RESEARCH ARTICLE

Interaction of XRCC1 and XPD Gene Polymorphisms with Lifestyle and Environmental Factors Regarding Susceptibility to Lung Cancer in a High Incidence Population in North East India

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Abstract

Background: This study aimed to explore the role of XRCC1 (Arg399Gln) and XPD (Lys751Gln) gene polymorphisms, lifestyle and environmental factors as well as their possible interactions in propensity to develop lung cancer in a population with high incidence from North East India. <u>Materials and Methods</u>: A total of 272 lung cancer cases and 544 controls were collected and XRCC1 (Arg399Gln) and XPD (Lys751Gln) genotypes were analyzed using a polymerase chain reaction based restriction fragment length polymorphism assay. Conditional multiple logistic regression analysis was used to calculate adjusted odds ratios and 95% confidence intervals after adjusting for confounding factors. <u>Results</u>: The combined Gln/Gln genotype of XRCC1 and XPD genes (OR=2.78, CI=1.05-7.38; p=0.040) was significantly associated with increased risk for lung cancer. Interaction of XRCC1Gln/Gln genotype with exposure of wood combustion (OR=2.56, CI=1.16-5.66; p=0.020), exposure of cooking oil fumes (OR=3.45, CI=1.39-8.58; p=0.008) and tobacco smoking (OR=2.54, CI=1.21-5.32; p=0.014) and interaction of XPD with betel quid chewing (OR=2.31, CI=1.23-4.32; p=0.009) and tobacco smoking (OR=2.13, CI=1.12-4.05; p=0.022) were found to be significantly associated with increased risk for lung cancer. <u>Conclusions</u>: Gln/Gln alleles of both XRCC1 and XPD genes appear to amplify the effects of household exposure, smoking and betel quid chewing on lung cancer risk in the study population.

Keywords: XRCC1 - XPD - environmental factors - interaction - lung cancer - high risk population - North-East India

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Introduction

Exposure to tobacco smoke, fumes and airborne particulates in the indoor environment and ionizing radiations are regarded as triggering factors for DNA damages (Sterpone and Cozzi, 2010; Tang et al., 2010). Converging lines of evidence suggest that cancer can be initiated by DNA damage, which if not repaired, can cause errors during DNA synthesis (Maynard et al., 2009). Humans are routinely exposed to mutagenic and carcinogenic aromatic amines via smoking, cooking of food and other sources (Zheng and Lee, 2009). DNA sodamaged is typically repaired by certain DNA-repairing enzymes. These enzymes are fundamental for the maintenance of genomic integrity in case of replication errors. Therefore individuals with impairment in DNA repair capability are often at an elevated risk of cancer development (Berwick and Vineis, 2000). In humans more than 100 proteins are involved in DNA repair system (Lopez-Cima et al., 2007). These proteins are implicated in various DNA repair pathways, including base excision repair (BER), nucleotide excision repair (NER) and mismatch repair (MMR) (Li et al., 2011). The X-ray repair cross-complementing group 1 (XRCC1) gene plays an important role in the development of lung cancer (Wang et al., 2014). XRCC1 protein plays a central role in base excision repair (BER) pathway by interacting with other DNA repair proteins (Yin et al., 2009). XRCC1 interacts with polynucleotide kinase enzyme, DNA pol- β , PARP1 and DNA ligase IIIa (Pramanik et al., 2011; Mutairi et al., 2013). Three coding polymorphisms in the XRCC1 gene are at codons 399 (Arg to Gln), 280 (Arg to His) and 194 (Arg to Trp) (Shen et al., 1998). In particular, 399 Gln/Gln allele is found to be significantly associated with higher level of DNA adducts, somatic mutations, sister chromatid exchanges and chromosomal damages (Lunn et al., 1999). Xerodermapigmentosum group D (XPD) encodes an evolutionary conserved ATP dependent helicase, a subunit of transcription factor II H (TFIIH) which is essential for transcription and NER (Coin et al., 1999; Li et al., 2013).

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Mutation of XPD codons 312 and 751 increases the risk of lung cancer (Zhou et al., 2012). XPD 751Gln/Gln has been demonstrated to have suboptimal DNA-repair capacity to remove UV photoproducts when compared to the XPD 751 Lys/Lys and Lys/Gln genotypes (Qiao et al., 2002).

