

Evaluation of the Role of Merromoside from *Ipomoea aquatica* Forsskal Hydroalcoholic Extract in the Downregulation of ROS Species in Overcoming MDR in Breast Cancer

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Abstract

Purpose: The medicinal plant *Ipomoea aquatica* belonging to *convulvulaceae* family is an effective natural herb for treatment of various ailments and possesses effective anticancer activity. The aim of the work is to characterize a secondary metabolite merromoside (a resin glycoside) for anti-breast cancer activity through down regulation of ROS species. **Methods:** The Extract of the whole plant has been prepared by maceration method using 50%v/v ethanol in distilled water to get a hydroalcoholic extract. The phytochemical evaluation reveals that the active secondary metabolite was isolated by using column chromatographic technique. The isolated compound was evaluated for its anticancer properties through invitro method such as 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide assay on Michigan Cancer Foundation-7 Cell lines. The purity and structural characterization were done by high-performance thin layer chromatography, Fourier Transform Infrared Spectroscopy, Proton and ¹³C Nuclear Magnetic Resonance spectroscopy and Liquid Chromatography–Mass Spectrometry. **Results:** The isolated compound (W04) from the derived extract showed Rf value of 0.79 that showed IC₅₀ of 182.8µg/ml. The chemical structure of W04 has been confirmed as [4,5-dihydroxy-6-[5-hydroxy-2-methyl-4-(2-methylpropanooyloxy)-6-[(24,25,26-trihydroxy-5,23-dimethyl-9-oxo-19-pentyl-2,4,8,20,22-pentaoxatricyclo[19.2.2.13,7]hexacosan-6-yl)oxy]oxan-3-yl]oxy-2-methyloxan-3-yl] 2-methyl propanoate with the molecular weight of 979.15268. The isolated compound merromoside from hydroalcoholic extract of *Ipomoea aquatica* has been evaluated for anti-breast cancer properties. The down regulation of ROS species will prevent reverse signalling and angiogenesis. This indicates that merromoside will overcome MDR in breast cancer especially DOX-resistant.

Keywords: Merromoside- resin glycoside- breast cancer- MCF-7 cell lines

Asian Pac J Cancer Prev, 23 (11), 3657-3663

Introduction

Plants are rich sources of medicinal compounds that provide preventive and curative therapy than synthetic drugs. Hence, there is an increased focus towards assembling and contributing herbal remedy for meeting the global exigency through drug development and research (Apu et al., 2012). The International Agency for Research on Cancer (IARC) estimates that globally, 1 in 5 people develop cancer during their lifetime, and 1 in 8 men and 1 in 11 women die from the disease. Breast cancer represents 1 in 4 cancers diagnosed among women globally. Colorectal, lung, cervical, and thyroid cancers are also common among women (MamtaSaxena et al., 2013; Marvibaigi et al., 2016). As per the report on the plant, *Convolvulaceae* plants would have resin glycosides like aquaterins, convolvulins and jalapins (Ono, 2017). Aquaterins (I to XIX) present in the *I. aquatica* plant had evaluated for aquaterin II induced G0/G1 arrest regulated by related proteins CDK4/6, cyclin

D/E and p21 used mitochondria-mediated apoptosis featured by MMP decrease, ROS accumulation, caspase cascade activation and Bax/Bcl-2 alteration (Bo-Yi Fan et al., 2015). Reversal of multidrug resistance (MDR) by thirty resin glycosides from the morning glory family (*Convolvulaceae*) was evaluated in vinblastine-resistant human breast carcinoma cells (MCF-7/Vin) (Figueroa-González et al., 2012). The mechanism of resin glycosides in improving the MDR reversal activity was revealed through a structure–activity relationship study that substituting Rha" C-3 with a trans-cinnamoyl group (Li et al., 2017). *I. aquatica* was screened for its activity against fibroblast cell lysis after *heterometrus laoticus* scorpion venom treatment but the result was clearly negative. The purified compound from the methanolic extract of *I. aquatica* showed cytotoxicity towards cell cultures with CTC50 values of 387 mg/ml against normal Vero cell line, (normal African green monkey kidney) and HEP-2 (human larynx epithelial carcinoma) cell and A-549 (human small cell lung carcinoma). The crude extract was

