RESEARCH ARTICLE

The -765G>C Polymorphism in the Cyclooxygenase-2 Gene and Digestive System Cancer: a Meta-analysis

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Abstract

Background: Published data regarding associations between the -765G>C polymorphism in cyclooxygenase-2 (COX-2) gene and digestive system cancer risk have been inconclusive. The aim of this study was to comprehensively evaluate the genetic risk of the -765G>C polymorphism in the COX-2 gene for digestive system cancer. Materials and Methods: A search was performed in Pubmed, Medline (Ovid), Embase, CNKI, Weipu, Wanfang and CBM databases, covering all studies until Feb 10, 2014. Statistical analysis was performed using Revman 5.2. Results: A total of 10,814 cases and 16,174 controls in 38 case-control studies were included in this meta-analysis. The results indicated that C allele carriers (GC+CC) had a 20% increased risk of digestive system cancer when compared with the homozygote GG (odds ratio (OR)=1.20, 95% confidence interval (CI), 1.00-1.44 for GC+CC vs GG). In the subgroup analysis by ethnicity, significant elevated risks were associated with C allele carriers (GC+CC) in Asians (OR = 1.46, 95% CI=1.07-2.01, and <math>p=0.02) and Africans (OR = 2.12, 95% CI=1.57-2.87, and p=0.02)p<0.00001), but not among Caucasians, Americans and mixed groups. For subgroup analysis by cancer type (GC+CC vs GG), significant associations were found between the -765G>C polymorphism and higher risk for gastric cancer (OR=1.64, 95% CI=1.03-2.61, and p=0.04), but not for colorectal cancer, oral cancer, esophageal cancer, and others. Regarding study design (GC+CC vs GG), no significant associations were found in then population-based case-control (PCC), hospital-based case-control (HCC) and family-based case-control (FCC) studies. Conclusions: This meta-analysis suggested that the -765G>C polymorphism of the COX-2 gene is a potential risk factor for digestive system cancer in Asians and Africans and gastric cancer overall.

Keywords: Cyclooxygenase-2 - digestive system cancer - meta-analysis - polymorphism

Asian Pac J Cancer Prev, 15 (19), 8301-8310

Introduction

Digestive system cancer is a sophisticated disease that results from complex interactions between environmental factors, genetic variants, and gene-environment interactions (Berlau et al., 2004; Pharoah et al., 2004; Yaghoobi et al., 2004). Helicobacter pylori (H. pylori) infection, environmental tobacco smokes and lifestyle represent the most important exogenous risk factors (Cook et al., 2010). Although these factors have been documented to influence the risk of cancer, not all individuals develop the disease, even though they are exposed in the same environment. This indicates that genetic differences, such as variants, may contribute to cancer pathogenesis. Therefore, numerous published studies have focused on the association of genetic variants with cancer susceptibility. And among them, cyclooxygenase (COX) gene has been extensively studied.

The COX gene, mapped to chromosome 1q25.2-q25.3 in human, is 8.3 kb in size, contains 10 exons and produces

an mRNA of 4.6 kb, which encodes a constitutive isoenzyme (COX-1) and an inducible isoenzyme (COX-2) (Tazawa et al., 1994). COX-1 is constitutively expressed and is involved in the homeostasis of various physiological functions (Dubois et al., 1998), while COX-2, known as rate-limiting enzyme produced during the production of prostaglandins, is often undetectable in normal tissue, whereas in tumor tissue specimens its expression is observably higher (Harrison et al., 1994; Seibert et al., 1994; Wu et al., 1996; Bakhle et al., 2001; Cao et al., 2002; Wang et al., 2007). This gene is polymorphic, and a large number of single nucleotide polymorphisms (SNPs) have been identified, such as -765G>C (reference SNP ID, rs20417), -1195G>A (rs689466), -8473T>C (rs5275), -1759G>A (rs3218625), -202C>T (rs2745557), and -1290A>G (rs689466). Among all of these polymorphisms, the -765G>C polymorphisms in COX-2 gene were the most widely studied for their implication in cancer risk. Several meta-analyses investigating this -765G>C polymorphism of COX-2

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gene with digestive system cancer risks were performed; however, the results were inconsistent for different ethnicity and cancer types (Dong et al., 2010; Liu et al., 2010; Wang et al., 2013; Yan et al., 2013). Therefore, we conducted this meta-analysis on all eligible case-control studies to estimate the overall digestive system cancer risk associated with this polymorphism.

Materials and Methods

Selection of studies

A systematic literature search of the Pubmed, Medline (Ovid), Embase, CNKI, Weipu database, Wanfang database and CBM database was carried out to identify studies involving association between the -765G>C polymorphism of COX-2 gene and digestive system cancer risk (updated on Feb 10, 2014). The search terms were used as follows: (cyclooxygenase-2 or COX-2 or PTGS2) in combination with (polymorphism or variant or mutation) and (cancer or carcinoma or neoplasm). The search results were limited to English and Chinese languages. Studies included in our meta-analysis met the following inclusion criteria: (1) evaluation of the -765G>C polymorphism of COX-2 gene and digestive system cancer risk, (2) the design had to be a case-control design published in a journal, (3) genotype distributions in both cases and controls were available for estimating an odds ratio with 95% confidence interval (CI) and P value, and (4) genotype distributions in control group should be consistent with Hardy-Weinberg equilibrium (HWE). Studies were excluded if one of the following existed: (1) no controls, (2) genotype frequencies or number not reported, and (3) abstracts, reviews. For duplication or overlapping publications, the studies with larger number of cases and controls or been published latest were included.

Data extraction

Two independent reviewers (FZ and CY) collected the data and reached a consensus on all items. In case of disagreement, a third author (HZ) would assess these articles. A standardized data form was used and included: first author's name, year of publication, original country, ethnicity, cancer type, study design, total number of cases and controls, genotyping method, and genotype distribution in cases and controls.

Statistical analysis

Odds ratios (ORs) with 95% CI was used to assess the strength of association between the COX-2-765G>C polymorphism and digestive system cancer risk. We first estimated with the risk of genetic model (CC+GC vs GG), and then estimated the risk of (C vs G) model. The pooled OR was calculated by a fixed-effects model or a random-effects model according to the heterogeneity. Heterogeneity was checked by a χ^2 -based Q statistic and P <0.10 was considered statistically significant. If the result was p>0.10, OR was pooled according to the fixed-effect model; otherwise, the random effect model was used. The statistical significance of OR was analyzed by Z test, and p<0.05 was considered as statistically significant. To evaluate the ethnicity-specific, cancer type-specific,

study design-specific effects, we performed stratification analyses on ethnicity, cancer type, and study design. For the subgroup analysis by ethnicity, the study populations were stratified into four groups: Asians, Caucasians, Americans, Africans, Mixed (if it was difficult to discriminate the ethnicity of participants according to the data presented, the study was termed "Mixed"). Subgroup analysis by cancer type were preformed if one cancer type contained three and more than three individual studies (if 1 cancer type was investigated <3 individual case-control studies, then it was combined into the group of "others"). In addition, subjects were categorized into different classifications according to study design: population-based case-control study (PCC), hospital-based case-control study (FCC).