Lung cancer (LC) is leading cause of cancer death worldwide with an annual mortality of 18.2 % cancer death (Ferlay et al., 2010a). India contributes 6.2% cases of LC with approximately 58,000 incidence cases reported in 2008 (Ferlay et al., 2010b). North Eastern (NE) parts of India represent a unique, strategic geographic location with a demographic diverse population. Manipur and Mizoram are two states from NE parts of India. LC is mostly predominant in NE parts of India, with highest ageadjusted rate (AAR) in Mizoram (28.3 per 10⁵ in male and 28.7 per 10⁵ in female). Manipur also contributes a very high incidence of LC (with AAR of 14.1 per 10⁵ in males and 11.9 per 10⁵ in females) (NCRP, 2013). These areas are also reported for a unique consumption of tobacco, betel quids and cooking habits that are different from other places (Phukan et al., 2001, 2005, 2006).

High risk of LC in the study population may be an outcome of genetic and environmental risk factors or a complex interaction of both. Studies have also reported association of XRCC1 and XPD allelic polymorphisms for LC (De-Ruyck et al., 2007; Li et al., 2013; Natukula et al., 2013). Lack of data on XRCC1 and XPD polymorphisms and high incidence of LC in NE parts of India prompted us to explore and evaluate any relevance of these

polymorphisms in the study population. We also wished to explore the interaction of XRCC1 and XPD gene with smoking, betel quid chewing, alcohol consumption, exposure of wood combustion during cooking and cooking oil fumes (COF).

Materials and Methods

Study design

Present study was an age (±5 years), sex and ethnicity matched population based case-control study. The study duration was from June 2010-May 2013. The work was carried out at Regional Medical Research Centre (RMRC) NE Region, Indian Council of Medical Research (ICMR); India in collaboration with Population Based Cancer Registry (PBCR), Imphal, Manipur and Aizawl, Mizoram, India. Incident cases and control subjects willing to participate in the study were indigenous people of Manipur and Mizoram. Histopathologically or cytologically confirmed LC cases with no evidence of pulmonary inflammation or benign lung tumors were included in the study. Cases too old to be interviewed elaborately and who refused to be interviewed were excluded from this study. Cancer free control subjects with age (± 5 years), sex and ethnicity matched were selected from healthy population of the states. None of the controls subjects had consanguinity with the cases or had any non-communicable diseases. Information of smoking, betel quid chewing, consumption of alcohol, exposure



Figure 1. RFLP Photograph of 2% Agarose Gel Electrophoresis for XRCC1 Genotype. Lane M represents 100 bp DNA Ladder. Lane 6 is characterised by 615bp that represents XRCC1 Gln/Gln genotype. Lane 1, 2, 3, 4, 5, 7, 8, 9, 10, 12, 13, 14 and 15 are characterised by 377bp and 238bp that represents XRCC1 Arg/Arg genotype. While, Lane 11 and 16 are characterised by 615bp, 377bp and 238bp that represents XRCC1 Arg/Gln genotype



Figure 2. RFLP Photograph of 2% Agarose gel electrophoresis for XPD Genotype. Lane M represents 50 bp DNA Ladder. Lane 1, 3, 4 and 5 are characterised by 734bp that represents XPD Lys/Lys genotype. Lane 2 and 6 are characterised by 734bp, 646bp and 88bp that represents XPD Lys/Gln genotype. While, Lane 7 is characterised by 646bp and 88bp that represents XPD Gln/Gln genotype

to household combustion and COF were recorded in a structured pre-designed questionnaire. The time period set for exposure of wood combustion during cooking and COF was 25 years, the participants were asked whether they have been exposed to the aforesaid time period or not. Those who were found to be exposed were taken as 'yes' for exposure of wood combustion and COF where as those who were not exposed for the set time period of 25 years were taken as 'no'. Written informed consent was taken from all subjects for participation in a protocol approved by the Institutional ethical committee of RMRC, NE Region (Indian Council of Medical Research). Thus a total of confirmed 272 LC cases and 544 controls matched for age (±5 years), sex and ethnicity were enrolled in the study.

DNA extraction

Four ml. of blood was collected from all subjects in EDTA vials. DNA was extracted by standard phenol chloroform method (Landi et al., 2006) and stored at -80° C till further analysis.

