

## REVIEW

# **RET Proto Oncogene Mutation Detection and Medullary Thyroid Carcinoma Prevention**

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### **Abstract**

Thyroid cancer is the most common endocrine neoplasia. The medullary thyroid carcinoma (MTC) is one of the most aggressive forms of thyroid malignancy, accounting for up to 10% of all types of this disease. The mode of inheritance of MTC is autosomal dominantly and gain of function mutations in the *RET* proto-oncogene are well known to contribute to its development. MTC occurs as hereditary (25%) and sporadic (75%) forms. Hereditary MTC has syndromic (multiple endocrine neoplasia type 2A, B; MEN2A, MEN2B) and non-syndromic (Familial MTC, FMTC) types. Over the last two decades, elucidation of the genetic basis of tumorigenesis has provided useful screening tools for affected families. Advances in genetic screening of the *RET* have enabled early detection of hereditary MTCs and prophylactic thyroidectomy for relatives who may not show any symptom of the disease. In this review we emphasize the main *RET* mutations in syndromic and non syndromic forms of MTC, and focus on the importance of *RET* genetic screening for early diagnosis and management of MTC patients, based on American Thyroid Association guidelines and genotype-phenotype correlation.

**Keywords:** Genetic screening - MEN2A - MEN2B - *RET* proto-oncogene - medullary thyroid cancer

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### **Introduction**

Neuroendocrine tumors (NETs) include a group of neoplasms of the neuroendocrine cells. The majority of NETs are sporadic. However, some of them may occur as familial inherited syndromes, such as multiple endocrine neoplasia type 1 (MEN1) and multiple endocrine neoplasia type 2 syndrome (MEN2), von Hippel Lindau (VHL) syndrome, neurofibromatosis, and tuberous sclerosis (Oberg, 2013). The term MEN was introduced by Steiner et al in 1968 to describe disorders that include a combination of endocrine tumors (Steiner et al., 1968). Over the last two decades, the genetic basis of tumorigenesis in these familial syndromes has been explained, which providing clinicians with useful screening tools for early diagnosis of affected families and even prevention of the disease in unaffected members (Öberg, 2013). This review explains briefly the medullary thyroid carcinoma, MEN2 syndromes, and mutations found in the main exons of *RET* proto oncogene and the usefulness of *RET* genetic screening in MTC management.

### **Medullary Thyroid Carcinoma**

Thyroid cancer is the most common endocrine neoplasia and accounts for 1% of all human cancers. Medullary thyroid carcinoma (MTC) is a rare malignant

tumor which was first described by Jaquet in the German literature as “malignant goiter with amyloid” (AJ, 1906). In 1959, Hazard et al. (1959) provided a definitive histological description, while Williams suggested that MTC originated from the calcitonin (Ct) secreting parafollicular C cells of the thyroid. Parafollicular C cells derive from the neural crest (Williams, 1966) and share a common neuroectodermal origin with the adrenal medulla. These cells secrete calcitonin, a hormone that plays an important role in the calcium regulation of certain animals but not in humans (Randolph and Maniar, 2000; Strosberg, 2013).

MTC accounts for 5-10% of all thyroid cancers (Leboulleux et al., 2004) and for 13.4% of all thyroid cancer-related deaths (Kebebew et al., 2000; Xu et al., 2012). The reported 10-year mortality rate for patients with MTC varies from 13.5% to 38% (Girelli et al., 1998; Modigliani et al., 1998). The Surveillance, Epidemiology, and End Results (SEER) program of the National Cancer Institute showed that MTC patients had a median age of 50 years at diagnosis and were white in many cases (Mulligan et al., 1993; Roman et al., 2006).

The mentioned tumors account for about 1200 new cancer diagnoses each year in the United States (2% of the 56500 new thyroid cancer cases in 2012) (Siegel et al., 2012). Prognosis can be affected by several factors, such as age, sex, tumor stage, and grading. In general,

patients with tumors confined to the thyroid gland have a 10-year survival rate greater than 95%, whereas patients with regional stage disease had an overall survival rate of 75%. Patients with distant metastases at diagnosis have a poor prognosis, with only 40% surviving 10 year (Roman et al., 2006; Agrawal et al., 2013). If MTC is diagnosed and treated at an early stage, the prognosis of disease will generally be favorable, with 10-year survival rates of 70-80% (Roman et al., 2006). Unfortunately, at the time of diagnosis, up to 70% of patients with MTC have cervical lymph node metastasis, 15% have symptoms of local invasion such as hoarseness or dysphagia, and 10% have distant metastasis (Scollo et al., 2003; Quayle and Moley, 2005).

Surgery is the basis of therapy, total thyroidectomy with bilateral central lymph node dissection is recommended for almost all patients (Kloos et al., 2009). Patients typically die of widespread metastasis affecting liver, lung, and bone or locally aggressive disease with tracheal or esophageal invasion (Agrawal et al., 2013).

MTC can occur as either a sporadic (non-familial) or hereditary disease (Campbell et al., 2013). About 70-80% of MTC cases occur as a sporadic form. Sporadic MTC (sMTC) can arise clinically at any age but its incidence peaks among the fourth and sixth decade of life, with a female dominance of 60% to 40% (Roman et al., 2006). The peak incidence of hereditary MTC (hMTC) can be diagnosed earlier, depending on the availability of genetic and biochemical screenings (LiVolsi, 1997).

Patients with sMTC usually present a solitary and palpable thyroid nodule or cervical lymph node. Most tumors occur in the upper and central areas of the thyroid where C cells are present at higher concentrations. The diagnosis is usually made on fine-needle aspiration (FNA). On average, more than 50% of patients have locoregional lymph node involvement at diagnosis and many present with palpable cervical adenopathy (Moley and DeBenedetti, 1999). Large primary tumor size and multifocality of tumors increases the risk of lymph node involvement (Machens et al., 2007). Distant metastases outside the neck, in the liver, lungs or bones are observed in about 10%-20% of patients at diagnosis (Saad et al., 1984). These patients may present with hormonal symptoms such as flushing and diarrhea, caused by excess secretion of calcitonin (Strosberg, 2013).