Sensitivity analysis was also performed by sequence excluding individual study to check the robustness of the result (Zhang et al., 2010). The possible publication bias was examined visually in a Begg's funnel plot and the degree of asymmetry was tested by Egger's test (Begg et al., 1994; Egger et al., 1997). HWE was tested by Pearson's χ^2 test (Zhang et al., 2010). Statistical analysis was performed using Reman5.2 software (The Cochrane Collaboration, www.cochrane.org).

Results

Study inclusion and characteristics

The initial search identified 194 studies from the selected electronic databases (Figure 1). After reading the titles and abstracts, 95 potential articles were included for full-text view. After reading the full texts, 50 studies were excluded for being irrelevant to digestive system cancer risk and *COX-2* gene. Therefore, 45 full-text articles remained for data extraction. 1 articles were excluded for not present usable data (Kamal et al., 2012), An additional 3 articles were excluded for repeat or overlapping studies (Zhang et al., 2005; Zhang et al., 2006; Zhang et al., 2011). Therefore, a total of 41 case-control studies published in 40 articles were identified. However, the control group genotypes in 4 case-control studies were not consistent with HWE (KX et al., 2008; Akkiz et al., 2011; Talar-Wojnarowska et al., 2011; Kamal et al., 2012), and these

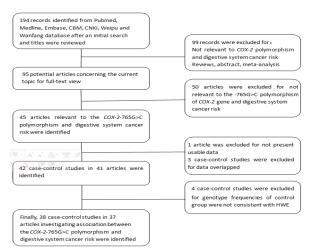


Figure 1. Flow Diagram of Included/Excluded Studies

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studies were excluded. Thus, a final total of 38 case-control studies in 37 articles which met our inclusion criteria were identified, including 10814 cases and 16174 controls. Among 38 cases and controls, there were 20 case-controls of Asians (Hamajima et al., 2001; Koh et al., 2004; Liu et al., 2006; Guo et al., 2007; Tan et al., 2007; Chiang et al., 2008; Lin et al., 2008; Saxena et al., 2008; Ueda et al., 2008; Xing et al., 2008; Xu et al., 2008; Tang et al., 2009; Upadhyay et al., 2009; Zhao et al., 2009; Mittal et al., 2010; He et al., 2011; Zhang et al., 2011; Chang et al., 2012; Li et al., 2012; Shin et al., 2012), 11 of Caucasians (Cox et al., 2004; Pereira et al., 2006; Hou et al., 2007; Moons et al., 2007; Sitarz et al., 2008; Andersen et al., 2009; Hoff et al., 2009; Iglesias et al., 2009; Kristinsson et al., 2009; Pereira et al., 2010; Daraei et al., 2012), 4 of Americans (Ulrich et al., 2005; Gunter et al., 2006; Gong et al., 2009; Thompson et al., 2009), and 1 of Africans (Bye et al., 2011), and 2 of Mixed (Bye et al., 2011; Wang et al., 2012). 9 investigated gastric cancer (Liu et al., 2006; Pereira et al., 2006; Hou et al., 2007; Saxena et al., 2008; Sitarz et al., 2008; Tang et al., 2009; Zhang et al., 2011; Li

et al., 2012; Shin et al., 2012), 16 investigated colorectal cancer (Hamajima et al., 2001; Cox et al., 2004; Koh et al., 2004; Ulrich et al., 2005; Gunter et al., 2006; Tan et al., 2007; Xing et al., 2008; Ueda et al., 2008; Andersen et al., 2009; Gong et al., 2009; Hoff et al., 2009; Iglesias et al., 2009; Thompson et al., 2009; Pereira et al., 2010; Daraei et al., 2012; Wang et al., 2012), 3 investigated oral cancer (Chiang et al., 2008; Lin et al., 2008; Mittal et al., 2010), 6 investigated esophageal cancer (Guo et al., 2007; Moons et al., 2007; Kristinsson et al., 2009; Upadhyay et al., 2009; Bye et al., 2011), and 4 investigated pancreatic cancer and hepatocellular carcinoma (HCC) (Xu et al., 2008; Zhao et al., 2009; He et al., 2011; Chang et al., 2012). Of these articles, 18 studies were performed in HB (Hamajima et al., 2001; Cox et al., 2004; Gunter et al., 2006; Pereira et al., 2006; Guo et al., 2007; Chiang et al., 2008; Lin et al., 2008; Saxena et al., 2008; Xing et al., 2008; Xu et al., 2008; Hoff et al., 2009; Iglesias et al., 2009; Upadhyay et al., 2009; Mittal et al., 2010; Pereira et al., 2010; He et al., 2011; Chang et al., 2012; Shin et al., 2012), 19 studies were performed in PB (Ulrich et

Table 1. Characteristics of the Studies Included in Meta-analysis

First author	Year	Country	Ethnicity	Cancer type	Study design	No. (Cases/Controls)	Genotyping method
Pereira et al	2006	Portugal	Caucasians	Gastric	HCC	73/ 210	PCR-RFLP
Liu et al	2006	China	Asians	Gastric	PCC	247/427	DHPLC
Hou et al	2007	Poland	Caucasians	Gastric	PCC	290/409	TaqMan
Saxena et al	2008	India	Asians	Gastric	HCC	62/241	PCR-RFLP
Sitarz et al	2008	Netherlands	Caucasians	Gastric	PCC	241/100	PCR-sequence
Tang et al	2009	China	Asians	Gastric	PCC	100/105	PCR-RFLP
Zhang et al	2011	China	Asians	Gastric	PCC	323/944	PCR-RFLP
Li et al	2012	China	Asians	Gastric	PCC	296/319	PCR-RFLP
Shin et al	2012	Korea	Asians	Gastric	HCC	100/100	PCR-RFLP
Hamajima et al	2001	Japan	Asians	Colorectal	HCC	148/241	PCR-CTPP
Cox et al	2004	Spain	Caucasians	Colorectal	HCC	220/257	TaqMan
Koh et al	2004	Singapore	Asians	Colorectal	PCC	310/1177	TaqMan
Ulrich et al	2005	America	Americans	Colorectal	PCC	494/584	PCR-RFLP
Gunter et al	2006	America	Americans	Colorectal	HCC	210/196	PCR-RFLP
Tan et al	2007	China	Asians	Colorectal	PCC	1000/1300	PCR-RFLP
Xing et al	2008	China	Asians	Colorectal	HCC	137/199	PCR-RFLP
Ueda et al	2008	Japan	Asians	Colorectal	PCC	455/1051	PCR-RFLP
Gong et al	2009	America	Americans	Colorectal	PCC	162/211	PCR-RFLP
Iglesias et al	2009	Spain	Caucasians	Colorectal	HCC	284/123	PCR-RFLP
Thompson et al	2009	America	Americans	Colorectal	PCC	421/479	TaqMan
Andersen et al	2009	Denmark	Caucasians	Colorectal	PCC	359/765	TaqMan
Hoff et al	2009	Netherlands	Caucasians	Colorectal	HCC	326/369	PCR-RFLP
Pereira et al	2010	Portugal	Caucasians	Colorectal	HCC	117/256	PCR-RFLP
Daraei et al	2012	Iran	Caucasians	Colorectal	PCC	110/120	PCR-RFLP
Wang et al	2012	Multicenter	Mixed	Colorectal	FCC	305/359	PCR-RFLP
Chiang et al	2008	China	Asians	Oral	HCC	178/205	PCR-RFLP
Lin et al	2008	China	Asians	Oral	HCC	297/280	PCR-RFLP
Mittal et al	2010	India	Asians	Oral	HCC	176/96	PCR-RFLP
Guo et al	2007	China	Asians	Esophageal	HCC	1026/1270	PCR-RFLP
Moons et al	2007	Netherlands	Caucasians	Esophageal	PCC	140/495	PCR-RFLP
Upadhyaya et al	2009	India	Asians	Esophageal	HCC	174/216	PCR-RFLP
Kristinsson et al	2009	Netherlands	Caucasians	Esophageal	PCC	222/236	PCR-RFLP
Bye et al (A)	2011	South Africa	Africans	Esophageal	PCC	347/462	TaqMan
Bye et al (M)	2011	South Africa	Mixed	Esophageal	PCC	190/422	TaqMan
Xu et al	2008	China	Asians	Pancreatic	HCC	283/566	PCR-RFLP
Zhao et al	2009	China	Asians	Pancreatic	PCC	393/786	PCR-RFLP
He et al	2011	China	Asians	HCC*	HCC	300/300	PCR-RFLP
Chang et al	2012	China	Asians	HCC*	HCC	298/298	PCR-RFLP