Genotyping of XRCC1 and XPD gene

Genotyping of XRCC1 and XPD were done by polymerase chain reaction based restriction fragment length polymorphism. All of the PCR reactions were

carried out by a Master cycler gradient thermo cycler (Bio-Rad, United States) in a final volume of 25 ul containing 200 ng of each primer (Sigma, United States), 50 ng genomic DNA, 1.0mM MgCl₂ (Roche, Germany), 200 ul dNTPs (Roche, Germany) and 2.0 unit of Taq DNA Polymerase (Roche). The PCR product was visualized in 2% agarose gel electrophoresis by gel documentation system (Cell Biosciences). XRCC1 PCR products were amplified with primers 5'-GCCCCGTCCCAGGTAAG-3' (sense) and 5'-AGCCCCAAGACCCTTTCACT-3'(antisense) (Park et al., 2002) followed by MspI (Promega) restriction digestion. The homozygous Gln allele was determined by presence of an uncut 615-bp band (indicative of absence of MspI cutting site), homozygous Arg allele was determined by presence of two bands at 377 and 238 bp while the heterozygous Arg/Gln allele was characterized by presence of three bands at 615, 377 and 238 bp (Figure 1). XPD PCR products were amplified with the primers 5'-CCTCTCCCTTTCCTCTGTTC-3' (sense) and 5'-CAGGTGAGGGGGGACATCT-3' (antisense) (Vettriselvi et al., 2007) and digested with PstI (New England BioLabs, Inc.). The homozygous Lys/Lys allele was characterized by an undigested band of 734bp, homozygous Gln/Gln allele determined by 646bp and 88bp, while heterozygous Lys/Gln genotype had three bands of 734bp, 646bp and 88bp (Figure 2). 10% of

Table 1.	Distribution	of Demographic	Characteristic and	l Risk of Lung Can	cer
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Category	Case n (%)	Control n (%)	Crude OR (95% CI)	p-value	Multivariate OR (95% CI)	p-value
Sample size	272 (100)	544 (100)				
Sex			0.529†			
Male	130 (47.8)	260 (47.8)				
Female	142 (52.2)	284 (52.2)				
Age (years)						
Range	21-88	21-89				
Means±SD	61.96±11.91	61.79±12.21		0.851‡		
Histological type						
Non-small squamous	160 (58.8)					
Non-small adenocarcinoma	65 (23.9)					
Small cell carcinoma	26 (9.6)					
Other §	21 (7.7)					
Exposure of wood combustion						
No	147 (54.0)	346 (63.6)	1.0 (Reference)		1.0 (Reference)	
Yes (Overall)	125 (46.0)	198 (36.4)	1.49 (1.11-1.20)	0.008	1.32 (0.96-1.81)	0.088
Yes (Males)	44 (16.2)	85 (15.6)	1.05 (0.67-1.65)	0.819	1.03 (0.64-1.63)	0.917
Yes (Females)	81 (29.8)	113 (20.8)	2.01 (1.34-3.02)	0.001	1.58 (1.01-2.48)	0.046*
Exposure of cooking oil fumes						
No	136 (50.0)	321 (59.0)	1.0 (Reference)		1.0 (Reference)	
Yes (Overall)	136 (50.0)	223 (41.0)	1.44 (1.07-1.93)	0.015	1.27 (0.93-1.75)	0.133
Yes (Males)	49 (18.0)	99 (18.2)	0.98 (0.64-1.52)	0.941	0.94 (0.60-1.48)	0.786
Yes (Females)	87 (32.0)	124 (22.8)	2.04 (1.35-3.08)	0.001	1.61 (1.02-2.53)	0.039*
Betel-quid chewing						
Nonchewer	85 (31.3)	208 (38.2)	1.0 (Reference)		1.0 (Reference)	
Chewer (Overall)	187 (68.7)	336 (61.8)	1.36 (1.00-1.86)	0.05	1.36 (0.99-1.87)	0.056
Chewer (Males)	83 (30.5)	158 (29.1)	1.14 (0.74-1.76)	0.556	1.09 (0.69-1.70)	0.718
Chewer (Females)	104 (38.2)	178 (32.7)	1.63 (1.05-2.54)	0.03	1.62 (1.02-2.56)	0.041*
Tobacco smoking						
Nonsmoker	75 (27.6)	203 (37.3)	1.0 (Reference)		1.0 (Reference)	
Smoker (Overall)	197 (72.4)	341 (62.7)	1.56 (1.14-2.15)	0.006	1.62 (1.17-2.24)	0.004*
Smoker (Males)	108 (39.7)	191 (35.1)	1.77 (1.04-3.03)	0.034	1.86 (1.07-3.23)	0.027*
Smoker (Females)	89 (32.7)	150 (27.6)	1.50 (0.99-2.27)	0.053	1.56 (1.02-2.39)	0.042*

*Significant; †Based on Chi-square test; ‡For independent samples T-test; § Other includes large cell carcinoma, bronchioalveolar carcinoma

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randomly selected samples were randomly sequenced to verify genotyping results and 100% concordance was found.

