RESEARCH ARTICLE

P53 Arg72Pro and MDM2 SNP309 Polymorphisms Cooperate to Increase Lung Adenocarcinoma Risk in Chinese Female Non-smokers: A Case Control Study

Yang-Wu Ren^{1,2}, Zhi-Hua Yin^{1,2}, Yan Wan^{1,2}, Peng Guan^{1,2}, Wei Wu^{1,2}, Xue-Lian Li^{1,2}, Bao-Sen Zhou^{1,2}*

Abstract

Background: Cell cycle deregulation is a major component of carcinogenesis. The *p53* tumor suppressor gene plays an important role in regulating cell cycle arrest, and mouse double minute 2 (MDM2) is a key regulator of *p53* activity and degradation. Abnormal expression of *p53* and MDM2 occurs in various cancers including lung cancer. Methods: We investigated the distribution of the *p53* Arg72Pro (rs1042522) and MDM2 SNP309 (rs2279744) genotypes in patients and healthy control subjects to assess whether these single nucleotide polymorphisms (SNPs) are associated with an increased risk of lung adenocarcinomas in Chinese female non-smokers. Genotypes of 764 patients and 983 healthy controls were determined using the TaqMan SNP genotyping assay. Results: The *p53* Pro/Pro genotype (adjusted OR = 1.55, 95% CI = 1.17–2.06) significantly correlated with an increased risk of lung adenocarcinoma, compared with the Arg/Arg genotype. An increased risk was also noted for MDM2 GG genotype (adjusted OR = 1.68, 95% CI = 1.27–2.21) compared with the TT genotype. Combined *p53* Pro/Pro and MDM2 GG genotypes (adjusted OR = 2.66, 95% CI = 1.54–4.60) had a supermultiplicative interaction with respect to lung adenocarcinoma risk. We also found that cooking oil fumes, fuel smoke, and passive smoking may increase the risk of lung adenocarcinomas in Chinese female non-smokers who carry *p53* or MDM2 mutant alleles. Conclusions: *P53* Arg72Pro and MDM2 SNP309 polymorphisms, either alone or in combination, are associated with an increased lung adenocarcinoma risk in Chinese female non-smokers.

Keywords: P53 - MDM2 - polymorphism - lung cancer - susceptibility

Asian Pac J Cancer Prev, 14 (9), 5415-5420

Introduction

Lung cancer is one of the leading causes of cancerrelated deaths in the world (Jemal et al., 2011). Cigarette smoking is responsible for about 80% of lung cancers; however, only 20% of smokers develop lung cancer (Smyth, 1996; Pesch et al., 2012). Unlike smokers, who commonly develop squamous cell carcinomas, nonsmokers (especially females) are more likely to develop adenocarcinomas (Zang et al., 1996), suggesting that genetic susceptibility and environmental factors may differentially affect lung cancer development. Single nucleotide polymorphisms (SNPs) in several genes involved in carcinogen metabolism, DNA damage repair, cell cycle control, and apoptosis are reported to be associated with lung cancer (Wu et al., 2002; Li et al., 2008; Yin et al., 2009). However, lung cancer development is a multistage process involving both genetic mutations and environmental risk factors. In Chinese women, lung cancer mortality is much higher than expected from the low prevalence of smoking (<4%) in this population (Jemal et al., 2011), indicating that Chinese women may be genetically susceptible to lung cancer development. Therefore, it is important to identify genes and gene–gene interactions that contribute to lung adenocarcinoma risk in Chinese women.