*HCC, hepatocellular carcinoma; population-based case-control study (PCC), hospital-based case-control study (HCC), and family-based case-control study (FCC)

al., 2005; Liu et al., 2006; Hou et al., 2007; Moons et al., 2007; Tan et al., 2007; Sitarz et al., 2008; Ueda et al., 2008; Andersen et al., 2009; Gong et al., 2009; Kristinsson et al., 2009; Tang et al., 2009; Thompson et al., 2009; Zhao et al., 2009; Bye et al., 2011; Zhang et al., 2011; Daraei et al., 2012; Li et al., 2012), 1 in FB (Wang et al., 2012). Different genotyping methods were used, including TaqMan, Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP), DHPLC (PCR-based denaturing high-performance liquid chromatography). The characteristics of each case-control study are listed in Table 1. Genotype and allele distributions for each case-control study are shown in Table 2.

Quantitative data synthesis

All studies: As shown in Figure 2, the heterogeneity of (CC+ GC vs GG) for all 38 studies was assessed and the value of χ^2 was 259.68 with 37 degrees of freedom and p<0.00001 in a random-effects model. Additionally, the I-square, which is another index of the test of heterogeneity,

was 86%, suggesting a moderate heterogeneity. Thus, we chose the random-effects model to synthesize the data. Overall, OR was 1.20 (95% CI=1.00-1.44), and the test for overall effect Z value was 2.01 (p=0.04) for (CC+GC vs GG) genetic model. The results suggested the C allele carriers (CC+GC) may have a 20% increased risk compared with the homozygote GG. Summary results for all comparisons are presented in Table 3.

Subgroup analyses: In the subgroup analysis by ethnicity (GC+CC vs GG, Figure 3), the analysis was stratified into four subgroups: Asians (6303 cases and 10121 controls), Caucasians (2382 cases and 3340 controls), Americans (12867 cases and 1470 controls), Africans (347 cases and 422 controls), and Mixed (495 cases and 821 controls). Significantly increased risks were found among Asians (OR=1.46, 95% CI=1.07-2.01, and p=0.02) and Africans (OR=2.12, 95% CI=1.57-2.87, and p<0.00001), but not among Caucasians (OR=1.01, 95% CI=0.83-1.22, and p=0.96), Americans (OR=0.99, 95% CI=0.84-1.17, and p=0.93), and Mixed (OR=0.63, 95%

Table 2. Distribution of COX-2-765G>C Genotype and Allele among Digestive System Cancers and Controls

Author	Cases (n)			Controls (n)				Cases (n)	Controls (n)		HWE ^a for control
	CC	GC	GG	CC	GC	GG	C	G	C	G	P
Pereira et al	5	32	36	13	67	130	42	104	93	327	0.28
Liu et al	0	27	220	0	43	384	27	467	43	811	0.27
Hou et al	10	70	210	11	110	288	90	490	132	686	0.9
Saxena et al	19	29	14	8	62	171	67	57	78	402	0.42
Sitarz et al	8	57	176	9	32	59	73	409	50	150	0.14
Tang et al	9	34	57	5	24	76	52	148	34	176	0.11
Zhang et al	35	0	288	41	0	903	70	576	82	1806	0.46
Li et al	2	53	241	1	43	275	57	535	45	593	0.62
Shin et al	0	18	82	0	10	90	18	182	10	190	0.6
Hamajima et al	0	8	140	0	11	230	8	288	11	471	0.023
Cox et al	11	59	150	10	77	170	81	359	97	417	0.73
Koh et al	NA	37*	273	NA	110*	1067	NA	NA	NA	NA	NA
Ulrich et al	10	140	344	20	159	405	160	828	199	969	0.37
Gunter et al	5	54	151	3	52	141	64	356	58	334	0.46
Tan et al	0	81	919	0	63	1237	81	1919	63	2537	0.37
Xing et al	1	17	119	1	29	169	19	255	31	367	0.84
Ueda et al	0	15	440	0	62	989	15	895	62	2040	0.32
Gong et al	9	45	108	12	72	127	63	261	96	326	0.67
Iglesias et al	13	99	172	4	43	76	125	443	51	195	0.48
Thompson et al	11	119	291	15	121	343	141	701	151	807	0.29
Andersen et al	9	83	267	13	186	566	101	617	212	1318	0.61
Hoff et al	10	75	241	8	112	249	95	557	128	610	0.26
Pereira et al	2	38	77	7	83	166	42	192	97	415	0.37
Daraei et al	5	67	38	9	58	53	77	143	76	164	0.20
Wang et al	11	87	207	10	111	238	109	501	131	587	0.49
Chiang et al	0	42	136	0	39	166	42	314	39	371	0.13
Lin et al	NA	104*	193	NA	173*	107	NA	NA	NA	NA	NA
Mittal et al	6	78	92	6	49	41	90	262	61	131	0.08
Guo et al	0	96	930	0	55	1215	96	1956	55	2485	0.43
Moons et al	7	41	92	4	107	384	55	225	115	875	0.24
Upadhyay et al	4	69	101	11	57	148	77	271	79	353	0.09
Kristinsson et al	7	62	153	6	73	157	76	368	85	387	0.47
Bye et al (A) ^b	80	167	100	122	230	110	327	367	474	450	0.94
Bye et al (M) ^b	34	75	81	44	183	195	143	237	271	573	0.91
Xu et al	0	28	255	0	24	542	28	538	24	1108	0.61
Zhao et al	0	36	357	0	30	756	36	750	30	1542	0.59
He et al	4	65	231	1	27	272	73	527	29	571	0.59
Chang et al	0	36	262	0	48	250	36	560	48	548	0.13