The hMTC form accounts for 25% of all MTC cases and it has autosomal dominant inheritance (Figlioli et al., 2013). The estimated prevalence is 2.5 per 100,000 (one in 30000) in the general population (Brauckhoff et al., 2008). hMTC characteristically presents as a multifocal process with C-cell hyperplasia (CCH) as the precursor lesion to MTC. Only familial primary CCH is a preneoplastic lesion (LiVolsi, 1997).

hMTC can be divided into three types: multiple endocrine neoplasia type 2A (MEN2A, 55-60% of all cases), multiple endocrine neoplasia type 2B (MEN2B, 5%-10%), and familial MTC alone (FMTC, 35%-40%) (Öberg, 2013), however more recent reports showed that FMTC as the most prevalent type of hMTC (up to 58%) (Romei et al., 2011). These three types differ with respect to incidence, genetics, age-related penetrance, association

with other diseases, aggressiveness of MTC, and prognosis (Brandi et al., 2001; Kloos et al., 2009).

MEN2 occurs in 1:200,000 live births (Moline and Eng, 2011) and is an autosomal-dominant inherited tumor syndrome with a prevalence of 1/30,000-1/40,000 individuals (Campbell et al., 2013; Figlioli et al., 2013),

In the germline DNA of patients with MEN2, activating mutations in the *RET* (REarranged during Transfection) proto oncogene are found in virtually all cases (Agrawal et al., 2013). The duration for the progression from CCH to microscopic carcinoma in MEN2 remains unclear and it may take years; also the age of transformation from CCH to MTC varies with different germline *RET* mutations. MTC in MEN2 patients is multicentric and concentrated in the upper third of the thyroid gland (Costante et al., 2007).

An early manifestation of all MEN2 syndromes is parafollicular C-cell hyperplasia, which eventually progresses to multicentric neoplasia (Strosberg, 2013). Parafollicular C cells associated with both germline and somatic *RET* mutations and secrete both calcitonin and carcinoembryonic antigen (CEA), which can be used as tumor markers to screen for tumor recurrence (Campbell et al., 2013).

Advances in genetic screening have enabled early detection of hMTCs and prophylactic thyroidectomy for affected kindreds (Strosberg, 2013). Each variant of MEN2 results from a different mutation of the *RET* gene with good genotype-phenotype correlations (Raue and Frank-Raue, 2012).

## Multiple Endocrine Neoplasia Type 2A

According to the literatures, MEN2A is the most common form of MEN2 syndrome (Frank-Raue et al., 2007; Raue and Frank-Raue, 2012). It makes up about 55%-80% or even 90% of cases of MEN2 and hMTC (Eng et al., 1996; Zbuk and Eng, 2007; Moline and Eng, 2011).

MEN2A, also known as "Sipple syndrome" was originally described by Sipple (Sipple, 1961). In 1961, he introduced a man who died of intracranial hemorrhage and autopsy findings were consistent with pheochromocytoma (PHEO), MTC, and parathyroid hyperplasia. The following year, Cushman (Cushman Jr, 1962) put a link between these endocrine tumors, which led to the classification of MEN2 as a unique entity.

Today, MEN2A is defined by MTC, bilateral PHEO, and multiple tumors of the parathyroid glands (parathyroid adenomas, primary hyperparathyroidism or HPT), within a single patient or family (Raue and Frank-Raue, 2012). The respective frequency of these tumors are 90%-100% for MTC (the penetrance of MTC in families with MEN2A is 100%) (Campbell et al., 2013), 40%-50% for PHEO, and 10%-25% for parathyroid hyperplasia (Raue and Frank-Raue, 2012). MTC is generally the first manifestation of MEN2A and presents when patients are between 5 and 25 years of age. Diarrhea, the most frequent systemic manifestation, occurs in affected individuals with a plasma calcitonin concentration of more than 10ng/mL and infers a poor prognosis. Patients with MTC typically present with a neck mass or neck pain, usually before age 35 years. Up to 70% of such individuals already have cervical lymph

node metastases (Cohen and Moley, 2003).

PHEO usually presents after MTC or concomitantly; however, it is the first symptom in 13-27% of individuals with MEN2A. PHEO in persons with MEN2A and

MEN2B are diagnosed at an earlier age, have subtler symptoms, and are more likely to be bilateral than sporadic tumors (Pacak et al., 2005).

HPT in MEN2A is typically mild and may range from

**Table 1. Genotype-Phenotype Correlations and Risk Levels for Aggressive Medullary Thyroid Cancer (Kloos et al., 2009)**

