

## MINI-REVIEW

# Long Non-coding RNAs and Drug Resistance

Jing-Jing Pan<sup>1</sup>, Xiao-Juan Xie<sup>1,2</sup>, Xu Li<sup>3</sup>, Wei Chen<sup>1\*</sup>

### Abstract

**Background:** Long non-coding RNAs (lncRNAs) are emerging as key players in gene expression that govern cell developmental processes, and thus contributing to diseases, especially cancers. Many studies have suggested that aberrant expression of lncRNAs is responsible for drug resistance, a substantial obstacle for cancer therapy. Drug resistance not only results from individual variations in patients, but also from genetic and epigenetic differences in tumors. It is reported that drug resistance is tightly modulated by lncRNAs which change the stability and translation of mRNAs encoding factors involved in cell survival, proliferation, and drug metabolism. In this review, we summarize recent advances in research on lncRNAs associated with drug resistance and underlying molecular or cellular mechanisms, which may contribute helpful approaches for the development of new therapeutic strategies to overcome treatment failure.

**Keywords:** lncRNA - drug resistance - cancer

*Asian Pac J Cancer Prev*, 16 (18), 8067-8073

### Introduction

There are only 20,000-25,000 protein-coding genes in the entire human genome (Costa FF, 2005; Djebali S, et al., 2012), only account for <2% of the whole human genome. However, the degree of complexity in human biological functions is more associated with non-coding RNAs (ncRNAs) (Frith et al., 2005; Taft et al., 2007), which have not significant open reading frame and are classified as small and long ones depend on the length of transcript size, those length >200 nt are long non-coding RNAs (lncRNAs) (Rinn et al., 2012). lncRNAs are a heterogeneous type of ncRNAs that include thousands of different species, with poorly conserved (Fatica et al., 2014). The number of lncRNAs in the genome is still debated, but it is estimated that there are >60,000 lncRNAs in the human genome (Derrien et al., 2012; Hangauer et al., 2013). Although lncRNAs are considered as transcriptional noises or experimental artifact at first (Werner et al., 2005), it has been known that lncRNAs play an essential role in biological pathways, including chromosome silencing and activation (Tian et al., 2010; Hung et al., 2011), genomic imprinting (Sleutels et al., 2002), differentiation (Kretz et al., 2013), acting as scaffolds for transcription factors and epigenetic modifiers (Brunner et al., 2012). lncRNAs themselves may also serve as precursors of microRNAs (miRNAs) encoding miRNAs, or function as miRNAs sponge and target certain mRNAs leading to RNA degradation or inhibit translation (Kallen et al., 2013). lncRNAs have been implicated in development events, therefore dysregulation of lncRNAs

links with cancer and lncRNAs may be potential biological markers of tumors (Shi et al., 2013).

Chemotherapy has been widely used in cancer treatment for a long time, it usually has an effect on cancer cells by interfering with the metabolism of nucleic acid, DNA replication and mitosis, synthesis of protein. Nevertheless, drug resistance, either de novo or acquired, is still a challenge for cancer therapy since it often cause therapeutic failure. The underlying mechanism of resistance to chemotherapeutic agents are not fully elucidated. With the development of new technologies in combination of bioinformatics, more and more genes related to drug resistance are discovered or predicted (Potti et al., 2006; Raguz et al., 2008; Crijns et al., 2009; Etemadmoghadam et al., 2009). Drug resistance results from diverse factors, including individual variations in patients, genetic and epigenetic changes within tumors (Roberti et al., 2006; Tan et al., 2010) such as mutations, translocations, deletions and amplifications of coding genes or promoter regions, gene rearrangement (Fojo T, 2007), alteration of tumor microenvironment and tumor stromal cell components (Bouzin et al., 2007), self-protection of cancer stem cells (Rosen et al., 2009), epithelial-mesenchymal transition (Han et al., 2014), energy metabolism and hypoxia (Broxterman et al., 1991; Robey et al., 2009; Ruan et al., 2009). In addition, other reasons like classical drug efflux (Gottesman et al., 2006), acceleration of drug metabolism and decreasing sensitivity to induction of apoptosis (Janne et al., 2009; Coley, 2010) also result in drug resistance.

Accumulating evidences have been revealed that

<sup>1</sup>Clinical Laboratory, the First Affiliated Hospital, School of Medicine, Xi'an Jiaotong University, <sup>2</sup>Shaanxi Center for Clinical Laboratory, Shaanxi Province People's Hospital, <sup>3</sup>Translational Medicine Center, the First Affiliated Hospital, School of Medicine, Xi'an Jiaotong University, Xi'an, China \*For correspondence: chenwei6311@163.com.

genetic or epigenetic alterations of some genes encoding membrane transporters, drug metabolizers, cell cycle regulators are responsible for development of drug resistance (Ganguly et al., 2011). MiRNAs and lncRNAs are the major regulatory non-coding RNAs that regulate gene expression in epigenetic or genetic levels. A number of contemporary studies have shown that altered miRNAs expression plays an important role in the chemoresistance of cancer cells by impairing cellular responses (Nagano et al., 2011; Wapinski et al., 2011). For instance, miR-340 is involved in cisplatin resistance of hepatocellular carcinoma cell lines due to regulating Nrf2-dependent antioxidant pathway (Shi et al., 2014). miR-130a plays an important role in gefitinib resistance in NSCLC by targeting Met (Zhou et al., 2014). The increasing prevalence of miRNA mediated drug resistance data has led to an increase in studies focused on targeting miRNAs as strategies for personal therapeutic intervention (Mishra, 2012). However, little remains known about the relationship between lncRNAs and drug resistance. Here, we review lncRNAs associated with drug resistance, then highlight describe the molecular mechanism of lncRNAs modulated drug resistance and evaluate effect of clinical treatment, which will allow us to identify new therapeutic targets, contributing to lncRNAs targeting therapy in the future.