more potent than that of the purified compound probably due to the combination of anthocyanin and other phenolic compounds. Isolation of a free radical-scavenging antioxidant from water spinach (*Ipomoea aquatica* Forsk) was reported (Parimala K et al., 2013). Anticancer activity of *Ipomoea aquatica*, a common leafy vegetable in prostate cancer and reduction of cisplatin-induced hepatotoxicity in vivo were reported. From the above literature information, it is clear that *I. aquatica* plant extract has potential as anti-tumor active while its effect has not been explored with breast cancer cell lines. Resin glycosides, as members of glycolipids have shown antitumor activity. The activities include mammalian cytotoxicity of tricolorin, antimetastatic activity of cairicoside, HepG2 inhibition by aquaterin-II (Chen et al., 2017; Fan BY et al., 2015; Pereda-Miranda et al., 2010; Rencurosi et al., 2004). Dichondrins, resin glycosides isolated from *Dichondra repens* were shown to reverse MDR and increase the cytotoxicity of vincristine (Song et al., 2015). In radiotherapy, blockage of HIF1 response has been shown to minimize lactate levels that worked as an aid to suppress anaerobic metabolism resulting in improved efficacy (Reuter et al., 2010). The role of resin glycosides in inhibiting P-gP has been reported by the use of resin glycosides from *Merremiamammosa* (Lour.) Hallier. F (Convolvulaceae) (Kim HR et al., 2004; Kitagawa I et al., 1997; Kokate CK et al., 2015; Lande et al., 2015). Efflux pump inhibitory ability of resin glycosides towards combating drug resistance in combinatorial therapy has also been reported that indicate the potential of the glycolipids in cancer treatment (Corona-Castañeda et al., 2016). Hence, the research was focused on exploring the anti-breast cancer potential of isolated compound from the opted plant extract for anti-breast cancer activity *Ipomoea aquatica* (Hu et al., 2014).

Materials and Methods

Materials

Plant Sample

The common name of the plant is Water Spinach, River Spinach. The botanical name is identified and authenticated as *Ipomoea aquatica* Forsskal belonging to the family Convolvulaceae. The plant was collected from the location Parambikulam – Aliyar Riverine, Pollachi. The part of the plant used for the study was whole plant. The plant was authenticated at BSI, Coimbatore-641003, Tamil Nadu, India, with the authentication number of BSI/SRC/5/23/2017/Tech./3269.

Chemicals/Reagents used

The chemicals and reagents used for the study were Petroleum Benzene boiling range 60.0°C-80.0°C GR, (Merck Specialities Private Limited, Mumbai – 400 018), Pyridine GR, Trichloro acetic acid, Sodium nitroprusside dehydrate purified, Silica gel G 60 – 120 mesh for column chromatography, o-toluidine, (Nice chemicals, Kochi – 682 024, Barfoed's reagent, (Oxford Laboratory, Mumbai – 400 002), Ethanol AR 99.9%, (Jiangsu Huaxi International Trade Co., Ltd.China), Zinc (metal) Powder, Laboratory Rasayan, Gelatin, Extra pure, (Hi Media Laboratories

Private Limited, Mumbai – 400 086), Potassium dichromate, Qualigens Fine Chemicals, Mumbai – 400 030. Sulphur powder, Distilled water, Seliwanoff's reagent LR, (Reachem Laboratory chemicals Private Limited, Chennai – 600 098), Molisch's reagent LR, 1-naphthol LR, Dimethyl sulphoxide LR, Toluene (Sulphur free), Formic acid LR, Phenyl hydrazine hydrochloride LR, Sodium oxalate LR, Dimethyl formamide LR, Ammonia solution LR, Benedict's quantitative reagent LR, Barium chloride LR, Fehling Solution A LR, (Chemspure, Chennai – 600 098), Acetic acid Glacial LR, Acetone LR, Biuret reagent, (LobaChemie Private Limited, Mumbai – 400 005), Mayer's solution, Fehling Solution No.2, Bial's reagent, Picric acid, Ferric chloride, Ammonium oxalate LR, Methanol LR, (S d Fine Chemicals Limited, Mumbai – 400 030), Potassium iodide, Oxalic acid, Sodium hydroxide pellets purified, Silica gel G for TLC, (LobaChemie Private Limited, Mumbai – 400 005), Potassium bromide for IR, MTT Powder, DMSO, Cell lines, ATCC, DMEM/F12, (Invitrogen), Fetal Bovine Serum, Penicillin, Streptomycin, Trypsin, EDTA, Glucose in PBS and DMBA, (Sigma Chemicals Co. St. Louis, MO, USA).