The p53 tumor suppressor protein regulates a number of cellular functions such as cell cycle arrest, gene transcription, and apoptosis in response to DNA damage (Kastan et al., 1991; Dulic et al., 1994). In as many as 50% of human cancers, the p53 gene is inactivated by mutation or deletion (Jin et al., 2001; Lain et al., 2003). The human homolog of mouse double minute 2 (MDM2) negatively regulates p53 by a number of different mechanisms: it inhibits p53 transcriptional activity by directly binding to transactivation domain of the p53 gene (Momand et al., 1992) and directly promotes p53 protein degradation through its E3 ubiquitin ligase activity (Haupt et al., 1997). MDM2 overexpression has been reported in non-small cell lung carcinoma, where it may substitute for p53 protein

¹Department of Epidemiology, School of Public Health, China Medical University, Shenyang 110001, PR China, 2Key Laboratory of Cancer Etiologic and Prevention (The First Hospital of China Medical University), Liaoning Provincial Department of Education, Liaoning, China *For correspondence: bszhou@mail.cmu.edu.cn

Yang-Wu Ren et al

inactivation (Eymin et al., 2002). By inhibiting MDM2 or blocking the p53–MDM2 interaction, tumor cells can reactivate p53 and thereby inhibit tumor growth (Chene, 2003). Thus, p53 and MDM2 play important roles in the development of lung cancer.

The *p53* gene harbors a $G \rightarrow C$ polymorphism in codon 72 that results in the replacement of arginine with proline (Arg72Pro, rs1042522) (Ara et al., 1990). The p53 codon 72 proline allele is less efficient than arginine allele at inducing apoptosis (Dumont et al., 2003). Genetic polymorphisms also exist in the MDM2 gene. For example, Bond et al. (Bond et al., 2004) described a common polymorphism located at position 309 in the first intron of MDM2 gene (SNP309, rs2279744) that leads to increased MDM2 expression and an earlier age of onset of some types of cancer, suggesting that this SNP may play an important role in cancer development. A number of studies have reported an association between p53Arg72Pro and MDM2 SNP309 polymorphisms and lung cancer (Jassem et al., 2005; Hu et al., 2006; Lind et al., 2006; Zhang et al., 2006; Caceres et al., 2009); however, results in different populations are contradictory and none of the studies considered the effects of environmental factors other than smoking.

In this case control study, we investigated the distribution of *p53* Arg72Pro and MDM2 SNP309 genotypes in Chinese female non-smokers to determine whether these SNPs are associated with an increased risk of lung adenocarcinoma. We also examined gene–gene and gene–environment interactions between these SNPs and potential environmental risk factors.

Materials and Methods

Ethics statement

This investigation was approved by the Institutional Review Board of China Medical University. Written informed consent was obtained from each participant.

Study subjects

Part of the patients used in this hospital-based case control study was described previously (Li et al., 2008; Yin et al., 2009). We included 764 lung adenocarcinoma patients with histologically confirmed diagnoses between January 2002 and December 2012. During the same period, 983 healthy controls with no evidence of lung or other cancer were recruited from a medical examination center in the same hospital. Participants were unrelated ethnic Han Chinese women. Face-to-face interviews of patients and healthy control subjects were done by trained interviewers and demographic data, including family history of cancer, and information on exposure to environmental lung cancer risk factors, including cooking oil fumes, fuel smoke, occupational exposure, and passive smoking, were collected. Estimates of the exposure of each participant to environmental factors were reported previously (Li et al., 2008; Yin et al., 2009).

Genotyping

DNA was extracted from 1 mL samples of whole blood using standard phenol–chloroform methods. Genotyping was performed on an Applied Biosystems 7500 FAST Real-Time PCR System (Foster City, CA, USA) using a TaqMan SNP genotyping assay (Affymetrix Inc., Cleveland, Ohio, USA). Each reaction (10 μ L) contained 5 μ L TaqMan Genotyping master mix, 0.5 μ L primers and probes (Applied Biosystems), 2.5 μ L water and 2 μ L DNA (15–25 ng/ μ L). Thermal cycling conditions were 95°C for 10 min, followed by 47 cycles of 92°C for 30 sec and 62°C for 1 min. Duplicates of 10% of the samples were re-tested for quality control purposes.