^aHWE: Hardy-Weinberg equilibrium; ^b(A) Afrians, ^b(M), Mixed; *Mumbers of GC+CC; NA, not available

Table 3. Stratified Analysis of the COX-2-765G>C Polymorphism on Cancer Risk

Variables		CC+G	C vs GG		C vs G☆			
	N°	Cases/controls	OR (95% CI)	p^*	N	Cases/controls	OR (95% CI)	p^*
Total	38	10814/16174	1.20 (1.00, 1.44)	0.04	36	20624/29422	1.23 (1.06, 1.42)	0.006
Total ^a	36	10207/14717	1.24 (1.05, 1.48)	0.01				
Subgroup by ethni	icity							
Asians	20	6303/10121	1.46 (1.07, 2.01)	0.02	18	11392/17326	1.57 (1.19, 2.06)	0.001
Caucasians	11	2382/3340	1.01 (0.83, 1.22)	0.96	11	4974/6670	1.00 (0.85, 1.18)	0.97
Americans	4	1287/1470	0.99 (0.84, 1.17)	0.93	4	2574/2940	0.97 (0.84, 1.12)	0.71
Afrians	1	347/422	2.12 (1.57, 2.87)	< 0.00001	1	694/924	0.85 (0.69, 1.03)	0.10
Mixed ^b	2	495/821	0.63 (0.29, 1.37)	0.24	2	990/1562	1.12 (0.86, 1.46)	0.39
Subgroup by canc	er type							
Gastric	9	1732/2855	1.64 (1.03, 2.61)	0.04	9	3674/5698	1.55 (0.97, 2.49)	0.07
Colorectal	16	5058/7687	1.01 (0.89, 1.14)	0.92	15	9496/13020	1.00 (0.90, 1.11)	0.96
Oral	3	651/581	0.66 (0.29, 1.50)	0.32	2	708/602	0.95 (0.56, 1.63)	0.86
Esophageal	6	2099/3101	1.32 (0.77, 2.25)	0.32	6	4198/6202	1.31 (0.96, 1.78)	0.09
Others ^c	4	1274/1950	1.89 (0.96, 3.72)	0.06	4	2548/3900	1.85 (0.97, 3.51)	0.06
Subgroup by study	y design							
HCC	18	4409/5423	1.27 (0.92, 1.76)	0.15	17	8434/10274	1.34 (1.02, 1.75)	0.04
PCC	19	6100/10392	1.17 (0.94, 1.46)	0.16	18	11580/18430	1.16 (0.97, 1.39)	0.09
FCC	1	305/359	0.93 (0.67, 1.29)	0.67	1	610/718	0.97 (0.74, 1.29)	0.86

number of case-control studies; *P value for Q-test. Due to that the number of case and control for GC and CC genotype in Koh et al and Lin et al were available, therefore, the comparisons of C vs G did not include the two studies. All studies excluding the studies without HWE. If it was difficult to discriminate the ethnicity of participants according to the data presented, the study was termed mixed. Cancers studied if 1 cancer type was investigated by did individual case-control studies, then it was combined and termed others. population-based case-control study (PCC), hospital-based case-control study (HCC), and family-based case-control study (FCC).

	Experin	nental	Cont	rol		OddsRatio	OddsRatio
Study or Subgroup	Events		Events		Weight	M-H, Random, 95% CI	MH, Random, 95% CI
Linet al 2008	104	297	173	280	2.9%	0.33 [0.24, 0.47]	-
Bye et al 2011 (M)	109	190	352	462	2.8%	0.42 [0.29, 0.60]	
Sitarz et al 2008	85	241	41	100	2.6%	0.53 [0.33, 0.87]	
Ueda et al 2008	15	455	62	1051	2.4%	0.54 [0.31, 0.97]	
Mittal et al 2010	84	176	55	96	2.6%	0.68 [0.41, 1.12]	
Chang et al 2012	36	298	48	298	2.6%	0.72 [0.45, 1.14]	
Hoffet al 2009	85	326	120	369	2.9%	0.73 [0.53, 1.02]	
Gong et al 2009	54	162	84	211	2.7%	0.76 [0.49, 1.16]	
Xing et al 2008	18	137	30	199	2.3%	0.85 [0.45, 1.60]	
Kristinsson et al 2009	69	222	79	236	2.8%	0.90 [0.61, 1.33]	
Houet al 2007	80	290	121	409	2.9%	0.91 [0.65, 1.27]	
Cox et al 2004	70	220	87	257	2.8%	0.91 [0.62, 1.34]	
Wanget al 2012	98	305	121	359	2.9%	0.93 [0.67, 1.29]	
Pereira et al 2010	40	117	90	256	2.6%	0.96 (0.60, 1.52)	
Andersen et al 2009	92	359	199	765	2.9%	0.98 [0.74, 1.31]	
Ulrich 2005	150	494	179	584	3.0%	0.99 [0.76, 1.28]	+
Gunter 2006	59	210	55	196	2.7%	1.00 [0.65, 1.54]	
Iglesias et al 2009	112	284	47	123	2.7%	1.05 [0.68, 1.63]	
Liuet al 2006	27	247	43	427	2.5%	1.10 [0.66, 1.82]	
Thompson et al 2009	130	421	136	479	2.9%	1.13 [0.85, 1.50]	+
Hamajima et al 2001	8	148	11	241	1.7%	1.19 [0.47, 3.04]	
Chianget al 2008	42	178	39	205	2.6%	1.31 [0.80, 2.15]	+
Koh et al 2004	37	310	110	1177	2.8%	1.31 [0.89, 1.95]	
Li et al 2012	55	296	44	319	2.7%	1.43 [0.93, 2.20]	
Daraei et al 2012	72	110	67	120	2.5%	1.50 [0.88, 2.55]	+
Upadhyaya et al 2009	73	174	68	216	2.7%	1.57 [1.04, 2.38]	l—.—
Pereira et al 2006	37	73	80	210	2.5%	1.67 [0.98, 2.86]	
Tan et al 2007	81	1000	63	1300	2.9%	1.73 [1.23, 2.43]	
Moons et al 2007	48	140	111	495	2.7%	1.80 [1.20, 2.71]	l ——
Shin et al 2012	18	100	10	100	1.9%	1.98 [0.86, 4.53]	
Tang et al 2009	43	100	29	105	2.4%	1.98 [1.10, 3.54]	
Bye et al 2011 (A)	247	347	227	422	2.9%	2.12 [1.57, 2.87]	
Guo et al 2007	96	1026	55	1270	2.9%	2.28 [1.62, 3.21]	
Xuet al 2008	28	283	24	566	2.4%	2.48 [1.41, 4.36]	
Zhao et al 2009	36	393	30	786	2.6%	2.54 [1.54, 4.19]	
Zhang et al 2011	35	323	41	944	2.6%	2.68 [1.67, 4.28]	
Zilang et al 2011 He et al 2011	30 89	300	28	300	2.6%		
Saxena et al 2008	48	62	70	241	2.0%	290 [1.81, 4.66] 8.38 [4.34, 16.16]	
Sauci ia ci di AlU0	40	62	70	241	2.2%	0.30 [4.34, 10.10]	
Total (95% CI)		10814		16174	100.0%	1.20 [1.00, 1.44]	•
Total events	2570		3229				
Heterogeneity: Tau ² = 0	.26; Chi² =	259.68,	df = 37 (F	< 0.00	001); l ² = 8	36%	05.07.4.45.6
Test for overall effect: Z					,		0.5 0.7 1 1.5 2