Statistical analysis

Difference in demographic characteristics, tobacco smoking, betel quid chewing and genotype frequencies between cases and controls were evaluated using Chi Square (χ^2) test. Estimates of LC risk, imparted by genotypes were determined by deriving odds ratio (OR) and its corresponding 95% confidence interval (95% CI) using multivariable conditional logistic regression after adjusting for potential confounders. For all tests, a two sided p≤0.05 was considered statistically significant. All statistical analysis were done using SPSS version 17.0.Tests for Hardy-Weinberg equilibrium amongst cases and control were conducted using observed genotype frequencies and a chi-square test featuring one degree of freedom.

Results

The details of demographic characteristics among cases and controls enrolled in this study are shown in Table 1. There was no statistically significant difference in term of mean age of cases (61.96 ± 11.91 years, range 21-88) and controls (61.79 ± 12.21 years, range 21-89) (p=0.711) of study subjects. 58.8% of cases were of non small squamous cell carcinoma, 23.9% were of non small adenocarcinoma and 9.6% were of small cell carcinoma and 7.7% others. Significant risk was observed for smoking (OR=1.62, CI=1.17-2.24; p=0.004). Risk was observed for betel-quid

chewing (OR=1.36, CI=0.99-1.87; p=0.056), exposure of wood combustion (OR=1.32, CI=0.96-1.81; p=0.088) and COF (OR=1.27, CI=0.93-1.75; p=0.133) but results are not statistically significant. Tests for Hardy-Weinberg equilibrium amongst cases and controls were conducted using observed genotype frequencies and a chi-square test featuring one degree of freedom. The distribution of genotypes for both XRCC1 and XPD genes among cases and controls were in Hardy-Weinberg equilibrium (Table 1).

Gln/Gln genotype was higher among cases than control groups in both XRCC1 (8.5% vs 6.3%) and XPD genes (12.9% vs 10.5%). Risk for LC was higher for individuals carrying Gln/Gln genotype in both XRCC1 (OR=1.37, CI=0.77-2.43; p=0.289) and XPD (OR=1.26, CI=0.78-2.04; p=0.352) genes, but results are not statistically significant. However a significant risk association was observed for combined effect Gln/Gln genotype of XRCC1 and XPD gene (OR=2.78, CI=1.05-7.38; p=0.040) after adjusting for potential confounders (Table 2).

Table 3. XRCC1 Arg399Gln and XPD Lys751Gln Allele Frequencies and Risk of Lung Cancer

Allele	Case n=544(%)	Control n=1088 (%)	Crude OR (95% CI)	p-value†
XRCC1 Arg39	9Gln		. ,	
Allele Arg	395 (72.6)	832 (76.5)	0.82 (0.65-1.03)	0.089
Allele Gln	149 (27.4)	256 (23.5)	1.23 (0.97-1.55)	0.089
XPD Lys751G	ln			
Allele Lys	358 (65.8)	749 (68.8)	0.87 (0.70-1.08)	0.216
Allele Gln	186 (34.2)	339 (31.2)	1.15 (0.92-1.43)	0.216

^{*}Based on Chi-square test

Table 2. Genetic Interaction and Distributions of XRCC1 Arg399Gln and XPD Lys751Gln Genotypes and Ris	k
of Lung Cancer	