## **LncRNAs Modulate Gene Expression**

It is uncovered that gene expression may be different dependent on the function of RNA molecules themselves as well as their interactions with DNA and/or proteins, especially emphasis long non-coding RNAs (Harries 2012; Ernst et al., 2013). Studies have revealed that lncRNAs have abilities of changing gene expression in response of various extracellular stimuli (Castelnuovo et al., 2013), which indicates that lncRNAs may adjust the timing of gene expression and there are fine-tuning steps in the regulation of gene expression at the epigenetic, transcriptional or post-transcriptional level (Wang et al., 2013). Long intergenic non-coding RNAs (lincRNAs) can interact with chromatin remodeling proteins to epigenetically silence genes in trans (Khalil et al., 2009; Tsai et al., 2010). LncRNAs such as X-inactive specific transcript (XIST) and homeobox A1 (HOXA1) have been shown to serve as molecular scaffolds for the recruitment of chromatin-modifying complexes, notably polycomb repressive complex 2 (PRC2) to alter gene expression in cis (Lee et al., 2013; Maamar et al., 2013). LncRNAs also bind specific gene clusters to prevent the binding of transcriptional activators (Chu et al., 2011). LncRNAs like enhancer RNAs (eRNAs) and lincRNA-p21 transcription may enhance expression of neighboring protein-coding genes through locally modulate chromatin structure or act as locus-restricted co-activators (Melo et al., 2013; Dimitrova et al., 2014). Similarly, transcription of lncRNAs near protein-coding loci can suppress transcription since the transcriptional machinery on lncRNA gene locus physically prevents binding to protein-coding genes (Malek et al., 2014). The post-transcriptional roles of lncRNAs including mRNA

splicing, editing, transport, translation and degradation. It is reported that lncRNA HOX antisense intergenic RNA (HOXAIR) has an influence on mRNA translation and is shown to induce ubiquitin-mediated proteolysis in a way of post-transcriptional. HOTAIR associates with certain E3 ubiquitin ligases containing RNA-binding domains such as Dzip3 and Mex3b, as well as with their respective ubiquitination substrates, Ataxin-1 and Snurportin-1. HOTAIR promotes the ubiquitination of Ataxin-1 by Dzip3 and Snurportin-1 by Mex3b respectively, then accelerating their degradation (John et al., 2012). These collective results suggest that lncRNAs may regulate many genes expression through interacting with DNA/RNA or proteins at different levels.

## **LncRNAs and Cell Death**

It is known that lncRNAs can promote or suppress cell death pathways through diverse molecular mechanisms including epigenetically silencing (Zheng et al., 2014), acting as sponge or precursors for miRNAs (Wang et al., 2014), activating or inhibiting signal molecules and interacting with proteins or nucleic acids (Wang, 2011) in various cell types. For example, recently one lncRNA named PANDAR (promoter of CDKN1A antisense DNA damage-activated RNA) is reported to interact with the transcription factor NF-YA to limit the expression of pro-apoptotic genes in lung cancer cell lines. The results demonstrate that overexpression of PANDAR leads to a loss of NF-YA binding at the promoter of Bcl-2 thus inhibiting Bcl-2 expression and promoting apoptosis (Han, 2015). In contrast, lncRNA urothelial cancer associated 1 (UCA1) may restrain apoptosis induced by Ets-2 knockdown in bladder cancer BLZ-211 cells. Knockdown of Ets-2 reduced expression of UCA1 which cause an increase of apoptosis via inactivating of Akt in BLZ-211 cells (Wu et al., 2013). Additionally, it is also reported that reduction of UCA1 may serve a pro-apoptotic role in cardiomyocyte partly through enhancing the protein level of p27 (Liu et al., 2015).

## **LncRNAs Link with Drug Resistance**

Numerous mechanisms can illustrate the molecular basis of drug resistance. Recent research demonstrate that there is a relationship between the drug-resistance phenotype and non-mutational regulation of gene expression. LncRNAs are the major modulators of non-mutational gene regulation. LncRNAs have been reported to be dynamically changed in response to various drugs, and could affect gene expression involved in cell cycle arrest, inhibition of apoptosis and DNA damage repair (Goldman 2003; d'Adda 2008; Lipovich et al., 2010). In this section, we will discuss current experimental findings showing molecular evidences of lncRNAs involved in the drug resistance rely on different drugs (Table 1).