Statistical analysis

The chi-squared test and Student's t-test were separately used to examine differences in categorical and continuous variables between patients and controls. Hardy–Weinberg equilibrium was tested using a Pearson chi-squared test. The odds ratio (OR) and its 95% confidence interval (95% CI) were obtained by logistic regression methodology to determine correlations between the two polymorphisms, exposure to environmental risk factors, and the incidence of lung adenocarcinoma. We also investigated gene–gene and gene–environment interactions using logistic regression analysis. All analyses were performed using SPSS 13.0 software (SPSS, Inc. Chicago, IL, USA), and a P < 0.05 was considered to be statistically significant.

Results

A total of 852 lung cancer patient (including 77 duplicates) and 1084 healthy control (including 99

Table 1. Demographics of Lung Cancer Patients and Healthy Controls

Variable	No. of patients (%)	No. of controls (%)	OR(95% CI)	P value
Total no.	764	983		
Age (years)	56.47 ± 11.28	56.04 ± 12.11		0.437
Income (Yuan/month)	567.32 ± 367.27	539.84 ± 370.73		0.124
Education				0.570
None	44 (5.8)	49 (5.0)		
Elementary school	322 (42.1)	412 (41.9)		
Junior school	272 (35.6)	337 (34.3)		
Senior school and upwards	126 (16.5)	185 (18.8)		
Fuel smoke exposure	226 (29.6)	263 (26.8)	1.15 (0.93-1.42)	0.192
Cooking oil fume exposure	246 (32.2)	245 (24.9)	1.43 (1.16-1.76)	0.001
Family history of cancer	106 (13.9)	102 (10.4)	1.39 (1.04–1.86)	0.025
Passive smoking	551 (72.1)	684 (69.6)	1.13 (0.92-1.39)	0.248

Table 2. Genotype Frequencies of p53 Arg72Pro andMDM2SNP309Polymorphisms among Patientsand Controls and Their Association with LungAdenocarcinoma Risk

Polymorphism	No. of	No. of	OR (95% CI)	P value
	patients (%)	controls (%)	
P53 Arg72Pro				
Arg/Arg	154 (20.2)	246 (25.0)	Reference	
Arg/Pro	413 (54.1)	522 (53.1)	1.30 (1.02–1.65)	0.035
Pro/Pro	197 (25.8)	215 (21.9)	1.55 (1.17-2.06)	0.002
Arg/Pro + Pro/Pro	610 (79.8)	737 (75.0)	1.37 (1.09–1.73)	0.008
MDM2 SNP309				
TT	202 (26.4)	335 (34.1)	Reference	
TG	391 (51.2)	472 (48.0)	1.42 (1.13-1.77)	0.002
GG	171 (22.4)	176 (17.9)	1.68 (1.27-2.21)	< 0.001
TG + GG	562 (73.6)	548 (65.9)	1.49 (1.21–1.84)	< 0.001

OR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, fuel smoke exposure, and cooking oil fume exposure

Table 3. Interaction of p53 Arg72Pro and MDM2SNP309 Polymorphisms on Lung AdenocarcinomaRisk

P53 Arg 72Pro	MDM SNP30		No. of %) controls	OR (95% CI) ^a	P value
Arg/Arg	TT	52 (6.8)	98 (10.0)	Reference	
Arg/Arg	TG	79 (10.3)	118 (12.0)	1.34 (0.86-2.09)	0.200
Arg/Arg	GG	23 (3.0)	30 (3.1)	1.51 (0.79–2.89)	0.209
Arg/Pro	TT	100 (13.1)	168 (17.1)	1.16 (0.76–1.77)	0.497
Arg/Pro	TG	215 (281)	246 (25.0)	1.78 (1.20-2.62)	0.004
Arg/Pro	GG	98 (12.8)	108 (11.0)	1.88 (1.21-2.93)	0.005
Pro/Pro	TT	50 (6.5)	69 (7.0)	1.52 (0.92-2.51)	0.104
Pro/Pro	TG	97 (12.7)	108 (11.0)	1.84 (1.19–2.86)	0.006
Pro/Pro	GG	50 (6.5)	38 (3.8)	2.66 (1.54-4.60)	<0.001

