ALEX1 Regulates Proliferation and Apoptosis in Breast Cancer Cells

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Abstract

Background: Arm protein lost in epithelial cancers, on chromosome X (ALEX) is a novel subgroup within the armadillo (ARM) family, which has one or two ARM repeat domains as opposed to more than six-thirteen repeats in the classical Armadillo family members. Materials and Methods: In the study, we explore the biological functions of ALEX1 in breast cancer cells. Overexpression of ALEX1 and silencing of ALEX1 were performed with SK-BR3 and MCF-7 cell lines. Cell proliferation and colony formation assays, along with flow cytometry, were carried out to evaluate the roles of ALEX1. Results: ALEX1 overexpression in SK-BR3 breast cancer cells inhibited proliferation and induced apoptosis. Furthermore, depletion of ALEX1 in MCF-7 breast cancer cells increased proliferation and inhibited apoptosis. Additional analyses demonstrated that the overexpression of ALEX1 activated the intrinsic apoptosis cascades through up-regulating the expression of Bax, cytosol cytochrome c, active caspase-9 and active caspase-3 and down-regulating the levels of Bcl-2 and mitochondria cytochrome c. Simultaneously, silencing of ALEX1 inhibited intrinsic apoptosis cascades through down-regulating the expression of Bax, cytosol cytochrome c, active caspase-9, and active caspase-3 and up-regulating the level of Bcl-2 and mitochondria cytochrome c. Conclusions: Our data suggest that ALEX1 as a crucial tumor suppressor gene has been involved in cell proliferation and apoptosis in breast cancer, which may serve as a novel candidate therapeutic target.

Keywords: Breast cancer - ALEX1 - cell proliferation - apoptosis - active caspase-3

Introduction

Cancer is a global problem that accounts for almost 13% of deaths worldwide, there will be between 15 and 17 million new cases of cancer every year, 60% of which will be in developing countries (Lopez-Gomez et al., 2013). Breast cancer is one of the most prevalent cancers in women worldwide, accounting for 23% of the total cancer cases. It is a major cause of morbidity and mortality among women accounting for 14% of all cancer deaths. Each year, more than one million newly diagnosed cases worldwide appeared and brought serious harm to women health (Jemal et al., 2011). Surgery together with chemotherapy is the main method for breast cancer treatment but brings serious side effects to the patients. Therefore, new diagnostic and treatment strategies with low toxicity and high efficiency for breast cancer are shortly needed.

Arm protein lost in epithelial cancers, on chromosome X (ALEX; also known as armadillo repeat containing, X-linked; ARMCX), characterized by the presence of a repeating 42 amino acid motif (arm repeat) is a novel subgroup within the ARM family. The ALEX gene family consists of at least three variants (ALEX1, ALEX2, and ALEX3), which have one or two ARM repeat domains. Though the classical ARM family was involved in a variety of processes such as cell adhesion, embryogenesis and tumorigenesis (Peifer et al., 1994; Hatzfeld, 1999), little is known about the ALEX genes. Recent report demonstrates that the ALEX3 directly interacts with the sex determining region Y (Sry)-box 10 (SOX10) transcription factor via the ARM repeat domains and alters its subcellular localization and transcriptional activity (Mou et al., 2009). In addition, the study have shown that the ALEX3 gene encodes a mitochondrial-targeted protein and the non-canonical Wnt/PKC pathway regulates mitochondrial distribution through ALEX3 protein degradation (Serrat et al., 2013). gene expression analysis revealed that both ALEX1 and ALEX2 mRNA is expressed in a variety of adult human tissues, but dramatically reduced or even undetectable in several human carcinoma cell lines and tissues, including breast cancer (Kurochkin et al., 2001). Recent studies have shown that ALEX1 gene is transcriptionally regulated by CREB and Wntβ-catenin signaling and ALEX1 suppresses colony formation ability of human colorectal carcinoma cell lines (Iseki et al., 2010; 2012). Despite that,
the function of ALEX1 in cancer is still poorly understood, especially, in breast cancer.