Instruments used

The analytical instruments utilized for the research were Precision Balance (Wensar), Hot plate (Cintex), Ultra Sonicator (Labman), Electrical Water bath (Technico), UV cabinet (CAMAG and Deep Vision), FTIR (Shimadzu), LCMS (PE Sciex API3000), FT-NMR (BRUKER AV400), Double beam UV/Visible spectrophotometer (Shimadzu UV 1800), CO₂ incubator (Thermo Fischer Scientific) and Microplate reader (Tecan).

Methods

Collection and Extraction

Maceration – a conventional extraction

In this process, the whole or coarsely powdered crude drug was placed in a stoppered container with the solvent and allowed to stand at room temperature for a period of at least 3 days with frequent agitation until the soluble matter has dissolved. The mixture then was strained, the marc (the damp solid material) was pressed, and the combined liquids were clarified by filtration or decantation after standing (Apu et al., 2012).

Preliminary Phytochemical Screening

Phytochemical process of the hydroalcoholic extract was carried out to detect the presence of secondary metabolite such as flavonoids, tannins, terpenoids, steroids, alkaloids, saponins and reducing sugars using standard phytochemical methods. Medicinal properties of the plants are determined by their phytochemical constituents (JB, 1973; Khadabadi SS et al., 2013; Shaikh., 2020).

Isolation and purification of bioactive compounds from plant samples

Completely dried plant extract sample should be mixed with silica gel to make a fine powdered form for easy distribution of sample in already packed silica gel

column. Sample powdered mass should be placed on the top of the pre-packed silica column and sample should be covered with a layer of cotton. The solvents of different polarities were passed through column at uniform rate under gravity to fractionate the sample extract (Bajpai et al., 2016; Corona-Castañeda et al., 2016; JB, 1973; Lande et al., 2015). Each fraction was collected separately in a test tube and numbered consecutively for further analysis on thin layer chromatography. The visualized spots of the components in the chromatogram are marked and the R_f value of each spot is calculated by the formula: R_f = distance travelled by the sample (cm)/ distance travelled by the solvent (cm) (Kokate CK et al., 2015). Based on the nature of the compounds, further spectral analyses such as infrared (IR), mass spectrometry and nuclear magnetic resonance (NMR) can be performed to elucidate the chemical structure of target compounds (L et al., 2012).

Anticancer Evaluation

Determination of effect of plant extract on cell proliferation by MTT assay

Cell culture and drug treatment

Human breast carcinoma cell lines MCF-7 and MDA-MB-231 were obtained from ATCC through NCCS, Pune, India. The cells were cultured and maintained in Modified Eagle's Medium (MEM) and Dulbecco's Modified Eagle's Medium (DMEM), respectively, supplemented with 10% (v/v) Fetal Bovine Serum (FBS) and 100 IU/mL Penicillin and 0.1 mg/mL streptomycin in a humidified atmosphere of 5% CO₂ at 37°C. Stock solutions of AZA (Sigma), TSA (Sigma), and SFN (Sigma) were prepared in dimethyl-sulphoxide (DMSO) whereas SAM (Sigma) was dissolved in milli-Q water. Stock solutions were further diluted to working concentrations in DMEM prior to use. Cells were harvested by trypsinization and cell number was counted by hemocytometer. The number of living cells was calculated by Trypan blue staining (0.2% v/v). For determining the concentration of drug that inhibited cell proliferation by 50% (IC₅₀), 5 X 10³ cells per well were seeded in a 96-well micro-titer plates and after 24 h incubation, were treated with the epigenetic modulators at different concentrations (AZA, SAM, SFN (1, 5, 10, 15, 20, 50 μM) and TSA (10, 20, 50, 100, 250, 500 nM)) mixed in respective medias supplemented with 5% FBS. Control cells were treated with DMSO only (Grigalius et al., 2017; JB, 1973; Juan-García et al., 2015; Raj et al., 2014).