^a OR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, fuel smoke exposure, and cooking oil fume exposure

duplicates) samples were genotyped. Genotyping was unsuccessful for 12 patient and two healthy control samples. However, all duplicate samples were concordant. After discarding duplicate samples, the study included 764 patients and 983 controls. The demographic characteristics of patients and controls, and their incidence of exposure to environmental lung cancer risk factors, are shown in Table 1. The mean ages of patients and healthy controls were almost identical (56.47 \pm 11.28 and 56.04 \pm 12.11 years). In addition, levels of education, as well as fuel smoke exposure and passive smoking, were similar between patients and controls. However, patients were more likely to have been exposed to cooking oil fumes (*P* = 0.001) and to have a family history of cancer (*P* = 0.025).

Distributions of the *p53* Arg72Pro and MDM2 SNP309 polymorphisms in patients and control subjects are shown in Table 2. The allele frequencies of *p53* Pro and MDM2 G were 0.485 and 0.419, respectively, in healthy controls. The genotype frequencies for *p53* Arg72Pro and MDM2 SNP309 polymorphisms in control subjects conformed to the Hardy–Weinberg equilibrium (P = 0.055 and P = 0.694, respectively). Multivariate logistic regression analysis determined the ORs for Arg/Pro and Pro/Pro genotypes to be 1.30 (95% CI, 1.02–1.65) and 1.55 (95% CI, 1.17–2.06), respectively, compared with *p53* Arg/ Arg genotype, after adjusting for age, family history of cancer, and passive smoking, fuel smoke exposure, and Table 4. Interaction of *p53* Arg72Pro and Exposure to Environmental Factors on Lung Adenocarcinoma Risk

Environmental exposure	1 0	No. of patients (%)			P value
Cooking oil fu	mes				
-	Arg/Arg	99 (13.0)	181 (18.4)	Reference	
-	Arg/Pro	277 (36.3)	387 (39.4)	1.34 (1.00-1.79) ^a	0.048
-	Pro/Pro	142 (18.6)	170 (17.1)	1.56 (1.12-2.18) ^a	0.009
+	Arg/Arg	55 (7.2)	65 (6.6)	1.53 (0.99-2.37) ^a	0.057
+	Arg/Pro	136 (17.8)	135 (13.9)	1.86 (1.32-2.62) ^a	< 0.001
+	Pro/Pro	55 (7.2)	45 (4.6)	2.27 (1.43-3.62) ^a	0.001
Fuel smoke					
-	Arg/Arg	98 (12.8)	171 (17.4)	Reference	1
-	Arg/Pro	305 (39.9)	400 (40.7)	1.34 (1.00-1.80) ^b	0.049
-	Pro/Pro	135 (17.7)	149 (15.2)	1.63 (1.16-2.30)b	0.005
+	Arg/Arg	56 (7.3)	75 (7.6)	1.15 (0.75-1.78) ^b	0.523
+	Arg/Pro	108 (14.1)	122 (12.4)	1.41 (0.97-2.04)b	0.069
+	Pro/Pro	62 (8.1)	66 (6.7)	1.56 (1.01-2.41)b	0.043
Passive smoking	ng				
-	Arg/Arg	56 (7.3)	83 (8.4)	Reference	
-	Arg/Pro	107 (14.0)	150 (15.3)	1.16 (0.76-1.78)°	0.494
-	Pro/Pro	50 (6.5)	64 (6.5)	1.25 (0.75-2.07)°	0.393
+	Arg/Arg	98 (12.8)	163 (16.6)	1.02 (0.65-1.58)°	0.948
+	Arg/Pro	306 (40.1)	374 (38.0)	1.40 (0.95-2.07)°	0.091
+	Pro/Pro	147 (19.2)	149 (15.2)	1.71 (1.11-2.63)°	0.014