### *Cisplatin Resistance and lncRNAs*

Cisplatin is one of the most common anticancer drugs by interfering with DNA synthesis to prevent cancer cells proliferation, and often result in drug resistance during

**Table 1. Effect up-/down-regulation of Different lncRNAs Expression on Sensitivity to Chemotherapy. The Table Presents the Findings Indicating a Negative Correlation between Expression of lncRNAs and Effect on Sensitivity to Chemotherapy (except AK126698, GAS5 and SnaR)**

lncRNAs	Up-/downregulation	Possible targets	Chemotherapeutic agent	Effect on sensitivity to chemotherapy	Cancer type	References
HOTAIR	↑	p21	Cisplatin	↓	Lung adenocarcinoma	Liu Z (2013)
AK126698	↓	Wnt/β-catenin	Cisplatin	↓	Non-small-cell lung cancer	Yang Y (2013)
UCA1	↑	Wnt6	Cisplatin	↓	Bladder cancer	Fan Y (2014)
H19	↑	MDR-1/P95	Doxorubicin	↓	Breast cancer, liver cancer	Doyle LA (1996), Tsang WP (2007)
ARA	↑	Cyclin B1, ToPo IIα, ACSL4, Bel-x1, Bax	Doxorubicin	↓	Breast cancer, liver cancer	Jiang M (2014)
CUDR	↑	Caspase-3	Doxorubici, etoposide	↓	Squamous carcinoma	Tsang WP (2007)
PANDA	↑	NF-YA	Doxorubicin	↓	Breast cancer	Sotillo E (2011), Hung T (2011)
GAS5	↓	cIAP2,SGK1	Glucocortico-id, docetaxel, nutlin-3a , mitoxantrone	↓	Prostate cancer, renal cell carcinoma	Kino T (2010), Qiao HP (2013), Pickard MR (2013)
PRNCR1	↑	DOTIL	Castration	↓	Prostate cancer	Yang L (2013), Pestell RG (2014)
ROR	↑	p53	Sorafenib, doxorubicin	↓	Hepatocellular cancers	Takahas-hi K (2014)
MRUL	↑	ABCBI	Doxorubicin, vincristine	↓	Gastric cancer	Wang Y (2014)
SnaR	↓	----	5-Fluorouril	↓	Colon cancer	Lee H(2014)

cancer therapy. HOTAIR as a well-documented long intervening non-coding RNA (lincRNA) that can regulate gene expression in cis, trans and epigenetically (Woo et al., 2007; Peschansky et al., 2014). It is reported that HOTAIR is linked with resistance of lung adenocarcinoma (LAD) cells to cisplatin. Expression of HOTAIR is highly upregulated in cisplatin-resistant cells and cisplatin-responding LAD tissues. Knockdown of HOTAIR will increase whereas overexpression of HOTAIR may decrease chemosensitivity of lung cancer resistant-cells to cisplatin. HOTAIR-mediated chemoresistance is correlated with enhancement of cell proliferation, inhibition of G0/G1 cell-cycle arrest and apoptosis via regulation of p21/WAF1/CIP1 (p21) expression which could mimic the effects of HOTAIR on cisplatin resistance within lung cancer cells. Moreover, HOTAIR can promote the resistance of LAD cells to cisplatin by targeting p21 in vivo. Dysregulation of HOTAIR in advanced LAD tissues may be associated with the response of patients to cisplatin-based chemotherapy (Liu et al., 2013). Consequently, it is possible to target HOTAIR for therapy of cisplatin-resistant LAD patients.

AK126698 as a new lncRNA (Ota et al., 2004) associated with cisplatin resistance in non-small-cell lung cancer cells. Expression of AK126698 is reduced in lung cancer cells with cisplatin resistance. Knockdown of AK126698 may decrease the vertebrate orthologs of naked cuticle family member NKD2 which can negatively regulate Wnt signaling pathway through binding to Dvl protein and enhance the accumulation and nuclear translocation of β-catenin, thus activating canonical Wnt/β-catenin pathway and significantly suppressing apoptosis induced by cisplatin in lung cancer cells. It is appeared that AK126698 promotes cisplatin resistance through targeting the Wnt pathway (Yang et al., 2013). However, more research are needed to elucidate whether there are other mechanisms by which AK126698 mediates within cisplatin resistance.

UCA1, a recently identified long non-coding RNA, whose expression is positively correlated with cell proliferation and migration in bladder cancer cells. It is shown that expression of UCA1 is upregulated in both cisplatin treatment patients with bladder cancer and cisplatin-resistant bladder cancer cells. UCA1 knockdown reduced the cell viability and wingless-type MMTV integration site family member 6 (Wnt6) expression levels, whereas overexpression of UCA1 strongly enhances the cell viability and expression of Wnt6 in human bladder cancer cell lines during cisplatin treatment, thus activating canonical Wnt signaling pathway. Moreover, inhibition of Wnt6 in UCA1-overexpressing cells can partially alleviate the increase of cell viability induced by UCA1 during cisplatin treatment. Expression of UCA1 and Wnt6 is also positively correlated in vivo. Therefore, UCA1 promotes cisplatin resistance by activating Wnt signaling pathway in a Wnt6-dependent manner (Fan et al., 2014), but how UCA1 intensifies Wnt6 expression will be investigated in future.