Cell viability analysis by colorimetric MTT assay

The effect of the epigenetic modulators, DNMT and HDAC inhibitors on cellular proliferation was assessed by 3-(4, 5-Dimethylthiazol-2-yl)-2, 5-Diphenyltetrazolium Bromide (MTT) assay, using standard protocol. Briefly, the drug-treated cells in each

of the 96 wells were washed twice with PBS. 0.8 mg/mL MTT solution was prepared from stock MTT solution (5 mg/mL PBS, pH 7.2). A total of 100 μL MTT solution was added to each well and incubated at 37°C for 4 h in dark. The supernatant was re-moved and 100 μL of DMSO was added into each well to dissolve the formazan crystals. The absorbance was measured at 570 nm and results were expressed as the mean of three replicates as a percentage of control (taken as 100%). The extent of cytotoxicity was defined as the relative reduction of the optical density (OD), which correlated to the amount of viable cells in relation to cell control (100%). The cell viability was plotted in a graph and the IC₅₀ was calculated accordingly to decide the optimum dosage of the drugs for further studies (Liaudanskas et al., 2021; Vijayarathna et al., 2012).

Statistical analysis

All data are presented as means ± SD. Statistical analysis was performed using the Student's t-test by SPSS software. Values of P < 0.05 were considered as significant value.

Structural characterization

FTIR (KBr Pressed Pellet Technique)

Sample is mixed with potassium bromide (0.1% w/v) and compressed into a thin transparent pellet using a hydraulic press at 3,000 psi or 200 bar pressure. The scanned spectra are meant for interpretation (Tossaton Charoonratana et al., 2014; YR, 2013).

Nuclear Magnetic Resonance Spectroscopic Analysis

D₂O was used as solvent for NMR analysis using both ¹H and ¹³C (Tossaton Charoonratana et al., 2014).

Spectroscopic Parameters

Mass Spectroscopic Analysis

Liquid chromatography and mass spectroscopic analysis using electron spray ionization method was used and the spectra for two compounds were evaluated for the various ion peaks to get complete information about chemical structure (Song et al., 2015; Willard et al., 1986)

Results

The cold maceration was conducted with hydroalcoholic menstruum for 7 days taking 350.0g of drug and 2000.0ml of menstruum. The percentage yield of hydroalcoholic extract was found to be 28.91 % w/w.

From the phytochemical tests, it was found that the primary metabolites such as carbohydrates, proteins & amino acids and the secondary metabolites like alkaloids, glycosides, flavonoids, tannins & triterpenoids were present in the prepared extract.

FTIR experiments for W04 as O-H Alcohols (3545

Table 1. HPTLC Analysis of Isolated Compound W04

Sample	Concentration	At 254 nm		At 366 nm	
		R _f value	Peak Area	R _f value	Peak Area
W04	1.0 μg/ spot	0.79	2305.6	0.79	2647.4