| Mutation Location                    | Exon        | Mutation                              | Phenotype         | ATA Risk Level |                  |   |
|--------------------------------------|-------------|---------------------------------------|-------------------|----------------|------------------|---|
| Extra cellular cadherine like domain | 5           | G321R                                 | FMTC/MEN2A        | A              |                  |   |
| Extra cellular cysteine rich domain  | 8           | C515S                                 | FMTC/MEN2A        | A              |                  |   |
|                                      |             | 532duplication                        | FMTC              | A              |                  |   |
|                                      |             | 529/531                               | FMTC              | ?              |                  |   |
|                                      |             | G533C                                 | FMTC/MEN2A        | A              |                  |   |
|                                      |             | 532duplication                        | FMTC              | A              |                  |   |
|                                      |             | 531/9bp duplication                   | FMTC/MEN2A        | A              |                  |   |
|                                      |             | R600Q                                 | FMTC/MEN2A        | A              |                  |   |
|                                      |             | K603E                                 | FMTC/MEN2A        | A              |                  |   |
|                                      |             | K603Q                                 | FMTC              | A              |                  |   |
|                                      |             | Y606C                                 | FMTC              | A              |                  |   |
|                                      | 10          | C609F/R/G/S/Y                         | FMTC/MEN2A/HSCR   | B              |                  |   |
|                                      |             | C611R/G/F/S/W/Y                       | FMTC/MEN2A/ HSCR  | B              |                  |   |
|                                      |             | C618R/G/F/S/Y                         | FMTC/MEN2A/ HSCR  | B              |                  |   |
|                                      |             | C620R/G/F/S/W/Y                       | FMTC/MEN2A/ HSCR  | B              |                  |   |
|                                      |             | C630R/F/S/Y                           | FMTC/MEN2A        | B              |                  |   |
|                                      |             | D631Y                                 | FMTC              | B              |                  |   |
|                                      |             | 633                                   | MEN2A             | '?'            |                  |   |
|                                      |             | 633/9bpduplication                    | FMTC/MEN2A        | B              |                  |   |
|                                      |             | 634/12bp duplication                  | FMTC/MEN2A        | B              |                  |   |
|                                      |             | C634R                                 | MEN2A/CLA         | C              |                  |   |
|                                      | 11          | C634G/F/S/W/Y                         | FMTC/MEN2A/CLA    | C              |                  |   |
|                                      |             | 634/12bp duplication                  | MEN2A             | B              |                  |   |
|                                      |             | 635/insertion ELCR/T636P              | FMTC/MEN2A        | A              |                  |   |
|                                      |             | 637                                   | MEN2A             | ?              |                  |   |
|                                      |             | S649L                                 | FMTC/MEN2A        | A              |                  |   |
|                                      |             | Intra cellular tyrosine kinase domain | 13                | K666E          | FMTC/MEN2A       | A |
|                                      |             |                                       |                   | E768D          | FMTC/MEN2A/ sMTC | A |
| N777S                                | FMTC        |                                       |                   | A              |                  |   |
| 778                                  | FMTC        |                                       |                   | ?              |                  |   |
| N776S                                | FMTC/MEN2A  |                                       |                   | A              |                  |   |
| 14                                   | 781         |                                       | FMTC              | ?              |                  |   |
|                                      | L790F       |                                       | FMTC/MEN2A        | A              |                  |   |
|                                      | Y791F       |                                       | FMTC/MEN2A        | A              |                  |   |
|                                      | V804L       |                                       | FMTC/MEN2A        | A              |                  |   |
|                                      | V804M       |                                       | FMTC/MEN2A/sMTC   | A              |                  |   |
|                                      | V804M+E805K |                                       | MEN2B             | D              |                  |   |
|                                      | V804M+Y806C |                                       | MEN2B             | D              |                  |   |
|                                      | G819K       |                                       | FMTC              | A              |                  |   |
|                                      | R833C       |                                       | FMTC/ MEN2B       | A              |                  |   |
|                                      | R844Q       |                                       | FMTC/ MEN2B       | A              |                  |   |
|                                      | V804M/E805K |                                       | MEN 2B            | D              |                  |   |
|                                      | V804M/Y806C |                                       | MEN 2B            | D              |                  |   |
|                                      | 804/844     |                                       | FMTC              | '?'            |                  |   |
|                                      | 852         |                                       | FMTC              | '?'            |                  |   |
| 15                                   | R866W       | FMTC/MEN2A                            | A                 |                |                  |   |
|                                      | 876         | FMTC                                  | '?'               |                |                  |   |
|                                      | A883F       | MEN2B/ sMTC                           | D                 |                |                  |   |
|                                      | S891A       | FMTC/MEN2A/                           | A                 |                |                  |   |
|                                      | 16          | R912P                                 | FMTC/MEN2A/ MEN2B | A              |                  |   |
| M918T                                |             | sMTC/MEN2B                            | D                 |                |                  |   |
| 920                                  |             | sMTC/MEN2B                            | D                 |                |                  |   |
| 922                                  |             | sMTC/MEN2B                            | D                 |                |                  |   |
| 13/14                                |             | V804M+V778I                           | FMTC/MEN2A        | B              |                  |   |
| 14/15                                |             | V804M+S904C                           | MEN2B/ MEN2A      | D              |                  |   |
| 13/16                                |             | 768/919                               | FMTC              | '?'            |                  |   |
| 14/15                                | V804M/S904C | MEN 2B                                | D                 |                |                  |   |

a single adenoma to marked hyperplasia. Most individuals with HPT have no symptoms; however, hypercalciuria and renal calculi may occur. If HPT is longstanding and unrecognized, symptoms may become severe. HPT usually presents many years after the diagnosis of MTC and the average age of its onset is 38 years (Pacak et al., 2005).

Rare variants of MEN2A also exist, including MEN2A with cutaneous lichen amyloidosis (CLA); this skin lesion is located over the upper portion of the back which may appear before the onset of MTC (Donovan et al., 1989; Gagel et al., 1989; Moline and Eng, 2011). Another rare variant is MEN2A/FMTC with Hirschsprung's disease (Raue and Frank-Raue, 2012), which occurs in about 16% of the cases (Panza et al., 2012).

The clinical course of MTC in patients with MEN2A is variable and the disease progression is associated with codon specific mutations (Eng et al., 1996; Machens et al., 2001). The majority of mutations in MEN2A kindred involve the cysteine-rich domain of the *RET* protein, including exons 10 and 11 (Randolph and Maniar, 2000; Fishbein and Nathanson, 2012).

All individuals with an MTC-predisposing *RET* mutation who have not undergone prophylactic thyroidectomy demonstrate biochemical evidence of MTC by age 35 years (Cohen and Moley, 2003).

## Multiple Endocrine Neoplasia Type 2B

Initial descriptions of MEN2B were reported by Wagenmann in 1922 (A, 1922), Froboese in 1923 (C, 1923), and then in 1966. The constellation of findings associated with MEN2B including thick lips, tongue lesions, MTC and PHEO was described in a patient and his family members by Williams and Pollock (Williams and Pollock, 1966; Williams ED, 1966).