Figure 2. Meta-analysis with a Random-effects Model for the Association between Digestive System Cancer Risk and the *COX-2-765G>C* Polymorphism (GC+CC *vs* GG)

CI=0.29-1.37 and p=0.24). Thus, the Asians and Africans C carriers (CC+GC) may have higher risk of digestive system cancer than others ethnicity. Summary results of other genetic comparisons are listed in Table 3. In the subgroup analysis by cancer type (GC+CC vs GG, Figure 4), the analysis was stratified into five subgroups: gastric cancer (1732 cases and 2855 controls), colorectal cancer (5058 cases and 7687 controls), oral cancer (651 cases and 581 controls), esophageal cancer (2099 cases and 3101 controls), and others (1274 cases and 1950 controls).

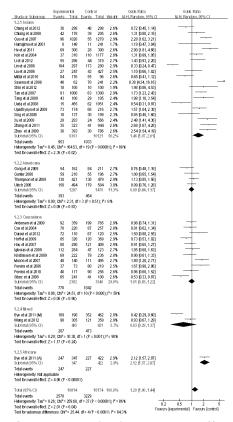


Figure 3. Meta-analysis with a Random-effects Model for the Association between Digestive System Cancer Risk and the *COX-2-765G>C* Polymorphism (GC+CC vs GG): Subgroup Analysis by Ethnicity

Significantly increased risks were found among gastric cancer (OR=1.64, 95% CI=1.03-2.61, and p=0.04), but not among colorectal cancer (OR=1.01, 95% CI=0.89-1.14, and p=0.92), oral cancer (OR=0.66, 95% CI=0.29-1.50, and p=0.32), esophageal cancer (OR=1.32, 95% CI=0.77-

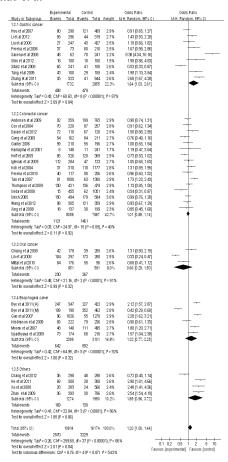


Figure 4. Meta-analysis with a Random-effects Model for the Association between Digestive System Cancer Risk and the *COX-2-765G>C* Polymorphism (GC+CC *vs* GG): Subgroup Analysis by Cancer Type

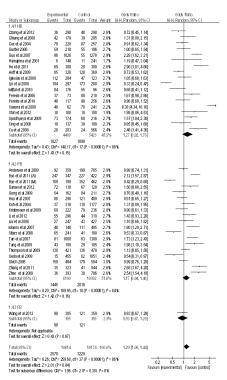


Figure 5. Meta-analysis with a Random-effects Model for the Association between Digestive System Cancer Risk and the *COX-2-765G>C* Polymorphism (GC+CC *vs* GG): Subgroup Analysis by Study Design

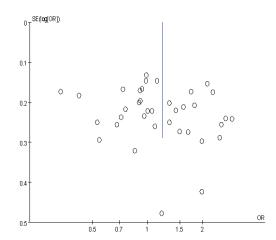


Figure 6. Begg's Funnel Plot for Publication Bias in Selection of Studies on the *COX-2-765G>C* Polymorphism (GC+CC vs GG)

2.25, and p=0.32), and others (OR=1.89, 95% CI=0.96-3.72, and p=0.06). Thus, the gastric cancer C carriers (CC+GC) may have a higher risk than others cancer type. Summary results of other genetic comparisons are listed in Table 3. For subgroup analysis by study design (GC+CC vs GG, Figure 5), no significant association between the -765G>C polymorphism of COX-2 gene and digestive system cancer risk was found in HCC (4409 cases and 5423 controls: OR=1.27,95% CI=0.92-1.76, and p=0.15), PCC (6100 cases and 10392 controls: OR = 1.20,95% CI = 0.99-1.46, and P = 0.16), or FCC (305 cases and 359 controls: OR = 0.93, 95% CI = 0.67-1.29, and p=0.67). Thus, the polymorphism may not increase cancer risk in different study design. Summary results of other genetic comparisons are listed in Table 3.

Sensitivity analysis

In order to assess the stability of the results of the meta-analysis, we performed a sensitivity analysis through sequentially excluded individual studies. After sequentially excluding each case-control study, statistically similar results were obtained for (GC+CC vs GG) (all *P* values were <0.05), suggesting the stability of this meta-analysis (data not shown).

Publication bias

The publication bias was assessed by Begg's funnel plot and Egger's test. The graphical funnel plot of 38 studies of the -765G>C polymorphism of *COX-2* gene appeared to be asymmetrical in the (CC+GC *vs* GG) (Figure 6), which suggested that the combined ORs were not stable, possible explanation may be that probably due to limited number of eligible studies included. Therefore, more studies with large sample size were required to minimize the likelihood of bias.

Discussion

COX-2, known as prostaglandin-endoperoxide synthase 2 (PTGS2), is a rate-limiting enzyme only expressed by various stimulus such as growth factors, cytokines, mitogens (Harrison et al., 1994; Seibert et al., 1994; Wu et al., 1996; Jongthawin et al., 2012).

Evidence suggests that *COX-2* plays an key role in the carcinogenesis pathway, such as in the inhibition of apoptosis, tumor growth, angiogenesis, invasion and metastasis (Leahy et al., 2000; Tatsuguchi et al., 2004; Wang et al., 2007; Gu et al., 2012; Huang et al., 2013; Saad et al., 2013), which are all crucial to many cancers, especially those belonging to the gastrointestinal tract, such as oral cancer (Chiang et al., 2008), gastric cancer (Liu et al., 2006), esophageal cancer (Kristinsson et al., 2009; Upadhyay et al., 2009), and colorectal cancer (Cox et al., 2004).