With this goal, we examined the expression of ALEX1, cell proliferation and apoptosis of SK-BR-3 cells and MCF-7 cells after overexpression and silencing of ALEX1. The results showed that ALEX1 overexpression in breast cancer cells inhibited proliferation and induced apoptosis. However, depletion of ALEX1 increased proliferation and inhibited apoptosis.

Materials and Methods

Materials

SK-BR3 and MCF-7 cells were obtained from American Type of Culture Collection. Dulbecco’s modified Eagle’s medium (DMEM) was from Gibco. Lipofectamine 2000 and fetal bovine serum (FBS) were from Invitrogen. Primary and secondary antibodies for ALEX1, Bcl-2, Bax and β-actin for Western blot analysis were purchased from Santa Cruz Biotechnology and antibodies for active caspase-3, active caspase-9 and cytchrome c were obtained from Abcam Biotechnology. All the other chemicals and regents were obtained from local commercial sources.

Cell culture

SK-BR3, MDA-MB-231, T47D and MCF-7 cells were grown in DMEM supplemented with 10% FBS, 100 units/ml Penicillin and 100 μg/ml Streptomycin in a cell culture incubator at 37ºC with 5% CO2. MCF-10A were grown in DMEM/F12 supplemented with 5% Horse serum, 10μg/ml Insulin, 20ng/ml EGF, 100ng/ml Cholera Toxin, 0.5μg/ml Hydrocortisone, 100 units/ml Penicillin and 100 μg/ml Streptomycin in a cell culture incubator at 37ºC with 5% CO2.

siRNA sequences

The Promega software system was used to select siRNA against ALEX1 mRNA (NM_016608). the ALEX1 siRNAs and control siRNA were chemically synthesized by Shanghai Genepharma Co. Ltd. The sequences used for primers were as follows: RNAi-1: 5'-CCUGGAGCGGACAAAGUAATTT-3' (sense) and 5'-AUCAUUGUGUGCUCACGAGTT-3' (anti-sense); RNAi-2: 5'-GCCUGCUACUGUAAUACATT-3' (sense) and 5'-UGUAAACAGAUGCCAGCTT-3' (anti-sense); RNAi-3: 5'-GCUGGCGGCAAGCGUAAUATT-3' (sense) and 5'-UUAAACAGCUUAGCCACGCTT-3' (anti-sense) for three of ALEX1 siRNAs; and 5'-UUUCGGGACGCU GUCAGGUTT-3' (sense) and 5'-ACGUGACGCUUUCGGAGATT-3' (anti-sense) for control siRNA. All of the siRNA sequences were chemically synthesized by Shanghai Genepharma Co. Ltd.

Construction of recombinant lentiviral vector

The ALEX1 coding region fragment DNA was cloned into shuttle plasmid LV5 (Genepharma, China). The recombinant construct (LV5-ALEX1) was confirmed by sequencing. The recombinant construct and the negative control (LV5-NC) were transfected into 293T cells with three packaging components (pGag/Pol, pRev and pVSV-G) by Lipofectamine 2000 (Invitrogen, USA), respectively. The recombinant lentivirus expressing ALEX1 and the negative control lentivirus were obtained from the cells supernatant and named as LV5-ALEX1, LV5-NC, respectively.

Transient transfection of siRNA oligonucleotides

One day before transfection, cells were seeded at a density of 100,000 cells in 1.0 ml of growth medium without antibiotics in 6-well plates so that cells would grow to 50-60% confluence at the time of transfection. MCF-7 cells were transfected with ALEX1-siRNA (SiALEX1) or control siRNA (SiCon) for 48 h via Lipofectamine 2000. Two groups were set up: (i) The 90 nmol/l SiALEX1 group, (ii) The 90 nmol/l SiCon group.

Stable transfection of lentivirus LV5

SK-BR3 cells were infected with lentivirus LV5-ALEX1 or LV5-NC at an MOI (multiplicity of infection) of 5 for 72h. Cell line was divided into two experimental groups: (i) LV5-ALEX1 group, (ii) LV5-NC group.