MEN2B accounts for about 5-10% of all cases affected with MEN2 syndrome (Moline and Eng, 2011). MEN2B is characterized by the early development of an aggressive form of MTC in all affected individuals (typically during the first year of life), PHEO (about 50% of individuals), the absence of hyperparathyroidism, and visible physical stigmata such as raised bumps on the lips and tongue (caused by cutaneous neuromas), ganglioneuromas of the intestine, and asthenic marfanoid body habitus with skeletal deformations often with kyphoscoliosis or lordosis, joint laxity, decreased subcutaneous fat, and proximal muscle wasting and weakness can also be seen (Raue and Frank-Raue, 2012). The lips become prominent (or "blubbery") over time, and submucosal nodules may be present on the vermilion border of the lips (Moline and Eng, 2011). The mucosal neuromas on the tongue and lips (bumpy lips) resulting in a distinctive facial appearance, together with a marfanoid body habitus (65%-75%), allows for diagnosis at a first glance (Moline and Eng, 2011). Interestingly, individuals with MEN2B may be identified in infancy or early childhood by the presence of mucosal neuromas on the anterior dorsal surface of the tongue, palate, or pharynx and a distinctive facial appearance. However, early diagnosis by clinical means is difficult because of the gradual development of the typical

clinical appearance during childhood (Wray et al., 2008).

About 40% of affected individuals have diffuse ganglioneuromatosis of the gastrointestinal which is responsible for the megacolon, constipation, and diarrhea that often present. Because of their similar clinical presentations, the clinician should carefully differentiate the diagnosis of Hirschsprung disease as a result of aganglioneuromatosis from MEN2B (Raue and Frank-Raue, 2012). Neuromas of the eyelids may cause thickening and eversion of the upper eyelid margins. Prominent thickened corneal nerves may be seen by slit lamp examination (Moline and Eng, 2011).

Brauckhoff et al. reported that an inability to cry tears is also one of the symptoms of MEN2B. It is the rarest form of MEN2 and accounts for 5-10% of such cases (Brauckhoff et al., 2008).

Individuals with MEN2B who do not undergo prophylactic thyroidectomy at an early age (1 year), are likely to develop metastatic MTC at an early age. Before intervention with early prophylactic thyroidectomy, the average age of death in individuals with MEN2B was 21 years (Moline and Eng, 2011).

Patients with MEN2B often do not have a family history of the disease; in more than 50% of cases the syndrome is due to a de novo germline *RET* mutation, especially in exons 16 and 15 (Majidi et al., 2011; Raue and Frank-Raue, 2012).

## Familial Medullary Thyroid Carcinoma

FMTC is characterized by a strong predisposition to MTC in families with a very low incidence of other endocrinopathies related to MEN2 (Raue and Frank-Raue, 2012). It is also considered as a "clinical variant of MEN2A in which MTC is the only manifestation" (Moline and Eng, 2011). Another characteristic feature of FMTC is the presence of amyloid deposits between tumor cells (Moley et al., 1998; Raue and Frank-Raue, 2009). Actually, FMTC is the mildest variant in which patients have familial, often more benign MTC and by definition no incidence of other endocrine neoplasms (Öberg, 2013).

FMTC is frequently bilateral and multifocal, and is generally the least aggressive phenotype, not appearing until the second or third decade of life (Lin, 2011). FMTC has been diagnosed more frequently in recent years and is reported to account for 35-60% of all MTC cases (Raue and Frank-Raue, 2012). Regarding MTC, in FMTC, penetrance is lower and the clinical course is more benign than in MEN2A and MEN2B, with a late onset or no clinically manifesting disease, and the prognosis is relatively good. Therefore, a family history is often inadequate for establishing familial disease (Raue and Frank-Raue, 2012).

The diagnosis of FMTC can only be considered when four or more family members across a wide range of ages have or have had isolated MTC (Raue and Frank-Raue, 2009). Careful genetic and biochemical screening often reveals a family history of MTC in patients originally thought to have the sporadic form of the disease (Figlioli et al., 2013).

To avoid neglecting the risk of PHEO, strict criteria



should be met before a family is classified as having FMTC. In the past, some believed that a kindred with FMTC should have more than 10 members with a *RET* mutation, multiple individuals with a *RET* mutation over the age of 50 years, and all members should have an adequate medical history demonstrating a lack of PHEO or PHPT (Machens et al., 2001; Machens and Dralle, 2006). In fact, the diagnosis of FMTC is based on the absence of PCC and PHPT in two or more generations within a family or by providing evidence of a *RET* mutation in a codon associated with susceptibility to only MTC, including codons 609, 611, 618, 620, 768, 790, 804, and 891 (Wohllk et al., 2010). In families with single affected generation or smaller families, the FMTC classification should be done cautiously, as there is the possibility of failure to PHEO and MEN2A diagnosis (Kloos et al., 2009).

### ***RET* Proto Oncogene**

The location of *RET* proto-oncogene was determined on human chromosome 10q11.2 in 1985, and the gene subsequently named *RET* [*RE-arranged during Transfection*] after it was found to be rearranged during transfection in 3T3 cell lines with DNA from lymphoma cells. In fact, it was shown to be activated by a DNA rearrangement (Takahashi et al., 1985).

After two years of *RET* locus identification, the genes responsible for MEN2 were found to be localized in centromeric region of chromosome 10 [10q11.21] by genetic linkage analysis in 1987 (Donis-Keller et al., 1993).

*RET* contains 21 exons and encodes a tyrosine kinase receptor (RTK) protein (Ceccherini et al., 1993). Its GC rich promoter locates in the nucleotide sequence of exon 1. The main *RET* transcript length is 5659 base pairs and is translated to 1114 amino acid-residue protein (Machens et al., 2003). The *RET* protein is 170 KDa, present on the cell surface, and highly phosphorylated on tyrosine residues (Asai et al., 1995).

### ***RET* Protein Kinase Structure**

*RET* is expressed primarily in peripheral enteric, sympathetic and sensory neurons, central motor, dopamine and noradrenaline neurons. It is also expressed in branching ureteric bud during embryogenesis and in differentiating spermatogonia (Pachnis et al., 1993; Meng et al., 2000).