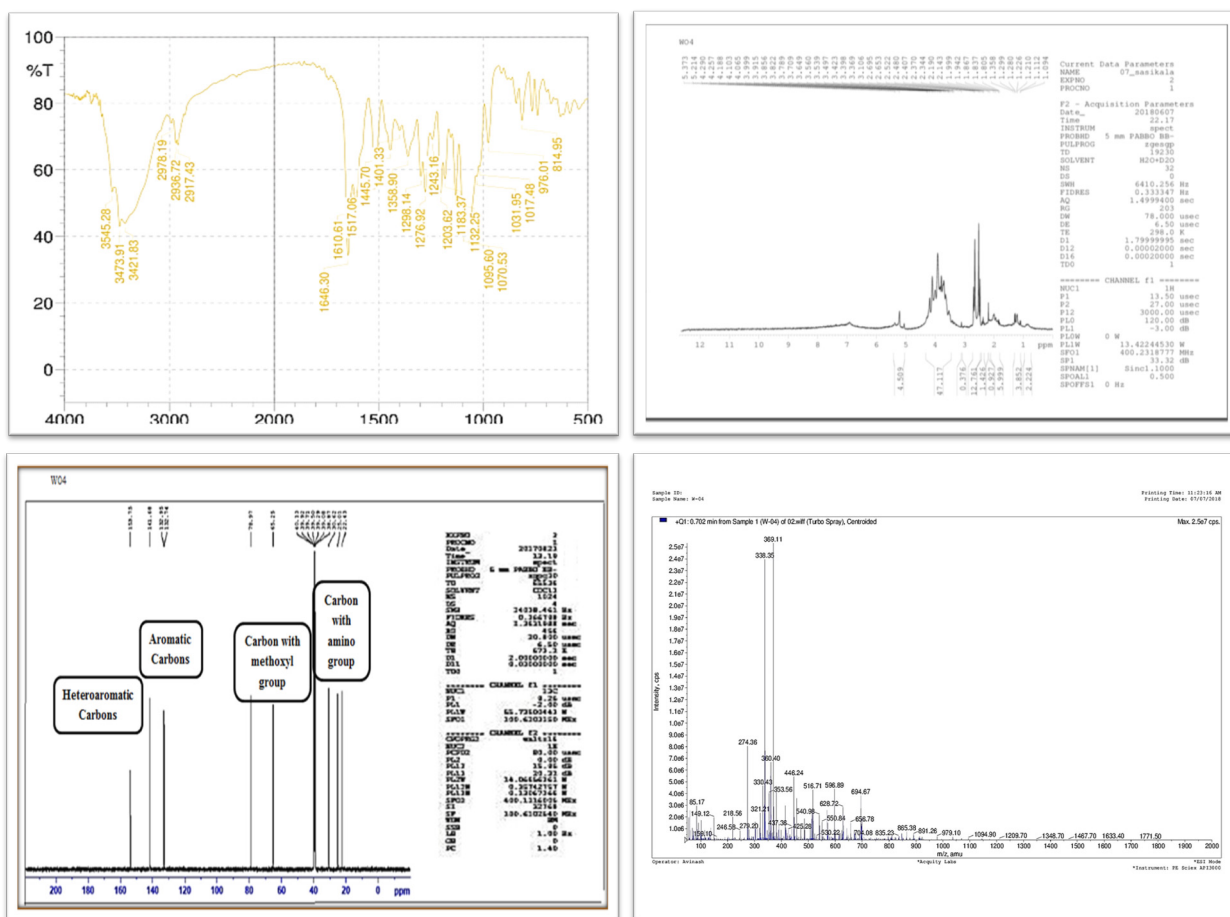


Figure 1. FTIR, 1H NMR, 13C NMR and LC/MS Spectral Data of Isolated Compound W04 Separately

& 3473 cm⁻¹); C-H Alkanes (2936 cm⁻¹); C=C Alkenes (1646 cm⁻¹); C=O Aromatics (1610 cm⁻¹) and C-C, C-O

Primary/Secondary Alcohols (1070 cm⁻¹). FT – NMR spectral studies for proton NMR spectra of compounds

