cancer Prev*, **12**, 2291-4.
- Yin H, Guo R, Xu Y, et al (2012). Synergistic antitumor efficiency of docetaxel and curcumin against lung cancer. *Acta Biochim Biophys Sin*, **44**, 147-53.
- Yin HT, Tian QZ, Guan L (2013). In vitro and in vivo evaluation of the antitumor efficiency of resveratrol against lung cancer. *Asian Pac J Cancer Prev*, **14**, 1703-6.
- Yin HT, Zhang DG, Wu XL (2013). In vivo evaluation of curcumin-loaded nanoparticles in a A549 xenograft mice model. *Asian Pac J Cancer Prev*, **14**, 409-12.
- Yin HT, Zhang DG, Wu XL, et al (2013). In vivo evaluation of curcumin-loaded nanoparticles in a A549 xenograft mice model. *Asian Pac J Cancer Prev*, **14**, 409-12.
- Yu DS, Huang XE, Zhou JN, et al (2012). A Comparative study on the value of anal preserving surgery for aged people with low rectal carcinoma in Jiangsu, China. *Asian Pac J Cancer Prev*, **13**, 2339-40.
- Zhan YP, Huang XE, Cao J (2012). Clinical safety and efficacy of Kanglaite® (Coix Seed Oil) injection combined with chemotherapy in treating patients with gastric cancer. *Asian Pac J Cancer Prev*, **13**, 5319-21.
- Zhan YP, Huang XE, Cao J (2012). Clinical study on safety and efficacy of Qinin® (cantharidin sodium) injection combined with chemotherapy in treating patients with gastric cancer. *Asian Pac J Cancer Prev*, **13**, 4773-6.
- Zhang CY, Zhang L, Yu HX, et al (2013). Curcumin inhibits invasion and metastasis in K1 papillary thyroid cancer cells. *Food Chem*, **139**, 1021-8.
- Zhang LQ, Huang XE, Wang J (2011). The cyclin D1 G870A polymorphism and colorectal cancer susceptibility: a meta-analysis of 20 populations. *Asian Pac J Cancer Prev*, **12**, 81-5.
- Zhang XZ, Huang XE, Xu YL, et al (2012). Phase II study on voriconazole for treatment of Chinese patients with malignant hematological disorders and invasive aspergillosis. *Asian Pac J Cancer Prev*, **13**, 2415-8.
- Zhou JN, Huang XE, Ye Z, et al (2009). Weekly paclitaxel/Docetaxel combined with a platinum in the treatment of advanced non-small cell lung cancer: a study on efficacy, safety and pre-medication. *Asian Pac J Cancer Prev*, **10**, 1147-50.