^aOR (95% CI) were calculated by logistic regression, adjusted for age **25.0** family history of cancer, passive smoking, and fuel smoke exposure; ^bOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, and cooking oil fume exposure; ^cOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, fuel smoke exposure, and cooking oil fume exposure **0**

 Table 5. Interaction of MDM2 SNP309 and Exposure to

 Environmental Factors on Lung Adenocarcinoma Risk

Environment	al MDN	12 No. of	No. of	OR (95% CI) ^a	P value
exposure	SNP3	09 patients(%	b) controls(%	6)	
Cooking oil:	fumes				
-	TT	117 (15.3)	252 (25.6)	Reference	
-	TG	276 (36.1)	353 (35.9)	1.69 (1.29-2.21) ^a	< 0.001
-	GG	125 (16.4)	133 (13.5)	2.02 (1.45-2.81) ^a	< 0.001
+	TT	85 (11.1)	83 (8.4)	2.12 (1.46-3.10) ^a	< 0.001
+	TG	115 (15.1)	119 (12.1)	2.09 (1.49-2.93) ^a	< 0.001
+	GG	46 (6.0)	43 (4.4)	2.31 (1.44-3.71) ^a	0.001
Fuel smoke					
-	TT	142 (18.6)	250 (25.4)	Reference	
-	TG	280 (36.6)	328 (33.4)	1.53 (1.17-1.98) ^b	0.002
-	GG	116 (15.2)	142 (14.4)	1.46 (1.06-2.02)b	0.022
+	TT	60 (7.9)	85 (8.6)	1.06 (0.71-1.59) ^b	0.761
+	TG	111 (14.5)	144 (14.6)	1.27 (0.91-1.76) ^b	0.155
+	GG	55 (7.2)	34 (3.5)	2.67 (1.65-4.33)b	< 0.001
Passive smoking					
-	TT	64 (8.4)	105 (10.7)	Reference	
-	TG	108 (14.1)	158 (16.1)	1.16 (0.78-1.73)°	0.469
-	GG	41 (5.4)	36 (3.7)	2.03 (1.17-3.53)°	0.012
+	TT	138 (18.1)	230 (23.4)	1.07 (0.72-1.59)°	0.730
+	TG	283 (37.0)	314 (31.9)	1.67 (1.15-2.41)°	0.007
+	GG	130 (17.0)	140 (14.2)	1.72 (1.15-2.59)°	0.009

^aOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, and fuel smoke exposure; ^bOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, and cooking oil fume exposure; ^cOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, fuel smoke exposure, and cooking oil fume exposure

cooking oil fume exposure. Participants carrying the Arg/Pro or Pro/Pro genotype had an elevated adjusted OR of 1.37 (95% CI, 1.09–1.73), compared with Arg/Arg genotype carriers. Similar results were obtained for MDM2 SNP309 genotypes: adjusted ORs for the TG and GG genotypes were 1.42 (95% CI, 1.13–1.77) and 1.68 (95% CI, 1.27–2.21), respectively, compared with the TT genotype. In addition, individuals with a TG or GG genotype had a 1.49-fold increased risk of lung cancer

56

6

Yang-Wu Ren et al

Table 6. Interaction of p53 Arg72Pro, MDM2SNP309 and Exposure to Environmental Factors onLung Adenocarcinoma Risk