*RET* has three different transcripts, encoding *RET* isoforms. *RET* exon 19 is present in all transcripts, however, differential splicing at the 3' end of exon 19 results in transcripts where exon 19 is unspliced, spliced to exon 20, or spliced to exon 21 (Ivanchuk et al., 1998). These transcripts encode *RET* isoforms with distinct c-terminal ends of 9 (*RET*9), 51 (*RET*51), or 43 (*RET*43) amino acids. *RET*9 and *RET*51, consisting of 1072 and 1114 amino acids respectively, are the main isoforms *in vivo*. These two isoforms are co-expressed in most tissues but have distinct developmental roles and different gene expression profiles, suggesting possible differences in downstream regulation of cell-cell interaction pathways (Takahashi et al., 1993; Myers et al., 1995; Hickey et

al., 2009). The final common amino acid for all three isoforms is a tyrosine (Y1062) which is phosphorylated during *RET* activation. Thus, alternative splicing places Y1062 in different amino acids contexts in the three *RET* isoforms, conferring different binding potentials (Arighi et al., 1997; Lorenzo et al., 1997).

*RET* has three domains, including an N-terminal extracellular segment which is included a ligand-binding domain, a cadherin (Ca<sup>2+</sup>-dependent cell adhesion)-like domain, and a cysteine-rich domain (near the cell membrane). This domain is a ligand for an activator protein called glial cell derived neurotrophic factor (GDNF) (Arighi et al., 2005).

The two other domains are a hydrophobic transmembrane domain, and an intracellular TK domain (Arighi et al., 2005). The TK domain contains multiple tyrosine residues (16 in *RET*9 and 18 in *RET*51), two of which, at positions 1019 and 1051, are only present in *RET*51 (Asai et al., 1995). The transmembrane domain ensures the close proximity of the *RET* monomers through noncovalent receptor-receptor interactions. The intracellular portion contains two TK subdomains, which are phosphorylated during receptor activation and are involved in the activation of the intracellular signaling pathways (de Groot et al., 2006; Colombo-Benkmann et al., 2008).

### ***RET* Protein Kinase Function**

The ligands of the *RET* receptor TK are growth factors belonging to the GDNF family or GDNF family ligands (GFLs), including GDNF, neurturin (NRTN), artemin (ARTN), and persephin (PSPN). Without the ligand, *RET* is monomeric, unphosphorylated, and inactive. Binding of the ligand due to GFR $\alpha$  co-receptors to the extracellular domain of *RET* induces receptor dimerization and autophosphorylation, creating intracellular binding sites for signaling proteins with the subsequent activation of multiple signaling pathways (Phay and Shah, 2010). In other words, once GFL binds to *RET* receptor, an intracytoplasmic domain within upstream portion of *RET* is autophosphorylated, which stabilizes the protein, and is necessary for further downstream activity of *RET* autophosphorylation cascade. In fact, phosphorylation of Tyr981, as well as of Tyr1015, Tyr1062 and Tyr1096, is important for initiating intracellular signal transduction processes (Airaksinen and Saarma, 2002). *RET* signaling is thought to provide growth and survival signaling via the RAF-MEK-ERK and PI3K-AKT-mTOR pathways (Takahashi, 2001).

### ***RET* Proto Oncogene Mutations**

*RET* was one of the first RTKs found to play a role in neoplasia (Figlioli et al., 2013). Interestingly, oncogenic activation of *RET* has been shown to activate both PI3K/AKT- and RAS/RAF/MAPK-dependent cell signaling (Segouffin-Cariou and Billaud, 2000; Raue and Frank-Raue, 2010). Various autosomal dominantly inherited mutations have been shown to cause a constitutive activation of *RET* (Figlioli et al., 2013). Both gain- and loss-of-function of *RET* mutations have been identified in

human diseases (Takahashi, 2001; Santoro et al., 2004). Gain of function mutations in *RET* have been implicated in the pathogenesis of several forms of thyroid cancer which leads to autophosphorylation of tyrosine sites within *RET* (Salehian and Samoa, 2013). Single point mutations in the extracellular domain of *RET* have been implicated in the malignant transformation of cells that originate from the neural crest during embryogenesis (Manie et al., 2001), such as the parafollicular cells of the thyroid. These point mutations can occur as both germline and somatic mutations (Boikos and Stratakis, 2008).

Germline missense mutations of *RET* are associated with MEN2A, MEN2B, and FMTC, as described previously (Takahashi et al., 1985), whereas sporadic MTC is thought to be due to a somatic mutation in the tumor cells (Romei et al., 1996). *RET* mutations are mostly missense and located in exons 10, 11 (extracellular domain of RET), 13, 14, 15, and 16 (in the TK domain) (Segouffin-Cariou and Billaud, 2000; Raue and Frank-Raue, 2010; Hedayati et al., 2011) (Table1). Mutation of the extracellular cysteine in codon 634 in exon 11 of *RET* causes ligand-independent dimerization of receptor molecules, enhanced phosphorylation of intracellular substrates and cell transformation. Mutation of the intracellular TK (for example codon 918) has no effect

on receptor dimerization but does cause constitutive activation of intracellular signaling pathways and also results in cellular transformation (Machens et al., 2003).

Further, rearrangements of *RET* (*RET/PTC*) are associated with papillary thyroid carcinoma (PTC) commonly seen in tumors of children and tumors associated with radiation exposure (Tallini and Asa, 2001).

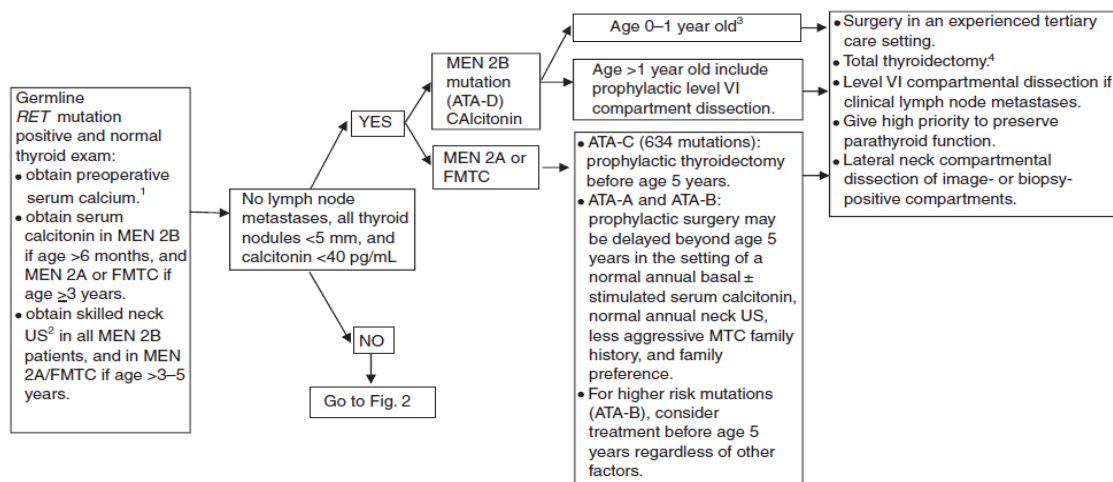
*RET* loss-of-function mutations are associated with a genetic disorder of neural crest development known as Hirschsprung disease (Arighi et al., 2005), which is a congenital absence of the enteric nervous system in the hindgut and lack of ganglion cells in the colon (Edery et al., 1994). Interestingly, some overlap has been reported between MEN2 and Hirschsprung’s phenotypes [especially MEN2A/Hirschsprung] (Oberg, 2013).