Genotypes		Case	Control	Crude	p-value	Adjusted	p-value
		n(%)	n (%)	OR (95% CI)		OR (95% CI) †	
XRCC1 Arg3	99Gln						
Arg/Arg		146 (53.7)	322 (59.1)	1.0 (Reference)		1.0 (Reference)	
Arg/Gln		103 (37.8)	188 (34.6)	1.21 (0.89-1.65)	0.231	1.08 (0.78-1.48)	0.654
Gln/Gln		23 (8.5)	34 (6.3)	1.49 (0.85-2.62)	0.165	1.37 (0.77-2.43)	0.289
Hardy-Weinb	erg equilibrium	test‡					
χ2		0.63	0.86				
p-value		0.43	0.35				
XPD Lys7510	Əln						
Lys/Lys		121 (44.5)	262 (48.2)	1.0 (Reference)		1.0 (Reference)	
Lys/Gln		116 (42.6)	225 (41.3)	1.12 (0.82-1.52)	0.488	1.04 (0.76-1.43)	0.795
Gln/Gln		35 (12.9)	57 (10.5)	1.33 (0.83-2.13)	0.238	1.26 (0.78-2.04)	0.352
Hardy-Weinberg equilibrium test#		test‡					
χ2		0.75	0.7				
p-value		0.39	0.4				
XRCC1	XPD						
Arg/Arg	Lys/Lys	68 (25.0)	163 (30.0)	1.0 (Reference)		1.0 (Reference)	
Arg/Arg	Lys/Gln	67 (24.6)	129 (23.7)	1.25 (0.83-1.87)	0.294	1.19 (0.78-1.80)	0.416
Arg/Arg	Gln/Gln	11 (4.0)	30 (5.5)	0.88 (0.42-1.85)	0.735	0.86 (0.40-1.83)	0.691
Arg/Gln	Lys/Lys	52 (19.1)	84 (15.4)	1.48 (0.95-2.32)	0.083	1.37 (0.87-2.16)	0.179
Arg/Gln	Lys/Gln	38 (14.1)	85 (15.6)	1.07 (0.67-1.72)	0.776	0.87 (0.53-1.43)	0.583
Arg/Gln	Gln/Gln	13 (4.8)	19 (3.5)	1.64 (0.77-3.51)	0.202	1.47 (0.68-3.19)	0.334
Gln/Gln	Lys/Lys	1 (0.4)	15 (2.8)	0.16 (0.02-1.23)	0.079	0.14 (0.02-1.07)	0.058
Gln/Gln	Lys/Gln	11 (4.0)	11 (2.0)	2.40 (0.99-5.79)	0.052	2.29 (0.93-5.62)	0.07
Gln/Gln	Gln/Gln	11 (4.0)	8 (1.5)	3.30 (1.27-8.55)	0.014	2.78 (1.05-7.38)	0.040*

* Significant; † Adjusted OR were estimated by adjusting exposure of wood combustion, cooking oil fumes, betel-quid chewing and tobacco smokingin conditional multiple logistic regression model; ‡Hardy–Weinberg equilibrium test is calculated for 1 (one) degree of freedom and values rounded to two decimals