Given the important roles of COX-2 in cancer etiology, it is possible that genetic variations of the COX-2 gene may affect the susceptibility to cancer development. Genetic variants, such as SNPs in the promoter region of the COX-2 encoding gene, is the most extensively studied polymorphism, which features guanine (G) converting to cytosine (C) at position -765 bp of the promoter region, affecting transcription activity of -765G>C polymorphism of COX-2 gene and its functional activity (Papafili et al., 2002; Szczeklik et al., 2004; Sitarz et al., 2008). To date, conclusions of the association of COX-2-765G>C polymorphism with digestive system cancer is still uncertain; thus, we performed a meta-analysis of 38 case-control studies, including 10814 cases and 16174 controls, to evaluate the associations between the COX-2-765G>C polymorphism and digestive system cancer risks. Considering the genetic background, cancer type and study design may affect the results of meta-analysis, subgroup analyses was performed by ethnicity, cancer type, and study design.

Our results showed that the COX-2-765G>C polymorphism was significantly associated with digestive system cancer risks in the (GC+CC vs GG) genetic model. In addition, in the (C vs G) model, we found significant associations between this polymorphism and digestive system cancer. These results indicated that this polymorphism may contribute to cancer risks. Consistent with the previous meta-analysis (Cao et al., 2010; Liang et al., 2011; Wang et al., 2013), we found significant increased risk -765G>C polymorphism with digestive system cancer, strongly suggesting that this polymorphism may contribute to digestive system cancer pathogenesis and help to explain individual differences of host susceptibility.

Considering the property of genetic background may affect the results of genetic association studies, we performed subgroup analysis by ethnicity. In this meta-analysis, we found that the variant C allele carriers (CC+GC) had increased risk of digestive system cancer in Asians and Africans, but not in Caucasians, Americans, and Mixed, suggesting a possible role of ethnic differences in genetic backgrounds and the environment they lived in. Thus, further studies are demanded to assess the effect of gene-environment interactions in different ethnicities and to validate these findings. In addition, significantly increased risks were found in Africans only one casecontrol study included, it may be due to chance because studies with small sample size may have insufficient statistical power to detect a slight effect. Therefore the results should be explained with great caution. When

stratified separately according to cancer type, we found that this polymorphism was significantly associated with increased risks of gastric cancer. Possible explanation may be that differences in etiology may exist in difference cancer type. Subgroup analysis was also performed by study design, no significant increased risk of digestive system cancer was found among in HCC, PCC, and FCC subjects. Based on our study, it is worth noting that, selection bias was well avoided by performing rigorous and scientific inclusion criteria and exclusion criteria.

Heterogeneity is one of the important issues when performing meta-analysis. We found that heterogeneity between studies existed in overall comparisons. After subgroup analysis by ethnicity or cancer type, the heterogeneity was effectively removed or decreased among Americans and colorectal cancer, possible explanations may be that differences in genetic backgrounds and environmental exposures existed among different ethnicities, and differences in etiology may exist in difference cancer type. Another important factor contributing to heterogeneity was that homogeneity in either the case or control groups was uncertain. Ideally, all cases and controls should be matched for age, sex, and environmental exposures. In this meta-analysis, these issues could not all be explained precisely because of insufficient information for individual person.

Some limitations of this meta-analysis should be acknowledged when explaining our results. Firstly, all eligible studies were published reports written in English and Chinese indexed by the selected databases. It is possible that some potential published studies in other languages or unpublished studies could be missed, which might bias the results. Secondly, some studies were excluded due to lack of original data by email from the corresponding author, we could not evaluate the potential interactions between this polymorphism and digestive system cancer risks, which may lead to a selection bias. Thirdly, this meta-analysis included data from Asians, Caucasians, Americans, Africans and Mixed, no studies from Dutch populations; thus, our study may be applicable to these ethnic groups only. And the last, data were not stratified by other factors such as age, gender, family history, lifestyle variables, because insufficient information could be extracted from the primary publication. It is worth mentioning a study published by Saad et al (Saad et al., 2013), the study indicated potential value of PA extracted from rice bran in reducing colonic cancer risk in rats. Which may have implications for further medical research concerning digestive system cancer and personalized therapy for digestive system cancer patients.

In conclusion, To our knowledge, this is the most comprehensive meta-analysis conducted to date with respect to the associations between the -765G>C polymorphism of COX-2 gene and digestive system cancer risks. Our results indicated the COX-2-765G>C polymorphism was significantly associated with increased risk of digestive system cancer, especially for Asians, Africans and gastric cancer. These results may have implications for further medical research concerning digestive system cancer and personalized therapy for

digestive system cancer patients. Regarding some limitations for this study, future large-scale studies will be needed to clarify the gene-gene and gene-environment interactions to better display the association between the -765G>C polymorphisms in *COX-2* gene and digestive system cancer risks.

Acknowledgements

This study was supported by grants from the National Natural Science Foundation of China (81171909). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

- Akkiz H, Bayram S, Bekar A, et al (2011). Functional polymorphisms of cyclooxygenase-2 gene and risk for hepatocellular carcinoma. *Mol Cell Biochem*, **347**, 201-8.
- Andersen V, Ostergaard M, Christensen J, et al (2009). Polymorphisms in the xenobiotic transporter Multidrug Resistance 1 (MDR1) and interaction with meat intake in relation to risk of colorectal cancer in a Danish prospective case-cohort study. *BMC Cancer*, **9**, 407.
- Bakhle YS (2001). COX-2 and cancer: a new approach to an old problem. Br J Pharmacol, 134, 1137-50.
- Begg CB, Mazumdar M (1994). Operating characteristics of a rank correlation test for publication bias. *Biometrics*, 50, 1088-101.
- Berlau J, Glei M, Pool-Zobel BL (2004). Colon cancer risk factors from nutrition. Anal Bioanal Chem, 378, 737-43.
- Bye H, Prescott NJ, Matejcic M, et al (2011). Populationspecific genetic associations with oesophageal squamous cell carcinoma in South Africa. *Carcinogenesis*, **32**, 1855-61.
- Cao H, Xu Z, Long H, Li XQ (2010). The -765C allele of the cyclooxygenase-2 gene as a potential risk factor of colorectal cancer: a meta-analysis. *Tohoku J Exp Med*, **222**, 15-21.
- Cao Y, Prescott SM (2002). Many actions of cyclooxygenase-2 in cellular dynamics and in cancer. J Cell Physiol, 190, 279-86.
- Chang WS, Yang MD, Tsai CW, et al (2012). Association of cyclooxygenase 2 single-nucleotide polymorphisms and hepatocellular carcinoma in Taiwan. *Chin J Physiol*, **55**, 1-7.
- Chiang SL, Chen PH, Lee CH, et al (2008). Up-regulation of inflammatory signalings by areca nut extract and role of cyclooxygenase-2 -1195G>a polymorphism reveal risk of oral cancer. *Cancer Res*, 68, 8489-98.
- Cook MB, Kamangar F, Whiteman DC, et al (2010). Cigarette smoking and adenocarcinomas of the esophagus and esophagogastric junction: a pooled analysis from the international BEACON consortium. J Natl Cancer Inst, 102, 1344-53.
- Cox DG, Pontes C, Guino E, et al (2004). Polymorphisms in prostaglandin synthase 2/cyclooxygenase 2 (PTGS2/COX2) and risk of colorectal cancer. *Br J Cancer*, **91**, 339-43.
- Daraei A, Salehi R, Mohamadhashem F (2012). PTGS2 (COX2) -765G>C gene polymorphism and risk of sporadic colorectal cancer in Iranian population. *Mol Biol Rep*, **39**, 5219-24.
- Dong J, Dai J, Zhang M, et al (2010). Potentially functional COX-2-1195G>A polymorphism increases the risk of digestive system cancers: a meta-analysis. J Gastroenterol Hepatol, 25, 1042-50.
- Dubois RN, Abramson SB, Crofford L, et al (1998). Cyclooxygenase in biology and disease. *FASEB J*, **12**, 1063-73.