The various *RET* mutations relate not only to the disease phenotype but also the clinical aggressiveness of MTC (Eng et al., 1996).

Analysis of *RET* in families with MEN2A and FMTC revealed that nearly all these families have germline mutations, and that only those family members with the germline missense mutations have the disease (Frank-Raue et al., 2007). Actually, germline *RET* mutations are observed in 98% of MEN2A, 95% of MEN 2B, and 88% of FMTC kindreds (Figlioli et al., 2013). Significantly,

**Table 2. American Thyroid Association Risk Level and Prophylactic Thyroidectomy Testing and Therapy (Kloos et al., 2009)**

| ATA risk level | Age of RET testing                               | Age of required first US                         | Age of required first serum Ct        | Age of prophylactic surgery  |
|----------------|--|--|---------------------------------------|--|
| D              | ASAP and within the 1 <sup>st</sup> year of life | ASAP and within the 1 <sup>st</sup> year of life | 6 months, if surgery not already done | ASAP and within the 1 <sup>st</sup> year of life   |
| C              | <3-5 years                                       | >3-5 years                                       | >3-5 years                            | Before age 5 years   |
| B              | <3-5 years                                       | >3-5 years                                       | >3-5 years                            | Consider surgery before age 5. May delay surgery beyond age 5 years if stringent criteria are met. |
| A              | <3-5 years                                       | >3-5 years                                       | >3-5 years                            | May delay surgery beyond age 5 years if stringent criteria are met.                                |



<sup>1</sup>Treat hyperparathyroidism with 4 gland resection and autograft to heterotopic site, or subtotal parathyroidectomy. Consider cryopreservation. PHEO preoperative screening should begin by age 8 years for MEN 2B and mutated *RET* codons 634 and 630; otherwise by age 20 years for other *RET* mutations.  
<sup>2</sup>Neck US to include the superior mediastinum and central and lateral neck compartments.  
<sup>3</sup>Insufficient data to recommend routine prophylactic level VI compartmental dissection.  
<sup>4</sup>Parathyroid glands resected or devascularized should be autografted in the neck in *RET*-negative, MEN 2B, and FMTC patients, while MEN 2A glands should be autografted to a heterotopic site.

**Figure 1. Initial Diagnosis and Therapy of Pre-clinical Disease (after Kloos et al., 2009)**

30%-50% of sMTC tumors also bear somatic *RET* mutations, the majority resembling those seen in the most aggressive hereditary form, MEN2B (Elisei et al., 2008; Agrawal et al., 2013). Most of these are the M918T mutation. *RET* mutations in sMTC cases were associated with increased risk of nodal and distant metastases, as well as a significantly inferior survival rate (Strosberg, 2013).

Activating mutations of *RET* involving exons 10, 11, 13, 14, 15 have been proved to cause MEN2A. The most common mutations involve exons 10 and 11 encoding the highly conserved cysteine-rich domain (Marx and Stratakis, 2005). The mutations for MEN2A are mostly located in exon 10, including codons 609, 611, 618, and 620 (10-15%), and exon 11, including codons 630 and 634 (80-85%) (Donis-Keller et al., 1993; Mulligan et al., 1993; Eng et al., 1996). Hyperparathyroidism in MEN2A shows a strong association with the presence of a mutation in codon 634 and C634R in particular (Eng et al., 1996; Hedayati et al., 2006; Alvandi et al., 2011; Masbi et al., 2014). Other uncommon mutations in the intracellular TK domain have also been reported in MEN2A kindred (Marx and Stratakis, 2005; Hoff and Hoff, 2007).

Pheochromocytoma is associated with exon 634 and 918 mutations in approximately 50% of patients, with exon 10 mutations (codons 609, 611, 618 and 620) in up to 17% of patients (Frank-Raue et al., 2011), and rarely with mutations in exons 13-15 (codons 791 and 804) (Machens et al., 2005; Quayle et al., 2007) and exon 8 (codon 533) (Donis-Keller et al., 1993; Mulligan et al., 1993; Eng et al., 1996).

There is a significant age-related progression from CCH to MTC that correlates with the transformative capacity for the particular *RET* mutation. C-cells are more susceptible to oncogenic *RET* activation than adrenal