*Doxorubicin resistance and lncRNAs*

Doxorubicin is a kind of broad-spectrum anthracycline anticancer drug which has strong cytotoxic effect. It is

one of the most widely employed chemotherapy agents for cancers, like breast cancer and liver cancer (Sen GS, et al., 2011). LncRNA H19 has been proved function as an oncogene. It is shown that compared with parental human breast carcinoma cells or drug-sensitive revertant cells, H19 gene is overexpressed within multidrug resistance phenotype of cells, which are characterized by overexpression of a 95-kilodalton membrane glycoprotein (p95) that correlates with lower accumulation and retention of doxorubicin. Another human lung carcinoma with p95-overexpressing multidrug-resistant cell lines, also displays high level of H19 mRNA (Doyle LA, et al., 1996). Besides, H19 gene is considered to induce P-glycoprotein expression and multidrug resistance 1 (MDR1)-associated drug resistance by mediating MDR1 promoter methylation in liver cancer cells. It is observed that expression of H19 mRNA, MDR1 gene and its protein product P-glycoprotein are all upregulated, contrast to a decrease of doxorubicin accumulation level in doxorubicin-resistant cells. H19 knockdown markedly increases the percentage of MDR1 promoter methylation, then reduces MDR1 and P-glycoprotein expression, thus enhancing the cellular doxorubicin accumulation level and sensitizes doxorubicin toxicity in both parent sensitive and resistance cells (Tsang WP, et al., 2007). It provides a new strategy (anti-H19) to improve the efficacy of cancer chemotherapy.

ARA-Doxorubicin Resistance Associated-as a new specific differently expressed lncRNA is validated in doxorubicin resistant cells. Sustained doxorubicin treatment can significantly upregulate ARA expression in parent sensitive cell lines. Knockdown of ARA can reverse drug resistance, diminish the proliferation, induce G2/M arrest and cell death through decreasing the accumulation of Topo II $\alpha$  (topoisomerase II alpha), expression of cyclin B1 and ACSL4 (acyl-CoA synthetase 4), downregulation Bcl-xl and upregulation Bax in doxorubicin resistant cells. ARA also involves in other multiple signaling pathways that are crucial for drug resistance, including MAPK signaling pathway, metabolism pathways, cell cycle and cell adhesion-related biological pathways (Jiang et al., 2014). It suggests that ARA has important roles in doxorubicin resistance development.

The long non-coding RNA novel gene cancer up-regulated drug resistant (CUDR) induces drug resistance in cells through suppressing apoptosis induced by drugs. Expression of CUDR is significantly upregulated in a doxorubicin-resistant subline of human squamous carcinoma cells, which are more resistant to drug-induced apoptosis. Overexpression of CUDR can increase resistance to doxorubicin and etoposide, decrease drug-induced apoptosis as well as expression and activity of caspase-3, promote anchorage-independent growth in squamous carcinoma cells. The present results demonstrate that CUDR may regulate the drug sensitivity through caspase 3-dependent apoptosis (Tsang et al., 2007). Whether or not CUDR will modulate other genes in drug resistance would require detailed investigation.

The P21-associated ncRNA DNA damage Activated lncRNA PANDA is an evolutionarily conserved lncRNA located between the protein-coding CDKN1A gene and

lincRNA-p21. Expression of PANDA is elevated in a subset of breast cancer cells that contribute to doxorubicin resistance, a crucial component of breast cancer chemotherapy. PANDA exerts effect through physically interacting with the nuclear transcription factor NF-YA subunit to diminish p53 dependent pro-apoptotic effect. Moreover, depletion of PANDA in human fetal lung fibroblasts has been shown to increase cell death induced by doxorubicin then enhancing sensitivity to doxorubicin (Sotillo E, et al., 2011). However, more studies is needed to validate the function of PANDA in drug resistance.

#### *Hormone drugs and lncRNAs*

The growth arrest-specific 5 lncRNA GAS5 as a long non-coding RNA encoded through its exonic sequences, is usually accumulated in growth-arrested cells. GAS5 can resist migration and invasion as well as promote apoptosis in renal cell carcinoma (RCC) cells, a role as a tumor suppressor in RCC has also been demonstrated (Pickard et al., 2013; Qiao et al., 2013). Downregulation of GAS5 contributing to glucocorticoid resistance through alleviating sensitivity of tumor cells to apoptosis via suppressing glucocorticoid-mediated induction of several responsive genes like anti-apoptotic cellular inhibitor of apoptosis 2 (cIAP2) and serum/glucocorticoid-regulated kinase 1 (SGK1). GAS5 acting as a decoy competes with cIAP2/SGK1 glucocorticoid response elements (GREs) for binding to the DNA-binding domain of the glucocorticoid receptor (GR) to prevent cIAP2/SGK1 transcriptional activation (Kino et al., 2010). Whether GAS5 interacts with other chromatin components to stimulate transcription of glucocorticoid-responsive genes need to be examined.

The lncRNA prostate noncoding RNA 1 (PRNCR1, also known as PCAT8) and prostate cancer gene expression marker 1 (PCGEM1) are both highly expressed in aggressive prostate cancer and enhance proliferation of prostate cancer cells. Hyperactive androgen receptor (AR) activity is a crucial determinant of resistance to current castration therapies. Knockdown of PRNCR1 and PCGEM1 can lessen the activity of AR and viability of castration-resistant prostate cancer cells. Moreover, PRNCR1 increases activity of AR through binding to acetylation AR C-terminus motif which is correlated with disruptor of telomeric silencing (DOT1)-like, histone H3 methyltransferase (DOT1L) and its association with DOT1L is required for sequential recruitment of the PCGEM1 to DOT1L-mediated methylated AR N-terminus (Yang et al., 2013; Pestell et al., 2014). Decrease of PRNCR1 may be a therapy strategy in castration-resistant prostate cancer.