RNA isolation and real-time PCR

Total RNA was isolated using E.Z.N.A. Total RNA Kit I (Omega, USA). cDNA was synthesized using PrimeScript® RT reagent Kit with gDNA Eraser (Takara, Japan). Real-time PCR was carried out by SYBR® Premix ExTaqTMII (Takara, Japan) and analyzed on MiniOpticon TM Real-Time PCR Detection System (Bio-Rad, USA). The sequences used for primers were as follows: 5'-TGATATCTGAGTGTCCCGACC-3' (sense) and 5'-TGTTTACCCAGAGTGACCAAGGCT-3' (antisense) for ALEX1; and 5'-TTTGGTGATCTGTGGAAGGAC-3' (sense) and 5'-GTAGGCGCAGTGAGGTCTTCT-3' (antisense) for GAPDH.

Cell proliferation assay

Cell proliferation was measured by a CCK8 kit (Beyotime, China) according to the manufacturer’s protocol.

Colony formation assay

SK-BR3 and MCF-7 cells were treated as described above. For each group of cells, 0.5x10^5 cells were plated in 12-well plates and incubated for 10 days. Medium was changed every 3 days. And then, colonies were fixed with 4% polyformaldehyde and stained with 0.2% crystal violet solution and counted.

Apoptosis analysis

SK-BR3 and MCF-7 cells were treated as described above. Two distinct methods were applied to get the convincing results of cell apoptosis. First, cell apoptosis was measured by fluorescence microscopy to identify apoptotic nuclear changes (cell shrinkage, chromatin condensation and fragmentation, nuclear membrane blebbing) after staining SK-BR3 and MCF-7 cells with Hoechst 33258. The percentage of apoptotic cells was determined with at least 100 cells per treatment group. Next, Cell apoptosis was evaluated by staining with AnnexinV-FITC/PI according to the protocol provided in the AnnexinV-FITC/PI apoptosis detection kit (ABGAB, USA) for 5 min at 25ºC, immediately analyzed by flow
cytometry, where apoptotic cells were defined as annexin V+ and PI-.

**Western blot**

SK-BR3 and MCF-7 cells were treated as described above. Total proteins from cells were extracted in RIPA Lysis Buffer (Beyotime, China) plus Phenylmethanesulfonyl fluoride. Mitochondrial and cytosolic proteins were isolated using Mitochondria/cytosol Fractionation Kit (Beyotime, China) following the manufacturer’s protocol. Proteins were probed with anti-ALEX1, anti-β-actin, anti-Bax, anti-Bcl-2 (Santa Cruz, USA) and anti-active caspase 3, anti-active caspase 9, anti-cytochrome c (Abcam, USA), 40 µg of protein were separated in 12% SDS-PAGE gels, transferred to nitrocellulosic membrane, and blocked with 5% nonfat milk in TBST on a shaker at room temperature for 1.5 h. The membranes were incubated with primary antibody on a shaker overnight at 4ºC. Goat anti-mouse and anti-rabbit, secondary antibodies conjugated to HRP (Santa Cruz, USA) were used to visualize the stained bands with an ECL kit (Santa Cruz, USA).

**Immunofluorescence**

Immunofluorescence staining with anti-ALEX1 antibody was performed in SK-BR3 cells. SK-BR3 cells were infected as described above and treated on coverslips. After fixation with 4% polyformaldehyde, cells were blocked in a buffer containing 10% goat serum and 1% BSA for 1h at room temperature and washed with PBS for three times; the slides were incubated with anti-ALEX1 antibody (1:50 dilution) overnight at 4ºC, then with Alexa Fluor 549-conjugated anti-mouse IgG (Invitrogen, USA) for 2h at 37ºC. After the final washes, DAPI (Sigma, USA) was added and used as counterstain for nuclei. Fluorescence images were acquired by an Olympus-BX51 microscope.

**Statistical analysis**

All statistical analyses were performed using the SPSS 17.0. data were represented as mean±SD. Student’s t test was used to compare data between control group and test group. p<0.05 was considered as statistically significant. Treatments of cells were analyzed in triplicate wells and the experiments were repeated three times.