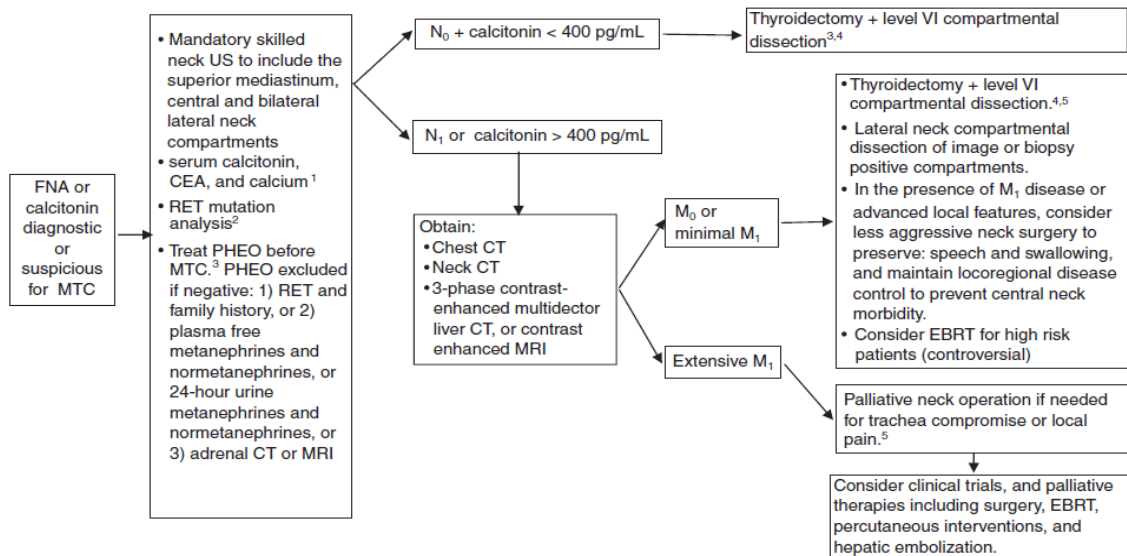
medullary or parathyroid cells (Machens et al., 2003).

The mutations characteristic of FMTC also occur in exons 10 and 11. However, non-cysteine point mutations also have been found in exon 8 (codons 532 and 533), exon 13 (codons 768,790, and 791), exon 14 (codons 804 and 844), exon 15 (codon 891), and exon 16 (codon 912) (Mulligan et al., 1994; Eng et al., 1996; Kouvaraki et al., 2005). A recently meta-analysis has shown that since 1993, a total of 39 different *RET* germline mutations were identified in FMTC patients from different families. Except for a 9-bp duplication (after codon 531, exon8), all mutations were missense type and were scattered among the exons 5, 8, 10, 11, 13, 14, 15, and 16. In FMTC, specific germline *RET* mutations were widely correlated with age-specific penetrance of cancer development and nodal metastases (Figlioli et al., 2013). Altogether, FMTC has been most commonly associated with mutations in codons 609, 611, 618, and 620 in exon 10; codon 768 in exon 13; and codon 804 in exon 14. When FMTC is associated with mutations in codon 634 in exon 11, it is almost never C634R and is most commonly C634Y (Eng et al., 1996).

About 95% of MEN2B patients carry a M918T mutation within exon16 and 5% have an A883F mutation in exon15 (Iwashita et al., 1996; Miyauchi et al., 1999). Mutation in codon 918 gives a more aggressive phenotype. Rare MEN2B patients compound heterozygotes for *RET* mutations (V804M/Y806C) were also observed (Gimm et al., 1997; Miyauchi et al., 1999).

## RET Genetic Screening:

Genetic testing for *RET* germline mutation (Figures 1-5) has shown 100 % sensitivity and specificity to



<sup>1</sup>Treat hyperparathyroidism with 4 gland resection and autograft to heterotopic site, or subtotal parathyroidectomy. Consider cryopreservation.

<sup>2</sup>Ideally performed with genetics counseling and completed preoperatively.

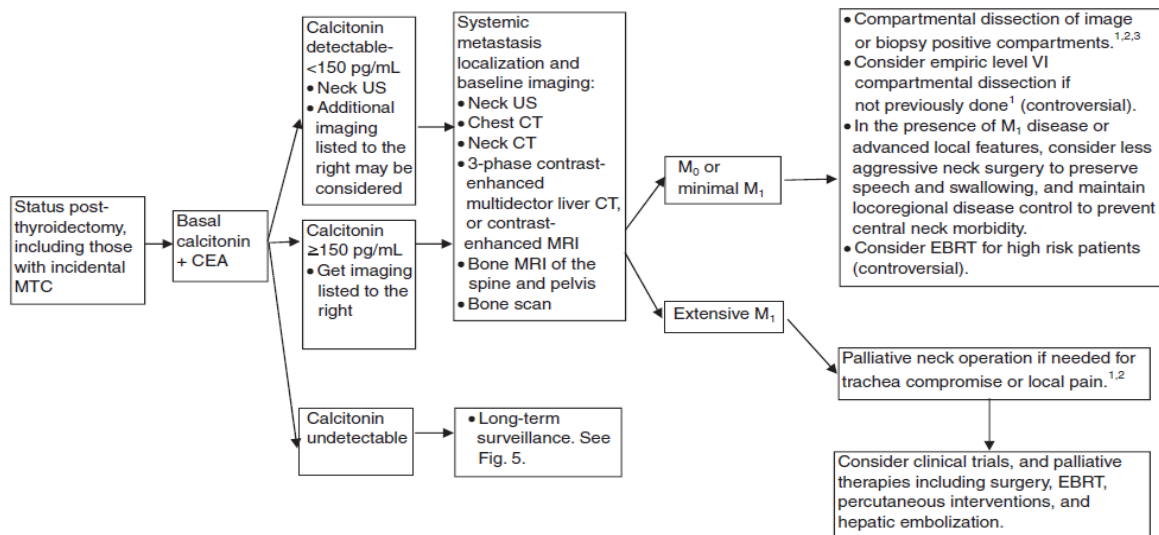
<sup>3</sup>PHEO preoperative screening should begin by age 8 years for MEN 2B and mutated *RET* codons 634 and 630; and by age 20 years for other *RET* mutations.

<sup>4</sup>Parathyroid glands resected or devascularized should be autografted in the neck in *RET*-negative, MEN 2B, and FMTC patients, while MEN 2A glands should be autografted to a heterotopic site.

<sup>5</sup>Consider external beam radiation of TNM stage T4 disease to prevent recurrent local disease.