#### *Other drugs resistance and lncRNAs*

Sorafenib is a multikinase inhibitor that exerts anti-angiogenic and anticancer effect by inhibiting multiple growth factor pathways (Wilhelm et al., 2004). LncRNA ROR is a stress-responsive lincRNA enriched within extracellular vesicles and upregulated in human hepatocellular cancers (HCC) cells, may as a modulator of chemotherapeutic response. Sorafenib raises linc-ROR expression in tumor cells and extracellular vesicles,

whereas knockdown of linc-ROR may enhance sorafenib-induced apoptosis and cell toxicity through significantly increasing p53 activity that can decrease cell viability of HepG2 cells during doxorubicin (Takahashi et al., 2014). Hence, targeting linc-ROR may improve responses to conventional therapeutic agents that are used for treatment of HCC.

Multidrug resistance (MDR) is a common reason of chemotherapy failure in cancer treatment. The long non-coding RNA MDR-related and upregulated lincRNA (MRUL) is strongly upregulated in both doxorubicin/vincristine-resistant gastric cancer (GC) cells and GC tissues. MRUL is negatively correlated with growth inhibition rates of GC specimens treated with chemotherapy drugs in vitro and indicates a poor prognosis for GC patients. MRUL knockdown not only results in increased rates of cell death by enhancing doxorubicin accumulation and reducing the Bcl-2/Bax ratio, but also decreased ABCB1 mRNA levels via the enhancer-like role of MRUL, whereas reversal of MRUL sequence and alteration of position of MRUL inserts do not change transcriptional enhancement, indicating that MRUL might positively affect ABCB1 expression in an orientation- and position-independent way (Wang Y, et al., 2014). It indicates that MRUL depletion would be helpful in GC MDR patients.

5-Fluorouracil (5-FU) is a classical anti-metabolite drug which is widely used for cancer treatment through affecting nucleoside metabolism to cause cell death (Zhang N, et al., 2008). lincRNA SnaR is transcribed by RNA polymerase III that links with nuclear factor 90 (NF90). It is shown that SnaR is downregulated in 5-FU-resistant colon cancer cells. Loss of SnaR in 5-FU-resistant colon cancer cells leads to increase the cell viability, decrease Annexin V-positive (ANN+) apoptotic cells after 5-FU treatment, which suggests that SnaR as a negative modulator in development of 5-FU resistance in colon cancer (Lee et al., 2014). But it does not describe the detail genes or pathways which SnaR modulates in 5-FU resistance of colon cancer cells.

Docetaxel is the first agent approved for treating hormone refractory prostate cancer by inhibiting tubulin to prevent proliferation and divide of cancer cells. GAS5 is aberrantly expressed in breast cancer, head and neck squamous cell carcinoma (HNSCC) and glioblastoma multiforme. It is reported that overexpression of GAS5 can enhance prostate cancer cell death induced by chemotherapeutic drugs docetaxel, nutlin-3a and mitoxantrone, while cell death is attenuated by downregulation of GAS5 expression. It is indicated that cell death is strongly correlated with cellular GAS5 levels and GAS5 is sufficient to mediate this activity. Accordingly, abnormally low levels of GAS5 expression may lessen the effectiveness of chemotherapeutic agents (Pickard et al., 2013). Enhancement of GAS5 in combination with chemotherapeutic drugs may improve the efficacy of cancer therapies.

## LncRNAs and Therapy

As discussed above, these studies have documented

lncRNAs roles during drug resistance, meaning lncRNAs may be the major players in contributing to drug resistance through modulating gene expression or signal pathways. The knowledge of lncRNAs related to drug resistance would be helpful in clinical therapy. It seems that some lncRNAs can be used as tools to evaluate the effectiveness of anticancer treatment in term of the response of drugs to adjust their dosage. For example, lncRNA Xist could be used as a biomarker to predict histone deacetylase inhibitors (HDACi, abexinostat) treatment effect on the breast cancer stem cell (CSC). Compared with low-dose sensitive breast cell lines (BCLs), high-dose BCLs tend to be enriched in Xist. X chromosomes number correlates with drug response to abexinostat. Low-dose sensitive BCLs present essentially X chromosome mono- or disomy, whereas high-dose sensitive BCLs present X chromosome normo- or polysomy. Only the CSC population will be reduced by abexinostat treatment in patient-derived xenografts (PDXs) with low Xist expression and abexinostat treatment targets the CSC population in vivo is inversely correlated to Xist expression (Salvador et al., 2013).

In addition, these lncRNAs themselves may be attractive therapeutic agents, since lncRNAs are differently expressed in chemosensitive and chemoresistance tumor cells. Although the molecular mechanism of lncRNA function in drug resistance are not full elucidated, some lncRNAs with unique characteristics can be potential candidates of therapeutic intervention. For instance, HOTAIR regulates cisplatin resistance through modulation of apoptosis and cell cycle distribution by affecting p21 expression in human LAD cells. This raises the possibility that anti-HOTAIR may have potential therapeutic value for those cisplatin-resistant LAD patients (Liu Z, et al., 2013). Because of the lower expression and baseline level of CUDR in human normal tissues compared with other current cancer biomarkers (for example, CEA), the gene might be a potential biomarker for assessment of cancer therapeutic response. Nevertheless, it need more investigation to support the future use of CUDR in cancer therapy (Tsang et al., 2007). These studies provide theoretical basis for targeting lncRNAs participated in anticancer therapy.