Mo	del Intera	ction Parameters	Case	Control	Crude	p-value	Adjusted	p-value			
			n (%)	n(%)	OR (95% CI)		OR (95% CI) #				
1†	XRCC1	Exposure of wood	combustion								
-	Arg/Arg	No	81 (29.8)	211 (38.7)	1.0 (Reference)		1.0 (Reference)				
	Arg/Arg	Yes	65 (23.9)	111 (20.4)	1.53 (1.02-2.27)	0.038	1.38 (0.91-2.10)	1.129			
	Arg/Gln	No	59 (21.7)	114 (21.0)	1.35 (0.90-2.02)	0.149	1.24 (0.82-1.87)	0.313			
	Arg/Gln	Yes	44 (16.2)	74 (13.6)	1.55 (0.99-2.44)	0.058	1.28 (0.80-2.05)	0.31			
	Gln/Gln	No	7 (2.6)	21 (3.9)	0.87 (0.36-2.12)	0.757	0.93 (0.38-2.30)	0.881			
	Gln/Gln	Yes	16 (5.9)	13 (2.4)	3.21 (1.48-6.96)	0.003	2.56 (1.16-5.66)	0.020*			
2‡	XRCC1	XRCC1 Exposure of cooking oil emission									
	Arg/Arg	No	71 (26.1)	195 (35.8)	1.0 (Reference)		1.0 (Reference)				
	Arg/Arg	Yes	75 (27.6)	127 (23.3)	1.62 (1.09-2.41)	0.016	1.44 (0.95-2.18)	0.082			
	Arg/Gln	No	56 (20.6)	101 (18.5)	1.52 (0.99-2.33)	0.052	1.38 (0.89-2.12)	0.148			
	Arg/Gln	Yes	47 (17.3)	87 (16.0)	1.48 (0.95-2.32)	0.084	1.22 (0.76-1.94)	0.413			
	Gln/Gln	No	9 (3.3)	25 (4.6)	0.99 (0.44-2.22)	0.978	0.91 (0.40-2.05)	0.811			
	Gln/Gln	Yes	14 (5.1)	9 (1.7)	4.27 (1.77-10.30)	0.001	3.45 (1.39-8.58)	0.008*			
3§	XRCC1	Betel-quid chewin	g								
	Arg/Arg	Nonchewer	56 (20.6)	124 (22.8)	1.0 (Reference)		1.0 (Reference)				
	Arg/Arg	Chewer	90 (33.1)	198 (36.4)	1.01 (0.67-1.51)	0.975	1.01 (0.67-1.51)	0.98			
	Arg/Gln	Nonchewer	26 (9.6)	72 (13.2)	0.80 (0.46-1.38)	0.424	0.72 (0.41-1.25)	0.241			
	Arg/Gln	Chewer	77 (28.3)	116 (21.3)	1.47 (0.96-2.25)	0.077	1.36 (0.88-2.10)	0.166			
	Gln/Gln	Nonchewer	3 (1.1)	12 (2.2)	0.55 (0.15-2.04)	0.374	0.56 (0.15-2.09)	0.388			
	Gln/Gln	Chewer	20 (7.4)	22 (4.1)	2.01 (1.02-3.99)	0.045	1.89 (0.94-3.79)	0.074			
4¶	XRCC1	Tobacco smoking									
	Arg/Arg	Nonsmoker	51 (18.8)	137 (25.2)	1.0 (Reference)		1.0 (Reference)				
	Arg/Arg	Smoker	95 (34.9)	185 (34.0)	1.38 (0.92-2.07)	0.12	1.46 (0.96-2.21)	0.075			
	Arg/Gln	Nonsmoker	19 (7.0)	50 (9.2)	1.02 (0.55-1.89)	0.948	1.01 (0.54-1.88)	0.983			
	Arg/Gln	Smoker	84 (30.9)	138 (25.4)	1.64 (1.07-2.49)	0.022	1.66 (1.08-2.54)	0.021*			
	Gln/Gln	Nonsmoker	5 (1.8)	16 (2.9)	0.84 (0.29-2.41)	0.745	0.86 (0.30-2.47)	0.773			
5.1.	Gln/Gln	Smoker	18 (6.6)	18 (3.3)	2.69 (1.30-5.56)	0.008	2.54 (1.21-5.32)	0.014*			
ЭŢ	XPD	Exposure of wood	combustion	17((20.4)	10 (D (10 (D (
	Lys/Lys	No	// (28.3)	1/6 (32.4)	1.0 (Kererence)		1.0 (Reference)				
	Lys/Lys	Ies No	44 (10.2) 55 (20.2)	128(25.2)	1.17(0.73-1.04)	0.497	1.03(0.04-1.00)	0.904			
	Lys/Gli	NO	55(20.2)	136 (23.3) 87 (16.0)	1.60(1.05, 2.45)	0.037	0.85(0.30-1.50) 1.26(0.87.2.12)	0.439			
	Clp/Clp	No	15(55)	37(10.0)	1.00(1.05-2.45) 1.07(0.55,2.00)	0.029	1.30(0.87-2.12) 1.00(0.55, 2.16)	0.174			
	Gln/Gln	No	13(3.3)	32(3.9)	1.07(0.33-2.09) 1.82(0.06.2.40)	0.04	1.09(0.33-2.10) 1.55(0.80,2.00)	0.197			
6+	XPD	Exposure of cooki	ng oil emission	25 (4.0)	1.05 (0.90-5.49)	0.007	1.55 (0.60-5.00)	0.195			
04	L ve/L ve	No	71(26.1)	152(27.9)	1.0 (Reference)		10 (Reference)				
	Lys/Lys Lys/Lys	Yes	50(184)	132(27.5) 110(20.2)	0.97 (0.63-1.51)	0.903	0.87 (0.55-1.38)	0 547			
	Lys/Gln	No	45 (16.5)	137(252)	0.70 (0.45-1.09)	0.116	0.66(0.42 - 1.02)	0.064			
	Lys/Gln	Yes	71 (26.1)	88 (16 2)	1 73 (1 13-2 63)	0.011	1 43 (0 91-2 23)	0.118			
	Gln/Gln	No	20 (7.4)	32 (5.9)	1.34 (0.72-2.50)	0.362	1.28 (0.68-2.43)	0.442			
	Gln/Gln	Yes	15 (5.5)	25 (4.6)	1.29 (0.64-2.59)	0.483	1.04 (0.50-2.15)	0.915			
78	XPD	Betel-quid chewin	g								
. 9	Lvs/Lvs	Nonchewer	46 (16.9)	107 (19.7)	1.0 (Reference)		1.0 (Reference)				
	Lvs/Lvs	Chewer	75 (27.6)	155 (28.6)	1.13 (0.72-1.75)	0.6	1.17 (0.75-1.83)	0.495			
	Lys/Gln	Nonchewer	34 (12.5)	73 (13.4)	1.08 (0.64-1.85)	0.769	1.06 (0.62-1.82)	0.836			
	Lys/Gln	Chewer	82 (30.1)	152 (27.9)	1.26 (0.81-1.94)	0.309	1.20 (0.77-1.87)	0.424			
	Gln/Gln	Nonchewer	5 (1.8)	28 (5.1)	0.42 (0.15-1.14)	0.089	0.41 (0.15-1.14)	0.087			
	Gln/Gln	Chewer	30 (11.0)	29 (5.3)	2.41 (1.30-4.46)	0.005	2.31 (1.23-4.32)	0.009*			
8¶	XPD	Tobacco smoking			. ,						
	Lys/Lys	Nonsmoker	35 (12.9)	105 (19.3)	1.0 (Reference)		1.0 (Reference)				
	Lys/Lys	Smoker	86 (31.6)	157 (28.9)	1.64 (1.03-2.61)	0.036	1.76 (1.09-2.82)	0.020*			
	Lys/Gln	Nonsmoker	31 (11.4)	77 (14.2)	1.21 (0.69-2.13)	0.513	1.16 (0.65-2.05)	0.614			
	Lys/Gln	Smoker	85 (31.3)	148 (27.1)	1.72 (1.08-2.75)	0.022	1.70 (1.06-2.73)	0.028*			
	Gln/Gln	Nonsmoker	9 (3.3)	21 (3.9)	1.29 (0.54-3.07)	0.571	1.32 (0.55-3.18)	0.537			
	Gln/Gln	Smoker	26 (9.6)	36 (6.6)	2.17 (1.15-4.08)	0.017	2.13 (1.12-4.05)	0.022*			