**Results**

**Expression of ALEX1 in breast cancer cells**

To examine the expression of ALEX1 in breast cancer cells, we have examined the expression level of ALEX1 in five breast cell lines (MCF-10A, MCF-7, MDA-MB-231, T47D and SK-BR3) (Figure 1). Endogenous ALEX1 was underexpressed with different levels in all of these four breast cancer cell lines compared with MCF-10A, but relatively high expression of ALEX1 was observed in MCF-7 and low expression of ALEX1 in SK-BR3 cell lines by real-time PCR and Western blot.

**LV5-ALEX1 increases ALEX1 expression in SK-BR3 cells**

To explore the role of ALEX1 in breast cancer, we have overexpressed ALEX1 in SK-BR3 cells by recombinant lentiviral infection. SK-BR3 cells were treated as described above. The efficiency of infection was at least 90 % as indicated by the fluorescence intensity of GFP (Figure 2a). To further confirm the efficiency of ALEX1 overexpression, real-time PCR and immunofluorescence were performed. The result revealed that the level of ALEX1 mRNA in the LV5-ALEX1 group was significantly higher than that in the LV5-NC group (p<0.01, Figure 2b). Western blot showed the same result in the protein levels (Figure 2c). In LV5-ALEX1 cells, immunofluorescence staining with anti-ALEX1 antibody showed that ALEX1 protein was expressed in a higher...
SiALEX1 inhibits ALEX1 expression in MCF-7 cells

MCF-7 Cells were transfected with ALEX1 RNAi-1, ALEX1 RNAi-2, ALEX1 RNAi-3 and RNAi-con. After 48 h, ALEX1 mRNA and protein was examined by real-time PCR and Western blot. The real-time PCR data showed that the lowest expression of ALEX1 in MCF-7 cells treated with ALEX1 RNAi-2 among three siRNAs compared to RNAi-con group (Figure 3a). Western blot showed the same result in the protein levels (Figure 3b). Therefore, ALEX1 RNAi-2 (SiALEX1) was used to perform further tests.

Overexpression of ALEX1 inhibited proliferation and clonogenic ability of SK-BR3 cells and silencing of ALEX1 increased proliferation and clonogenic ability of MCF-7 cells

Cell proliferation assays were performed for SK-BR3 cells that were infected with either LV5-ALEX1 or LV5-NC. According to results from the CCK-8 assay, the absorbance of SK-BR-3 cells infected with LV5-ALEX1 was significantly decreased when compared with LV5-NC group at 48, 72 and 96h (p<0.05; Figure 4a). However, the absorbance of MCF-7 cells transfected with SiALEX1 was markedly increased , compared with SiCon group at 48, 72 and 96h (p<0.05; Figure 4b). In the colony formation assay, the results revealed that the overexpression of ALEX1 in SK-BR3 cell line significantly inhibited the number of foci, relative to LV5-NC group (p<0.01; Figure 5a, b). But, the number of foci in SiALEX1 group increasingly increased, compared to SiCon group in MCF-7 cells (P<0.05; Figure 5c, d).

Overexpression of ALEX1 induced SK-BR3 cells apoptosis and silencing of ALEX1 resisted MCF-7 cells apoptosis

We speculated that ALEX1 could induce breast cancer cells apoptosis. First, Hoechst 33258 staining was performed. The result exhibited typical apoptosis changes with chromatin condensation and fragmentation in LV5-ALEX1 group. However, the typical apoptosis changes were not found in SiALEX1 group of MCF-7 cells (Figure 6). To quantify the apoptotic cells with overexpression and silencing of ALEX1, SK-BR3 and MCF-7 cells were stained with Annexin-V/PI and analyzed by flow cytometry, which showed markedly increased apoptotic cells in ALEX1 overexpression cells, whereas the apoptotic rate was significantly less in ALEX1 silencing cells (p<0.05; Figure 7).
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Effects of ALEX1 on protein expression levels of intrinsic apoptosis pathway in SK-BR3 and MCF-7 cells