## Conclusion

Despite the progression of knowledge on resistance mechanisms, there is still a considerable lack of understanding of the detailed mechanisms and intracellular pathways mediated by lncRNAs. In this review, we attempt to summarize the roles of lncRNAs in drug resistance. These results demonstrated that lncRNAs are differently expressed in sensitive and resistant cells and most of them positively correlates with chemoresistance, directly or indirectly modulate drugs efficacy via regulating gene expression. One lncRNA can regulate many drugs resistance, and one drug resistance can be modulated by many lncRNAs. Studies to better understand the molecular mechanisms of lncRNAs in drug resistance offers promise for the development of more effective cancer therapies.

## References

- Bouzin C, Feron O (2007). Targeting tumor stroma and exploiting mature tumor vasculature to improve anti-cancer drug delivery. *Drug Resist Updates*, **10**, 109-20.
- Broxterman HJ, Pinedo HM (1991). Energy metabolism in multidrug resistant tumor cells: a review. *J Cell Pharmacol*, **2**, 239-47.
- Brunner AL, Beck AH, Edris B, et al (2012). Transcriptional profiling of lncRNAs and novel transcribed regions across a diverse panel of archived human cancers. *Genome Biol*, **13**, R75.
- Castelnuovo M, Rahman S, Guffanti E, et al (2013). Bimodal expression of PHO84 is modulated by early termination of antisense transcription. *Nat Struct Mol Biol*, **20**, 851-8.
- Chu C, Qu K, Zhong FL, et al (2011). Genomic maps of long noncoding RNA occupancy reveal principles of RNA-chromatin interactions. *Mol Cell*, **44**, 667-78.
- Coley HM (2010). Overcoming multidrug resistance in cancer: Clinical studies of p-glycoprotein inhibitors. *Methods Mol Biol*, **596**, 341-58.
- Costa FF (2005). Non-coding RNAs: New players in eukaryotic biology. *Gene*, **357**, 83-94.
- Crijns APG, Fehrmann RS, de Jong S, et al (2009). Survival-related profile, pathways, and transcription factors in ovarian cancer. *PLOS Med*, **6**, 24.
- d'Adda di FF (2008). Living on a break: cellular senescence as a DNA-damage response. *Nat Rev Cancer*, **8**, 512-22.
- Derrien T, Johnson R, Bussotti G, et al (2012). The GENCODE v7 catalog of human long noncoding RNAs: Analysis of their gene structure, evolution, and expression. *Genome Res*, **22**, 1775-89.
- Dimitrova N, Zamudio JR, Jong RM, et al (2014). LincRNA-p21 activates p21 In cis to promote polycomb target gene expression and to enforce the G1/S checkpoint. *Mol Cell*, **54**, 777-90.
- Djebali S, Davis CA, Merkel A, et al (2012). Landscape of transcription in human cells. *Nature*, **489**, 101-8.
- Doyle LA, Yang W, Rishi AK, et al (1996). H19 gene overexpression in atypical multidrug-resistant cells associated with expression of a 95-kilodalton membrane glycoprotein. *Cancer Res*, **56**, 2904-7.
- Ernst C, Morton CC (2013). Identification and function of long non-coding RNA. *Front Cell Neurosci*, **7**, 168.
- Etemadmoghadam D, de Fazio A, Beroukhi R, et al (2009). Integrated genomewide DNA copy number and expression analysis identifies distinct mechanisms of primary chemoresistance in ovarian carcinomas. *Clin Cancer Res*, **15**, 1417-27.
- Fan Y, Shen B, Tan M, et al (2014). Long non-coding RNA UCA1 increases chemoresistance of bladder cancer cells by regulating Wnt signaling. *FEBS J*, **281**, 1750-8.
- Fatica A, Bozzoni I (2014). Long non-coding RNAs: new players in cell differentiation and development. *Nat Rev Genet*, **15**, 7-21.
- Fojo T (2007). Multiple paths to a drug resistance phenotype: Mutations, translocations, deletions and amplification of coding genes or promoter regions, epigenetic changes and microRNAs. *Drug Resist Updates*, **10**, 59-67.