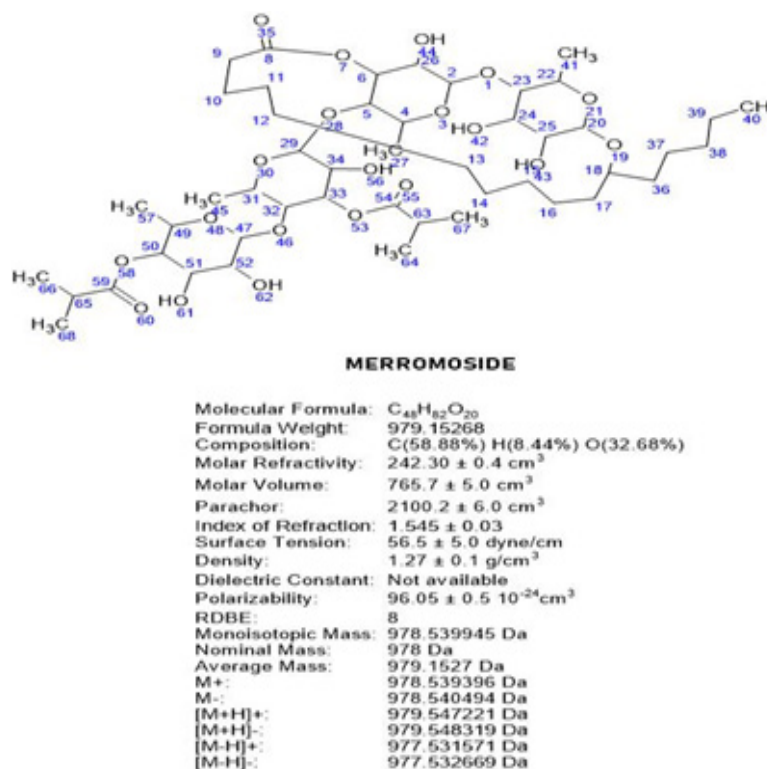


Figure 2. Possible Predictable Structure for W04

Table 2. Evaluation of IC₅₀ Values of Samples

Compound name	Conc. (µg/ml)	MCF-7		
		OD at 590nm	% Inhibition	IC ₅₀ (µg/ml)
Control	0	0.923	0	220.9
Extract	10	0.884	4.23	
	20	0.835	9.53	
	40	0.779	15.6	
	80	0.685	25.79	
	160	0.594	35.64	
	320	0.432	53.23	
	320	0.448	51.44	
W04	10	0.885	4.12	182.8
	20	0.855	7.37	
	40	0.779	15.56	
	80	0.68	26.34	
	160	0.549	40.5	
	320	0.408	55.82	

were concluding the expected set of protons with active functional groups. The ¹H spectrum resulted for W04 existed with 25-CH₃ (δ-0.927), 5-CH₃ -sugar (δ-1.426), 11-CH₂ (δ-2.224), 5-H-sugar (δ-3.852), 4-H-sugar (δ-4.509), 3-H-sugar (δ-5.999), 6-OH (δ-2.522 & 3.560), 9-H (δ-2.653) and 7-H (δ-2.675). FT – NMR spectral studies for ¹³C NMR spectra of compounds were finalizing the predictable set of carbons with active functional groups. The spectrum gives rise to following groups for W04, C-6 C=O (δ-153.75), C-8 C=O (δ-143.49), C-1 (δ-132.95 & 132.74), C-4-sugar (δ-78.97), C-5-sugar (δ-65.25), C-7 C=O (δ-40.13), C-11 (δ-39.71 & 39.72), C-23 (δ-39.50, 39.29, 39.09 & 38.87), C-12 (δ-30.42) and C-24 (δ-25.02 & 22.43). LC/MS spectral data revealed that the compound found with the molecular weight of 979.1527 and the molecular formula of C₄₈H₈₂O₂₀. The spectra are collected in Figure 1.

Finally, all the spectral data including FTIR, ¹H NMR, ¹³C NMR and LCMS give rise to the prediction of possible chemical structure through the advanced instrumental software for compound (W04) as merremoside with IUPAC name of [4,5-dihydroxy-6-[5-hydroxy-2-methyl-4-(2-methylpropanoyloxy)-6-[(24,25,26-trihydroxy-5,23-dimethyl-9-oxo-19-pentyl-2,4,8,20,22-pentaoxatricyclo

[19.2.2.1^{3,7}] hexacosan-6-yl)oxy]oxan-3-yl]oxy-2-methyloxan-3-yl] 2-methyl propanoate (Figure 2).

HPTLC chromatogram of isolated compounds R_f value showed the single peak at specific values, compound 1 at 0.37 and compound 2 at 0.79. The results are shown in the Figure 3 and Table 1 respectively.

AEIA and its isolated compounds were evaluated for their *in vitro* anticancer activity using MCF – 7 cell lines by colorimetric assay which showed noteworthy inhibitory action on the proliferating cells.

The analysis of half maximal inhibitory concentration (IC₅₀) of all samples concluded the action of W04 was significant with IC₅₀ value of 182.8 µg/ml when it was compared with the extract 220.9 µg/ml, individually, as shown in the Table 2.