Environm	ental No. of	No. of	No. of	OR (95% CI) P value		
exposure	mutation alle	les patients(%) controls(%)		
Cooking of	Cooking oil fumes					
-	0	26 (3.4)	71 (7.2)	Reference		
-	1-2	314 (41.1)	468 (47.6)	1.85 (1.15-2.97) ^a 0.011		
-	3–4	178 (23.3)	199 (20.2)	2.48 (1.51-4.07) ^a <0.001		
+	0	26 (3.4)	27 (2.7)	2.50 (1.23-5.07) ^a 0.011		
+	1-2	153 (20.0)	163 (16.6)	2.58 (1.56-4.26) ^a <0.001		
+	3–4	67 (8.8)	55 (5.6)	3.35 (1.88-5.96) ^a <0.001		
Fuel smol	ke					
-	0	26 (3.4)	71 (7.2)	Reference		
-	1-2	349 (45.7)	458 (46.6)	2.09 (1.30-3.37) ^b 0.002		
-	3–4	163 (21.3)	191 (19.4)	2.40 (1.45-3.95) ^b 0.001		
+	0	26 (3.4)	27 (2.7)	2.16 (1.06-4.42) ^b 0.035		
+	1-2	118 (15.4)	173 (17.6)	1.71 (1.02-2.87) ^b 0.043		
+	3-4	82 (10.7)	63 (6.4)	3.40 (1.93-6.00) ^b <0.001		
Passive sr	noking					
-	0	20 (2.6)	39 (4.0)	Reference		
-	1-2	130 (17.0)	189 (19.2)	1.55 (0.85-2.80)° 0.152		
-	3–4	63 (8.2)	71 (7.2)	2.05 (1.07-3.92)° 0.030		
+	0	32 (4.2)	59 (6.0)	1.26 (0.62-2.57)° 0.947		
+	1-2	337 (44.1)	442 (45.0)	1.82 (1.02-3.25)° 0.091		
+	3-4	182 (23.8)	183 (18.6)	2.43 (1.33–4.44) ^c 0.004		

^aOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, and fuel smoke exposure; ^bOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, passive smoking, and cooking oil fume exposure; ^cOR (95% CI) were calculated by logistic regression, adjusted for age, family history of cancer, fuel smoke exposure, and cooking oil fume exposure

development compared with TT carriers.

We next examined whether a correlation existed between the p53 and MDM2 polymorphisms (Table 3). Using p53 Arg/Arg and MDM2 TT genotype carriers as a reference, we found that individuals carrying the p53Pro/Pro and MDM2 GG genotype had much higher risk of lung adenocarcinoma (2.66) than either p53 Pro/Pro and MDM2 TT genotype carriers (1.52) or p53 Arg/Arg and MDM2 GG genotype carriers (1.51). These results indicate that a supermultiplicative interaction exists between the p53 Pro/Pro and MDM2 GG genotypes in lung adenocarcinoma development (Brennan, 2002).

Gene-environment interactions were also investigated (data shown in Tables 4 and 5). We detected a higher risk of developing lung adenocarcinoma in p53 Pro/Pro genotype carriers exposed to cooking oil fumes or passive smoking: OR = 2.27 and 95% CI = 1.43–3.62 for *p53* Pro/ Pro genotype carriers exposed to cooking oil fumes; OR = 1.71 and 95% CI = 1.11–2.63 for *p53* Pro/Pro genotype carriers exposed to passive smoking. These data suggest that a multiplicative interaction exists between these pairs of genetic and environmental factors. Relative to TT genotype carriers without cooking oil fume exposure, the OR (2.31) for GG genotype carriers with prior exposure to cooking oil fumes is larger than the OR (2.12) for MDM2 TT carriers with cooking oil fume exposure or the OR (2.01) for MDM2 GG genotype carriers without cooking oil fume exposure, but less than expected. These results suggest that antagonism exists between MDM2 SNP309 genotypes and cooking oil fume exposure. Similar results were obtained when MDM2 SNP309 and passive smoking were examined.