- Frith MC, Pheasant M, Mattick JS (2005). The amazing complexity of the human transcriptome. *Eur J Hum Genet*, **13**, 894-7.
- Ganguly A, Banerjee K, Chakraborty P, et al (2011). Overcoming multidrug resistance (MDR) in cancer and by a quinoline derivative. *Biomed Pharmacother*, **65**, 387-94.
- Goldman B (2003). Multidrug resistance: can new drugs help chemotherapy score against cancer? *J Natl Cancer Inst*, **95**, 255-7.
- Gottesman MM, Ling V (2006). The molecular basis of multidrug resistance in cancer: the early years of P-glycoprotein research. *FEBS Lett*, **580**, 998-1009.
- Han L, Zhang EB, Yin DD, et al (2015). Low expression of long noncoding RNA PANDAR predicts a poor prognosis of non-small cell lung cancer and affects cell apoptosis by regulating Bcl-2. *Cell Death Dis*, **6**, 1665.
- Han RF, Ji X, Dong XG, et al (2014). An epigenetic mechanism underlying doxorubicin induced EMT in the human BGC-823 gastric cancer cell. *Asian Pac J Cancer Prev*, **15**, 4271-4.
- Hangauer MJ, Vaughn IW, McManus MT (2013). Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. *PLoS Genet*, **9**, 1003569.
- Harries LW (2012). Long non-coding RNAs and human disease. *Biochem Soc Trans*, **40**, 902-6.
- Hung T, Wang Y, Lin MF, et al (2011). Extensive and coordinated transcription of noncoding RNAs within cell-cycle promoters. *Nat Genet*, **43**, 621-9.
- Janne PA, Gray N, Settleman J (2009). Factors underlying sensitivity of cancers to small-molecule kinase inhibitors. *Nat Rev Drug Discov*, **8**, 709-23.
- Jiang M, Huang O, Xie Z, et al (2014). A novel long non-coding RNA-ARA: adriamycin resistance associated. *Biochem Pharmacol*, **87**, 254-83.
- Kallen AN, Zhou XB, Xu J, et al (2013). The imprinted H19 lncRNA antagonizes Let-7 MicroRNAs. *Mol Cell*, **52**, 101-12.
- Khalil AM, Guttman M, Huarte M, et al (2009). Many human large intergenic noncoding RNAs associate with chromatin-modifying complexes and affect gene expression. *Proc Natl Acad Sci USA*, **106**, 11667-72.
- Kino T, Hurt DE, Ichijo T, et al (2010). Noncoding RNA gas5 is a growth arrest-andstarvation-associated repressor of the glucocorticoid receptor. *Sci Signal*, **3**, 8.
- Kretz M, Siprashvili Z, Chu C, et al (2013). Control of somatic tissue differentiation by the long non-coding RNA TINCR. *Nature*, **493**, 231-5.
- Lee H, Kim C, Ku JL, et al (2014). A long non-coding RNA snaR contributes to 5-fluorouracil resistance in human colon cancer cells. *Mol Cells*, **37**, 540-6.
- Lee JT, Bartolomei MS (2013). X-Inactivation, Imprinting, and Long Noncoding RNAs in Health and Disease. *Cell*, **152**, 1308-23.
- Lipovich L, Johnson R, Lin CY (2010). MicroRNA underdogs in a microRNA world: evolutionary, regulatory, and biomedical significance of mammalian long non-protein-coding RNA. *Biochim Biophys Acta*, **1799**, 597-615.
- Liu YB, Zhou DL, Li GN, et al (2015). Long non coding RNA-UCA1 contributes to cardiomyocyte apoptosis by suppression of p27 expression. *Cell Physiol Biochem*, **35**, 1986-98.
- Liu Z, Sun M, Lu K, et al (2013). The long noncoding rna hotair contributes to cisplatin resistance of human lung adenocarcinoma cells via downregulation of p21WAF1/CIP1 expression. *PLoS One*, **8**, 77293.
- Maamar H, Cabili MN, Rinn J, et al (2013). Linc-HOXA1 is a noncoding RNA that represses Hoxa1 transcription in cis. *Genes Dev*, **27**, 1260-71.
- Malek E, Jagannathan S, Driscoll JJ (2014). Correlation of long non-coding RNA expression with metastasis, drug resistance and clinical outcome in cancer. *Oncotarget*, **5**, 8027-38.
- Melo CA, Léveillé N, Agami R et al (2013). eRNAs reach the heart of transcription. *Cell Res*, **23**, 1151-2.
- Mishra PJ (2012). The miRNA-drug resistance connection: a new era of personalized medicine using noncoding RNA