Table 4. Interaction of XRCC1 Arg399Gln and XPD Lys751Gln Genotypes with Exposure of Wood Combustion, Cooking Oil Fumes, Betel-Quid Chewing and Tobacco Smoking

*Significant; †Exposure of cooking oil fumes, betel-quid chewing and tobacco smokingwere adjusted to estimate adjusted OR in each model; ‡Exposure of wood combustion, betel-quid chewingand tobacco smokingwere adjusted to estimate adjusted OR in each model; \$Exposure of wood combustion, exposure of cooking oil fumes and betel-quid chewing and tobacco smokingwere adjusted OR in each model; \$Exposure of wood combustion, exposure of cooking oil fumes and betel-quid chewing were adjusted to estimate adjusted OR were estimated through conditional multiple logistic regression model

Allele frequency Gln was also higher in cases than control groups in both XRCC1 (27.4% vs 23.5%) and XPD (34.2 vs 31.2%) genes. Risk was observed for Gln Allele in both XRCC1 (OR=1.23, CI=0.97-1.55; p=0.089) and XPD (OR=1.15, CI=0.92-1.43; p=0.216) genotype; however result are not statistically significant (Table 3).

Interaction of XRCC1Gln/Gln genotype with exposure

exposure of COF (OR=3.45, CI=1.39-8.58; p=0.008) and tobacco smoking (OR=2.54, CI=1.21-5.32; p=0.014) were also significantly associated with increased risk of LC. Similarly interaction of XPD with betel quid chewing (OR=2.31, CI=1.23-4.32; p=0.009) and tobacco smoking (OR=2.13, CI=1.12-4.05; p=0.022) were also significantly associated with increased risk for LC after adjusting for potential confounders (Table 4).