- Egger M, Davey Smith G, Schneider M, Minder C (1997). Bias in meta-analysis detected by a simple, graphical test. *BMJ*, **315**, 629-34.
- Gong Z, Bostick RM, Xie D, et al (2009). Genetic polymorphisms in the cyclooxygenase-1 and cyclooxygenase-2 genes and risk of colorectal adenoma. *Int J Colorectal Dis*, **24**, 647-54.
- Gu KS, Chen Y (2012). Mechanism of P-glycoprotein expression in the SGC7901 human gastric adenocarcinoma cell line induced by cyclooxygenase-2. *Asian Pac J Cancer Prev*, 13, 2379-83.
- Gunter MJ, Canzian F, Landi S, et al (2006). Inflammation-related gene polymorphisms and colorectal adenoma. *Cancer Epidemiol Biomarkers Prev*, **15**, 1126-31.
- Guo Y, Zhang X, Tan W, et al (2007). Platelet 12-lipoxygenase Arg261Gln polymorphism: functional characterization and association with risk of esophageal squamous cell carcinoma in combination with *COX-2* polymorphisms. *Pharmacogenet Genomics*, **17**, 197-205.
- Hamajima N, Takezaki T, Matsuo K, et al (2001). Genotype Frequencies of cyclooxygenease 2 (COX2) rare polymorphisms for Japanese with and without colorectal cancer. *Asian Pac J Cancer Prev*, **2**, 57-62.
- Harrison JR, Lorenzo JA, Kawaguchi H, et al (1994). Stimulation of prostaglandin E2 production by interleukin-1 alpha and transforming growth factor alpha in osteoblastic MC3T3-E1 cells. *J Bone Miner Res*, **9**, 817-23.
- He JH, Li YM, Zhang QB (2011). Cyclooxygenase-2 promoter polymorphism -899G/C is associated with hepatitis B-related liver cancer in a Chinese population of Gansu province. *Chinese Med J*, **124**, 4193-7.
- Hoff JH, te Morsche RH, Roelofs HM, et al (2009). *COX-2* polymorphisms -765G-->C and -1195A-->G and colorectal cancer risk. *World J Gastroenterol*, **15**, 4561-5.
- Hou L, Grillo P, Zhu ZZ, et al (2007). COX1 and COX2 polymorphisms and gastric cancer risk in a Polish population. Anticancer Res, 27, 4243-7.
- Huang F, Lin C, Shi YH, et al (2013). MicroRNA-101 inhibits cell proliferation, invasion, and promotes apoptosis by regulating cyclooxygenase-2 in Hela cervical carcinoma cells. *Asian Pac J Cancer Prev*, **14**, 5915-20.
- Iglesias D, Nejda N, Azcoita MM, et al (2009). Effect of COX2 -765G>C and c.3618A>G polymorphisms on the risk and survival of sporadic colorectal cancer. *Cancer Causes Control*, **20**, 1421-9.
- Jongthawin J, Techasen A, Loilome W, et al (2012). Antiinflammatory agents suppress the prostaglandin E2 production and migration ability of cholangiocarcinoma cell lines. *Asian Pac J Cancer Prev*, **13**, 47-51.
- Kamal MM, Youssef OZ, Lotfy AN, et al (2012). Association of folate intake, dietary habits, smoking and COX-2 promotor -765G>C polymorphism with K-ras mutation in patients with colorectal cancer. J Egypt Natl Canc Inst, 24, 115-22.
- Koh WP, Yuan JM, van den Berg D, et al (2004). Interaction between cyclooxygenase-2 gene polymorphism and dietary n-6 polyunsaturated fatty acids on colon cancer risk: the Singapore Chinese health study. *Br J Cancer*, **90**, 1760-4.
- Kristinsson JO, van Westerveld P, te Morsche RH, et al (2009). Cyclooxygenase-2 polymorphisms and the risk of esophageal adeno- or squamous cell carcinoma. World J Gastroenterol, 15, 3493-7.
- KX Z, YM L, X L (2008). Association of *COX-2* genetic polymorphisms and Hpylori infection with gastric cancer in high-incidence Hexi area of Gansu Province in China. *Shijie Huaren Xiaohua Zazhi*, **16**, 2364-70.
- Leahy KM, Koki AT, Masferrer JL (2000). Role of cyclooxygenases in angiogenesis. *Curr Med Chem*, 7, 1163-70.