- begins. *Pharmacogenomics*, **13**, 1321-4.
- Nagano T, Fraser P (2011). No-nonsense functions for long noncoding RNAs. *Cell*, **145**, 178-81.
- Ota T, Suzuki Y, Nishikawa T, et al (2004). Complete sequencing and characterization of 21,243 full-length human cDNAs. *Nat Genet*, **36**, 40-5.
- Peschansky VJ, Wahlestedt C (2014). Non-coding RNAs as direct and indirect modulators of epigenetic regulation. *Epigenetics*, **9**, 3-12.
- Pestell RG, Yu Z (2014). Long and noncoding RNAs (lnc-RNAs) determine androgen receptor dependent gene expression in prostate cancer growth *in vivo*. *Asian J Androl*, **16**, 268-9.
- Pickard MR, Mourtada-Maarabouni M, Williams GT (2013). Long non-coding RNA GAS5 regulates apoptosis in prostate cancer cell lines. *Biochim Biophys Acta*, **1832**, 1613-23.
- Potti A, Dressman HK, Bild A, et al (2006). Genomic signatures to guide the use of chemotherapeutics. *Nat Med*, **12**, 1294-300.
- Qiao HP, Gao WS, Huo JX, et al (2013). Long non-coding RNA GAS5 functions as a tumor suppressor in renal cell carcinoma. *Asian Pac J Cancer Prev*, **14**, 1077-82.
- Raguz S, Yague E (2008). Resistance to chemotherapy: New treatments and novel insights into an old problem. *Brit J Cancer*, **99**, 387-91.
- Rinn JL, Chang HY (2012). Genome regulation by long non coding RNAs. *Annu Rev Biochem*, **81**, 145-66.
- Roberti A1, La Sala D, Cinti C (2006). Multiple genetic and epigenetic interacting mechanisms contribute to clonally selection of drug-resistant tumors: current views and new therapeutic prospective. *J Cell Physiol*, **207**, 571-81.
- Robey RB, Hay N (2009). Is Akt the "Warburg kinase"?-Akt-energy metabolism interactions and oncogenesis. *Semin Cancer Biol*, **19**, 25-31.
- Rosen JM, Jordan CT (2009). The increasing complexity of the cancer stem cell paradigm. *Science*, **324**, 1670-3.
- Ruan K, Song G, Ouyang G (2009). Role of hypoxia in the hallmarks of human cancer. *J Cell Biochem*, **107**, 1053-62.
- Salvador MA, Wicinski J, Cabaud O, et al (2013). The Histone Deacetylase Inhibitor Abexinostat Induces Cancer Stem Cells Differentiation in Breast Cancer with Low Xist Expression. *Clin Cancer Res*, **19**, 6520-31.
- Sen GS, Mohanty S, Hossain DM, et al (2011). Curcumin enhances the efficacy of chemotherapy by tailoring p65NFkB-p300 cross-talk in favor of p53-p300 in breast cancer. *J Biol Chem*, **286**, 42232-47.
- Shi L, Chen ZG, Wu LL, et al (2014). miR-340 reverses cisplatin resistance of hepatocellular carcinoma cell lines by targeting Nrf2-dependent antioxidant pathway. *Asian Pac J Cancer Prev*, **15**, 10439-44.
- Shi X, Sun M, Liu H, et al (2013). Long non-coding RNAs: A new frontier in the study of human diseases. *Cancer Lett*, **339**, 159-66.
- Sleutels F, Zwart R, Barlow DP (2002). The non-coding Air RNA is required for silencing autosomal imprinted genes. *Nature*, **415**, 810-3.
- Sotillo E, Thomas TA (2011). The long reach of noncoding RNAs. *Nat Gene*, **43**, 616-7.
- Taft RJ, Pheasant M, Mattick JS (2007). The relationship between non protein-coding DNA and eukaryotic complexity. *Bioessays*, **29**, 288-99.
- Takahashi K, Yan IK, Kogure T, et al (2014). Extracellular vesicle-mediated transfer of long non-coding RNA ROR modulates chemosensitivity in human hepatocellular cancer. *FEBS Open Bio*, **4**, 458-67.
- Tan DS, Gerlinger M, Teh BT, et al (2010). Anti-cancer drug resistance: Understanding the mechanisms Through the use of integrative genomics and functional RNA interference. *Eur J Cancer*, **46**, 2166-77.
- Tian D, Sun S, Lee JT (2010). The long noncoding RNA, Jpx, is a molecular switch for X chromosome inactivation. *Cell*, **143**, 390-403.
- Tsai MC, Manor O, Wan Y, et al (2010). Long noncoding RNA as modular scaffold of histone modification complexes. *Science*, **329**, 689-93.
- Tsang WP, Kwok TT (2007). Riboregulator H19 induction of MDR1-associated drug resistance in human hepatocellular carcinoma cells. *Oncogene*, **26**, 4877-81.
- Tsang WP, Wong TW, Cheung AH, et al (2007). Induction of drug resistance and transformation in human cancer cells by the noncoding RNA CUDR. *RNA*, **13**, 890-8.
- Wang K, Long B, Zhou LY, et al (2014). CARL lncRNA inhibits anoxia-induced mitochondrial fission and apoptosis in cardiomyocytes by impairing miR-539-dependent PHB2 downregulation. *Nat Commun*, **5**, 3596.
- Wang KC, Chang HY (2011). Molecular mechanisms of long noncoding RNAs. *Mol Cell*, **43**, 904-14.
- Wang S, Tran EJ (2013). Unexpected functions of lncRNAs in gene regulation. *Commun Integr Biol*, **6**, 27610.
- Wang Y, Zhang D, Wu K, et al (2014). Long non-coding RNA MRUL promotes ABCB1 expression in multidrug-resistant gastric cancer cell sublines. *Mol Cell Biol*, **34**, 3182-93.
- Wapinski O, Chang HY (2011). Long noncoding RNAs and human disease. *Trends Cell Biol*, **21**, 354-61.
- Werner A, Berald A (2005). Natural antisense transcripts: sound or silence? *Physiol Genomics*, **23**, 125-31.
- Wilhelm SM, Carter C, Tang L, et al (2004). BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res*, **64**, 7099-109.
- Woo CJ, Kingston RE (2007). HOTAIR lifts noncoding RNAs to new levels. *Cell*, **129**, 1257-9.
- Wu W, Zhang S, Li X, et al (2013). Ets-2 regulates cell apoptosis via the akt pathway, through the regulation of Urothelial Cancer Associated1, a Long Non-Coding RNA, in Bladder cancer cells. *PLoS One*, **8**, 73920.
- Yang L, Lin C, Jin C, et al (2013). lncRNA-dependent mechanisms of androgen-receptor-regulated gene activation programs. *Nature*, **500**, 598-602.
- Yang Y, Li H, Hou S, et al (2013). The noncoding RNA expression profile and the Effect of lncRNA AK126698 on cisplatin resistance in non-small-cell lung cancer cell. *PLoS One*, **8**, 65309.
- Zhang N, Yin Y, Xu SJ, et al (2008). 5-Fluorouracil: mechanisms of resistance and reversal strategies. *Molecules*, **13**, 1551-69.
- Zheng H, Yang S, Yang Y, et al (2014). Epigenetically silenced long noncoding-SRHC promotes proliferation of hepatocellular carcinoma. *J Cancer Res Clin Oncol*, **16**, 171-5216.
- Zhou YM, Liu J, Sun W (2014). MiR-130a overcomes gefitinib resistance by targeting met in non-small cell lung cancer cell lines. *Asian Pac J Cancer Prev*, **15**, 1391-6.