Discussion

In this study, we examined whether association of XRCC1 and XPD genes polymorphisms and their interaction with indoor household exposure during cooking, tobacco smoking and betel quid chewing are implicated in development of LC in population with high incidence of lung cancer from North East India. Observation on association of XRCC1 and XPD on LC was inconsistent in different ethnic and geographical region with varying allele frequency (Lopez-Cima et al., 2007; Improta et al., 2008; Karkucak et al., 2012). In present study, no significant independent association of XRCC1 and XPD polymorphisms for LC was observed. These findings are concordant with some of the previous reports over different ethnic population (David-Beabes et al., 2001; Huang et al., 2008; Sun et al., 2013). However present study reveals significant association when XRCC1 Gln/Gln genotype interact with exposure of wood combustion, exposure of COF and tobacco smoking, while XPD Gln/Gln genotype with betel quid chewing and tobacco smoking. Combined effect of XRCC1 (Arg399Gln) and XPD (Lys751Gln) were also analysed. Result suggested that individuals with both XRCC1 Gln/Gln and XPD Gln/Gln genotype seemed to have synergistically increased risk for LC compared with those of either of them. Environmental exposure primarily tobacco smoke and other household exposure contain complex mixture of certain substances, it is plausible that repair of DNA damage intrigued by these mixed substances either by BER pathway or NER pathway. The failure or diminished on either side may cause LC risk. As expected, our study also confirmed the well established association between tobacco smoking and LC. Because of the traditional culture of study population, responsibility of cooking lies mostly with women; they are more exposed to COF and other household exposure. Interestingly in the present study significantly higher risk was observed in women than in males for LC in terms of exposure of wood combustion (OR=1.58, CI=1.01-2.48; p=0.046) and COF (OR=1.61, CI=1.02-2.53; p=0.039). Study conducted in Shenyang also observed a positive association between COF and LC risk among women (Li et al., 2008). Another study conducted by Hung et al. (2007) reported that COF is capable of causing cellular destruction of genetic material.

Wood combustion, cooking oil emission, tobacco, betel quids primarily constitute large number of polycyclic aromatic hydrocarbon (PAH), alkaloids and other phenolic compounds which are considered as a prime risk factors of LC (Seow et al., 2000; Pfeifer et al., 2002; Li et al., 2008; Hosgood et al., 2010; Mandal et al., 2013). Individuals differ widely in their capacity to repair DNA damage from both exogenous agents, such as wood and tobacco smoke, exposure to COF as well as endogenous reactions. Present study reports for XRCC1 and XPD gene polymorphisms, their interaction with exposure of COF, wood combustion, betel quid chewing, tobacco smoking and alcohol consumption and its association with LC in a high risk area from NE parts of India. Though no significant association for XRCC1 and XPD genotype on LC was observed, the Gln/Gln allele

of XRCC1 seems to contribute significant risk modifiers for exposure of wood combustion, exposure of COF and tobacco smoking while the Gln/Gln allele of XPD with betel quid chewing and tobacco smoking. Studies conducted by Lunn et al. (1999) reveals that Gln allele is associated with higher DNA adduct level or lower DNA repair efficiency. PAH-induced bulky DNA adducts, such as benzo[a]pyrenediol epoxide-DNA adducts, which are the most potent premutagenic adducts are mainly repaired by NER. A variety of reactive oxygen species, such as hydroxyl radical and hydrogen peroxide are generated during enzymatic oxidation of PAH (Park et al., 2002). These reactive oxygen species can lead to DNA damages which may be quantitatively a predominant PAH-induced DNA damage. Oxidative DNA damages are primarily removed via BER, including XRCC1.

Our study has several strengths and findings. It was a population based case-control study with a high participation rates. Our cases were incident; therefore possibility of observer or recall bias can be nullified. Also case-control matching was done in reference to age (± 5 years), sex and ethnicity, thereby controlling for any confounding effect on account of these variables.

Present study indicates that there is no significant relationship between XRCC1, XPD polymorphisms in study population. Significant risk was observed for interaction of these genes with some environmental factors. However a validation of these results will require its replication in a larger sample size. Taking into account other factors such as susceptibility differences of familial aggregation studies, epigenetic mechanism and infection (Human papilloma virus) will gives us more probable factors for increase risk of LC in NE parts of India.

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