- Li Y, Dai L, Zhang J, et al (2012). Cyclooxygenase-2 polymorphisms and the risk of gastric cancer in various degrees of relationship in the Chinese Han population. *Oncol Lett*, **3**, 107-12.
- Liang Y, Liu JL, Wu Y, et al (2011). Cyclooxygenase-2 polymorphisms and susceptibility to esophageal cancer: a meta-analysis. *Tohoku J Exp Med*, **223**, 137-44.
- Lin YC, Huang HI, Wang LH, et al (2008). Polymorphisms of COX-2 -765G>C and p53 codon 72 and risks of oral squamous cell carcinoma in a Taiwan population. Oral Oncol, 44, 798-804.
- Liu F, Pan K, Zhang X, et al (2006). Genetic variants in cyclooxygenase-2: Expression and risk of gastric cancer and its precursors in a Chinese population. *Gastroenterology*, 130, 1975-84.
- Liu JL, Liang Y, Wang ZN (2010). Cyclooxygenase-2 polymorphisms and susceptibility to gastric carcinoma: a meta-analysis. *World J Gastroenterol*, **16**, 5510-7.
- Mittal M, Kapoor V, Mohanti BK, Das SN (2010). Functional variants of COX-2 and risk of tobacco-related oral squamous cell carcinoma in high-risk Asian Indians. Oral Oncol, 46, 622-6.
- Moons LM, Kuipers EJ, Rygiel AM, Groothuismink AZ (2007). COX-2 CA-haplotype is a risk factor for the development of esophageal adenocarcinoma. Am J Gastroenterol, 102, 2373-9.
- Papafili A, Hill MR, Brull DJ, et al (2002). Common promoter variant in cyclooxygenase-2 represses gene expression: evidence of role in acute-phase inflammatory response. Arterioscler Thromb Vasc Biol, 22, 1631-6.
- Pereira C, Pimentel-Nunes P, Brandao C, et al (2010). COX-2 polymorphisms and colorectal cancer risk: a strategy for chemoprevention. Eur J Gastroenterol Hepatol, 22, 607-13.
- Pereira C, Sousa H, Ferreira P (2006). -765G>C COX-2 polymorphism may be a susceptibility marker for gastric adenocarcinoma in patients with atrophy or intestinal metaplasia. World J Gastroenterol, 12, 5473-8.
- Pharoah PD, Dunning AM, Ponder BA, Easton DF (2004). Association studies for finding cancer-susceptibility genetic variants. *Nat Rev Cancer*, 4, 850-60.
- Saad N, Esa NM, Ithnin H (2013). Suppression of beta-catenin and cyclooxygenase-2 expression and cell proliferation in azoxymethane-induced colonic cancer in rats by rice bran phytic acid (PA). Asian Pac J Cancer Prev, 14, 3093-9.
- Saxena A, Prasad KN, Ghoshal UC (2008). Polymorphism of 765G C COX-2 is a risk factor for gastric adenocarcinoma and peptic ulcer disease in addition to H pylori infection A study from northern India. World J Gastroenterol, 14, 1498-503.
- Seibert K, Zhang Y, Leahy K, et al (1994). Pharmacological and biochemical demonstration of the role of cyclooxygenase 2 in inflammation and pain. *Proc Natl Acad Sci USA*, 91, 12013-7.
- Shin WG, Kim HJ, Cho SJ (2012). The *COX-2-1195AA* genotype is associated with diffuse-type gastric cancer in Korea. *Gut Liver*, **6**, 321-7.
- Sitarz R, Leguit RJ, de Leng WW, et al (2008). The *COX-2* promoter polymorphism -765 G>C is associated with early-onset, conventional and stump gastric cancers. *Modern Pathology*, **21**, 685-90.
- Szczeklik W, Sanak M, Szczeklik A (2004). Functional effects and gender association of *COX-2* gene polymorphism G-765C in bronchial asthma. *J Allergy Clin Immunol*, **114**, 248-53.
- Talar-Wojnarowska R, Gasiorowska A, Olakowski M, et al (2011). Role of cyclooxygenase-2 gene polymorphisms in pancreatic carcinogenesis. World J Gastroenterol, 17,

- Tan W, Wu J, Zhang X, et al (2007). Associations of functional polymorphisms in cyclooxygenase-2 and platelet 12-lipoxygenase with risk of occurrence and advanced disease status of colorectal cancer. *Carcinogenesis*, 28, 1197-201.
- Tang XF, Li YM, Li SX (2009). Correlation between the nucleotide polymorphisms of *COX-2* and the susceptibility to gastric cancer in Hui ethnic group. *Shijie Huaren Xiaohua Zazhi*, **17**, 1772-6.
- Tatsuguchi A, Matsui K, Shinji Y, et al (2004). Cyclooxygenase-2 expression correlates with angiogenesis and apoptosis in gastric cancer tissue. *Hum Pathol*, **35**, 488-95.
- Tazawa R, Xu XM, Wu KK, Wang LH (1994). Characterization of the genomic structure, chromosomal location and promoter of human prostaglandin H synthase-2 gene. *Biochem Biophys Res Commun*, **203**, 190-9.
- Thompson CL, PLummer SJ, Merkulova A (2009). No association between cyclooxygenase-2 and uridine diphosphate glucuronosyltransferase 1A6 genetic polymorphisms and colon cancer risk. *World J Gastroenterology*, **15**, 2240.
- Ueda N, Maehara Y, Tajima O, et al (2008). Genetic polymorphisms of cyclooxygenase-2 and colorectal adenoma risk: the Self Defense Forces Health Study. *Cancer Sci*, **99**, 576-81.
- Ulrich CM, Whitton J, Yu JH, et al (2005). PTGS2 (*COX*-2) -765G>C promoter variant reduces risk of colorectal adenoma among nonusers of nonsteroidal anti-inflammatory drugs. *Cancer Epidemiol Biomarkers Prev*, **14**, 616-9.
- Upadhyay R, Jain M, Kumar S, et al (2009). Functional polymorphisms of cyclooxygenase-2 (*COX*-2) gene and risk for esophageal squmaous cell carcinoma. *Mutat Res*, **663**, 52-9.
- Wang J, Joshi AD, Corral R, et al (2012). Carcinogen metabolism genes, red meat and poultry intake, and colorectal cancer risk. *Int J Cancer*, **130**, 1898-907.
- Wang MT, Honn KV, Nie D (2007). Cyclooxygenases, prostanoids, and tumor progression. *Cancer Metastasis Rev*, **26**, 525-34.
- Wang XF, Huang MZ, Zhang XW, et al (2013). *COX-2-765G>C* polymorphism increases the risk of cancer: a meta-analysis. *PLoS One*, **8**, 73213.
- Wu KK (1996). Cyclooxygenase 2 induction: molecular mechanism and pathophysiologic roles. *J Lab Clin Med*, 128, 242-5.
- Xing LL, Wang ZN, Jiang L, et al (2008). Cyclooxygenase 2 polymorphism and colorectal cancer: -765G>C variant modifies risk associated with smoking and body mass index. *World J Gastroenterol*, **14**, 1785-9.
- Xu D, ZHANG XM, Zhao P (2008). Association between single nucleotide polymorphisms in the promoter of cyclooxygenase COX-2 gene and hereditary susceptibility to pancreatic cancer. National Medical J China, 88, 1961-5.
- Yaghoobi M, Rakhshani N, Sadr F, et al (2004). Hereditary risk factors for the development of gastric cancer in younger patients. *BMC Gastroenterol*, **4**, 28.
- Yan WF, Sun PC, Nie CF (2013). Cyclooxygenase-2 polymorphisms were associated with the risk of gastric cancer: evidence from a meta-analysis based on case-control studies. *Tumour Biol*, **34**, 3323-30.
- Zhang X, Miao X, Tan W, et al (2005). Identification of functional genetic variants in cyclooxygenase-2 and their association with risk of esophageal cancer. *Gastroenterology*, **129**, 565-76.
- Zhang X, Zhong R, Zhang Z, et al (2011). Interaction of cyclooxygenase-2 promoter polymorphisms with *Helicobacter pylori* infection and risk of gastric cancer.

- Molecular Carcinogenesis, **50**, 876-83.
- Zhang XM, Miao XP, Tan W (2006). Genetic polymorphisms in the promoter region of cyclooxygenase-2 and their association with risk of gastric cancer. Zhongguo Yi Xue Ke Xue Yuan Xue Bao, 28, 119-23.
- Zhang XM, Zhong R, Liu L (2011). Smoking and COX-2 functional polymorphisms interact to increase the risk of gastric cardia adenocarcinoma in Chinese population. PLoS One, 6, 21894.
- Zhang YG, Huang J, Zhang J, et al (2010). RANTES gene polymorphisms and asthma risk: A meta-analysis. Arch Med Res, 41, 50-8.
- Zhao D, Xu D, Zhang X, et al (2009). Interaction of cyclooxygenase-2 variants and smoking in pancreatic cancer: a possible role of nucleophosmin. Gastroenterology, 136, 1659-68.