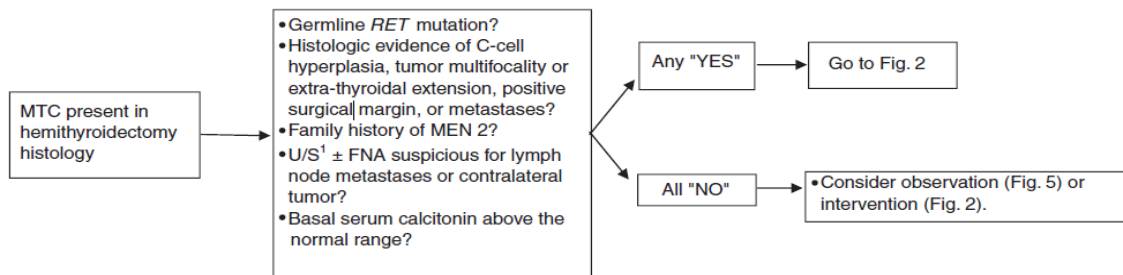
FNA, fine-needle aspiration biopsy.

Figure 2. Initial Diagnosis and Therapy of Clinically Apparent Disease (after Kloos et al., 2009)



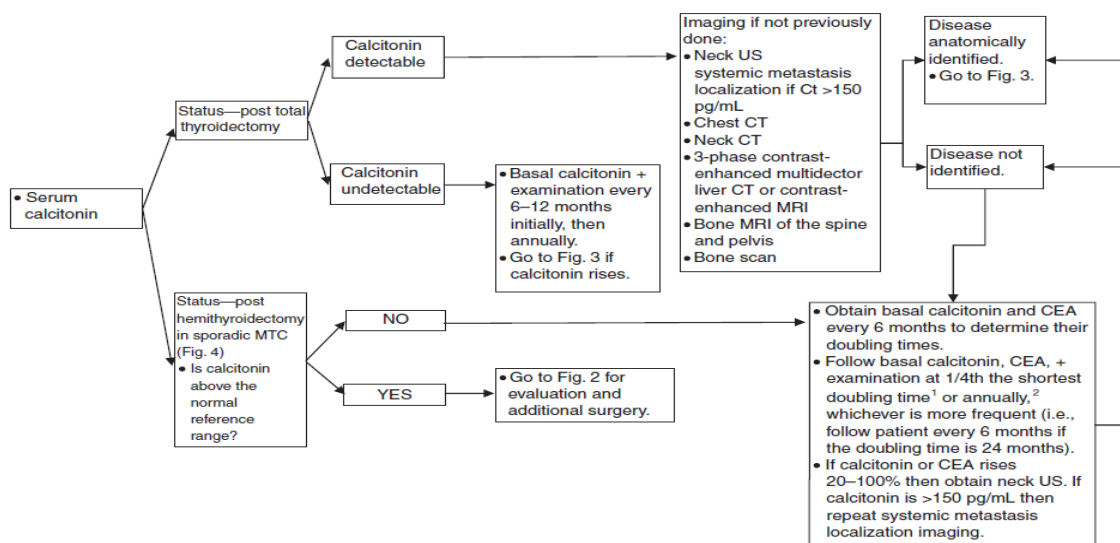
<sup>1</sup>Parathyroid glands resected or devascularized should be autografted in the neck in *RET*-negative, MEN 2B, and FMTC patients, while MEN 2A glands should be autografted to a heterotopic site.  
<sup>2</sup>Consider external beam radiation of T<sub>4</sub> disease to prevent recurrent local disease.  
<sup>3</sup>Observation of nonthreatening locoregional disease <1 cm may be considered.

Figure 3. Initial Evaluation and Treatment of Postoperative Patients (after Kloos et al., 2009)



<sup>1</sup>Neck US to include the superior mediastinum and central and lateral neck compartments.

Figure 4. Management of MTC after Hemithyroidectomy (after Kloos et al., 2009)



<sup>1</sup>Doubling time may be estimated or optimally calculated by fitting data to single exponentials by nonlinear least-square analysis (calculator available at [www.thyroid.org](http://www.thyroid.org)).

<sup>2</sup>Patients with *RET* mutations associated with PHEO or primary hyperparathyroidism should be screen annually beginning at age 8 years in MEN 2B and mutated *RET* codons 634 and 630, and from age 20 years in carriers of other MEN 2A *RET* mutations, while those associated only with FMTC should be screened at least periodically.

Figure 5. Long Term Surveillance (after Kloos et al., 2009)



identify individuals at risk of MTC development. Use of this genetic assay allows earlier and more definitive identification and clinical management of persons with a familial risk for MTC when compared to the existing standard of annual biochemical monitoring, such as serum calcitonin. Once a person is found to be positive for a *RET* mutation, they must be carefully counseled. At-risk family members need to be identified and should undergo genetic testing, because even asymptomatic patients that are identified as *RET* mutation carriers can be offered a prophylactic thyroidectomy (Sippel et al., 2008).

Since hMTC can be prevented with prophylactic thyroidectomy, the American Thyroid Association (ATA) recommends that all patients with MTC be offered germline *RET* testing (Strosberg, 2013). Recommendations for the timing of prophylactic thyroidectomy and the extent of surgical resection is based on a model which uses the genotype-phenotype correlations to stratify mutations into risk levels (A-D). ATA level D (ATA-D) mutations carry the highest risk for MTC. These mutations include codons 883 (exon 15) and 918 (exon 16) that are associated with the youngest age of onset and highest risk of metastases and mortality. ATA level C (ATA-C) mutations carry a lower, yet still high risk of aggressive MTC and include mutations at codon 634 (exon 11). ATA level B (ATA-B) mutations carry a lower risk for aggressive MTC mutations and include mutations at codons 609, 611, 618, 620 (exon 10), and 630 (exon 11). ATA level A (ATA-A) mutations carry the “least high” risk. Compared to ATA-B mutation carriers of the same age, these patients have lower serum calcitonin levels, lower tumor stage, and a higher rate of biochemical cure when they undergo prophylactic thyroidectomy at age  $\geq 4$  years (Frank-Raue et al., 2006). ATA-A mutations include *RET* mutations at codons 768, 790, 791 (exon 13), 804 (exon 14), and 891 (exon 15). (Table2) (Kloos et al., 2009).

ATA chose to create specific MTC Clinical Guidelines that would bring together and update the diverse MTC literature and combine it with evidence-based medicine and input from a panel of expert clinicians (Figure 1-5) (Kloos et al., 2009).

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