Discussion

Role of natural compounds in cancer treatment has been attributed to increasing concentration of drugs that resulted in suppressing cancer cell viability, and migration in addition to causing increased cell death (Nanayakkara et al., 2018). Biochanin A and other natural compounds have exerted MDR reversal through P-gp inhibition (Chung et

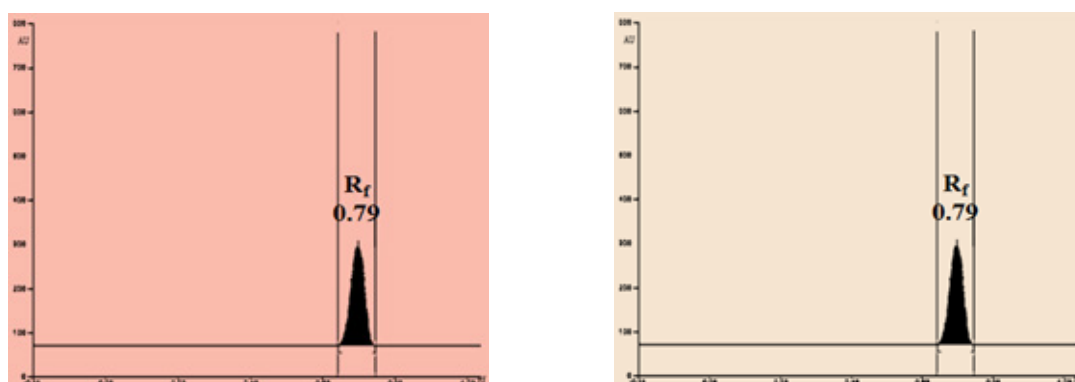


Figure 3. HPTLC Chromatogram of Isolated Compound for W04 at 254nm and at 366nm respectively

al., 2005). Presence of ortho-dihydroxy function in flavone C ring has been suggested to be important for antioxidant and anticancer activity (Raj et al., 2014). Resin glycosides with acyl sugar coupled with amphiphilic nature have been reported to show P-gp inhibition leading to overcoming MDR in cancer (Lira-Ricárdez J et al., 2020; Rybalkina et al., 2021; Syed et al., 2016).

The present results of higher antioxidant activity leading to anticancer activity against breast cancer cell lines are in line with reports of plant phenolics in herbal medicine with strong antioxidant activity (Liaudanskas et al., 2021). Oxidative stress in causing changes in metabolic processes that trigger cancer cell proliferation and the role of quenching of free radical species has been found to be critical to control cancer (Pandey et al., 2009). Higher phenolic content from *Scurrula ferruginea* (Jack) Danser had shown to enhance apoptosis in breast cancer cell lines, due to high level of radical quenching ability (Iriti et al., 2017; Marvibaigi et al., 2016; Rybalkina et al., 2021; Syed et al., 2016; Veena Sharma et al., 2017). In view of the above, the anticancer activity of fraction W04 from *Ipomoea aquatica*, could be correlated with high phenolic content through its antioxidant activity.

In conclusion, the isolated resin glycoside (merromoside) from the *Ipomoea aquatica* has been evaluated for its anticancer activity against human breast cancer cell lines using MCF-7. From the chemical and biological characterization of a lead molecule from Indian water spinach containing flavone hydroxyl group and the acyl groups in resin glycoside W04 might be responsible for the antioxidant activity coupled with inhibiting P-gp efflux through down regulation of ROS and inflammatory cytokines, thereby resulting in inhibiting tumorigenesis. The anticancer effect was highly significant and it correlates with standard anticancer drug Vinblastine. And its potential will be further examined in MDR cancer cell lines.

Author Contribution Statement

SM and SS contributed to the study conception and design. Material preparation, data collection and analysis were performed by SM and SS. MS and NA supervised the process of data collection and analysis. The first draft of the manuscript was written by SM and NA and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This examination was encouraged by the Management and Principal of Karpagam Educational Institutions, Coimbatore. We acknowledge our colleagues from Karpagam College of Pharmacy and Faculty of Pharmacy, Karpagam Academy of Higher Education, Coimbatore, who afforded intuitiveness and proficiency that greatly assisted the research.

This work is a part of approved Ph.D thesis of an author SM.

Ethical Approval Statement

As only invitro cell lines studies are reported in this manuscript, ethical approval statement has not been enclosed.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material; further inquiries can be directed to the corresponding author.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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