To determine whether breast cancer cells apoptosis is modulated by intrinsic apoptosis pathway, we analyzed the expression of Bax, Bcl-2, active caspase-9, active caspase-3 proteins with Western blot after overexpression or silencing of ALEX1. The results showed that overexpression of ALEX1 in SK-BR3 cells promoted the expression of Bax, active caspase-9, active caspase-3 and reduced Bcl-2 expressions (Figure 8a, b). However, silencing of ALEX1 in MCF-7 cells decreased the expression of Bax, active caspase-9, active caspase-3 and increased Bcl-2 expressions, β-actin was the internal control, suggesting that the protein loading amount was the same (Figure 8c, d). Collectively, these results suggest that ALEX1 induced apoptosis by causing to increase the ratio of Bax/Bcl-2 and then release cytochrome c from mitochondria to cytoplasm and activate caspase-9 and caspase-3, ultimate, triggered SK-BR3 cells apoptosis.

Discussion

The arm repeats were first found in the segment polarity gene armadillo in Drosophila in 1989 (Riggleman et al., 1989). Since then a number of proteins containing arm repeats have been noticed and studied. Studies have revealed that arm repeats regulate protein-protein interaction with multiple binding partners involved in nuclear transport, transcription activation and cell junction (Ozawa et al., 1995; Rubinfeld et al., 1995; Troyanovsky et al., 1996). Recently, the biological function of some classical arm repeat proteins have been well clarified. Such as β-catenin, a classical member of ARM family plays important roles both at cells adherence junctions and in Wnt signaling through interaction with E-cadherin and T cell factor/lymphoid enhancer factor (TCF/LEF) family transcription factors (Xu et al., 2007; Bass-Zubek et al., 2009). Tumor suppressor adenomatous polyposis coli (APC) is also an ARM family member and acts with casein kinase I, glycogen synthase kinase-3β and AXIN to regulate Wnt signaling through β-catenin degradation (Munemitsu et al., 1995; Rubinfeld et al., 1995).
ALEX1 overexpression increased the expression of Bax (Ding et al., 2015). Further studies by Western blot showed that ALEX1 overexpression reduced the protein level of cytochrome c in mitochondria and increased cytochrome c in cytosol. However, we came to the opposited conclusion in ALEX1 depletion (Figure 8). These results have suggested that ALEX1 promote the ratio of Bax/Bcl-2 and then result in cytochrome c release from mitochondria to cytosol. It is well known that caspase-8 and caspase-9 are essential proteases of extrinsic and intrinsic apoptotic pathways (Chen et al., 2014). Reports showed that the intrinsic apoptosis pathway is characterized as mitochondrial dysfunction and stimulation of caspase-9 and caspase-3 and caspase-3 is the cuultimate executionor for the nucleic changes with apoptosis (Chen et al., 2002). Therefore, mitochondrial damage is associated with the high level of caspase-9 and caspase-3. Our results displayed the higher level of active caspase-9 and active caspase-3 in LV5-ALEX1 group compared with in LV5-NC group (Figure 8), it is reasonable to infer that the elevated level of active caspase-3 is responsible for the increased apoptosis in SK-BR3 cells infected LV5-ALEX1. Taken together, ALEX1 overexpression induces cell apoptosis or ALEX1 silencing inhibits cell apoptosis in SK-BR3 or MCF-7 cells through intrinsic apoptosis depended on caspase activation.

In summary, we demonstrated that the overexpression of ALEX1 inhibited SK-BR3 breast cancer cells growth through upregulating the expression of apoptosis precursor protein Bax, downregulating anti-apoptotic protein Bcl-2 expression, increasing mitochondrial membrane permeability, triggering the caspase-3 cascade and inducing cell apoptosis. On the other hand, the silencing of ALEX1 promoted MCF-7 breast cancer cells growth by the same signal. Results in the study suggest that ALEX1 as a novel tumor suppressor gene could be involved in tumorigenesis and progression of human breast cancer and be utilized as a potential biomarker and therapeutic target in breast cancer. We will further study the molecular mechanisms of ALEX1 on apoptosis, migration and invasion of human breast cancer.

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References