Results of the combined analysis of polymorphisms and environmental factor exposure are shown in Table

6. Subjects were divided into three groups according to the number of mutant alleles (0, 1–2, 3–4) they carry. Compared with individuals without mutant alleles and cooking oil fume exposure, ORs for individuals carrying 3–4 mutant alleles (without smoke/fume exposure) or those exposed to cooking oil fumes (without mutations) were 2.48 and 2.50, respectively. However, for individuals carrying 3–4 mutant alleles and exposed to cooking oil fumes, the OR increased to 3.35. Similar results were obtained for subjects carrying 3–4 mutant alleles who were previously exposed to either fuel smoke or passive smoking.

Discussion

In this case control study, we examined whether (1) genetic polymorphisms in p53 and MDM2 and (2) individual environmental factors including cooking fume exposure, fuel smoke exposure, and passive smoking, either alone or in combination, are associated with an increased lung adenocarcinoma risk. We found a statistically significant association between p53 Arg72Pro and MDM2 SNP309 polymorphisms and lung adenocarcinoma risk. Compared with the p53 Arg/Arg genotype, the Pro/Pro genotype correlated with a 1.55-fold increased risk of lung adenocarcinoma; in addition, there was a 1.68-fold increased risk for carriers of the MDM2 GG genotype compared with the MDM2 TT genotype. We also observed a supermultiplication interaction between *p53* Arg72Pro and MDM2 SNP309 polymorphisms. Cooking fume exposure, fuel smoke exposure, and passive smoking all modified the relationship between both polymorphisms and the development of lung adenocarcinoma. A two-sided test with $\alpha = 0.05$ had >80% power to detect an OR of 1.55 for the p53 Pro/Pro genotype and an OR of 1.68 for the MDM2 GG genotype in Chinese female non-smokers.

Our observations are in line with those of several other clinical and biological studies. The allele frequencies of p53 Pro and MDM2 G were 0.485 and 0.419, respectively, among healthy control subjects in this study; these values are similar to those of other studies on Chinese populations (0.38–0.52 and 0.42–0.52, respectively) (Hu et al., 2006; Zhang et al., 2006; Chua et al., 2010; Liu et al., 2013). Li et al. (Li et al., 2009) reported the OR for Asians carrying Pro/Pro genotype to be 1.395 (95% CI = 1.206 - 1.613), which is much higher than that of Caucasians. Zhang et al. (2006) found that smoking modified the association between the p53 Arg72Pro polymorphism and lung cancer; in contrast, both Piao et al. (2011) and Sakiyama et al. (2005) found that it did not. To remove the possible confounding effects of cigarette smoking, all participants in our study were female nonsmokers. Consistent with previous studies (Sakiyama et al., 2005; Piao et al., 2011), we found the p53 Arg72Pro polymorphism to be associated with increased lung adenocarcinoma risk. This result is supported by reports that the Pro allele of the p53 Arg72Pro polymorphism, which is less efficient at inducing apoptosis, reduces the tumor suppression function of p53 and increases cancer risk (Thomas et al., 1999). MDM2 is a key regulator of p53: even a modest change in MDM2 protein levels can affect cancer development by influencing the *p53* pathway (Bond et al., 2005). The MDM2 SNP309 functional SNP is located in the promoter region of MDM2; the G allele is associated with increased MDM2 expression, resulting in *p53* inhibition (Bond et al., 2004). We found G allele carriers to have a higher risk of lung cancer, consistent with results from one Chinese study (Zhang et al., 2006) and a recent meta-analysis (Zhuo et al., 2012). However, another study showed that this MDM2 polymorphism may not be associated with lung cancer in a Chinese population (Hu et al., 2006). These contradictory data are likely to be caused by differences in the lifestyle and geographical location of the participants because our results indicate that environmental factors can cooperate with such polymorphisms to influence lung cancer development.

Lung cancer is a complex multifactorial, polygenic disease, and therefore a single SNP or environmental factor may only have a modest effect on its development. Thus, investigating multiple biologically relevant SNPs and environmental factors may be a more accurate way of evaluating lung cancer risk. p53 and MDM2 form a negative feedback loop that plays a central role in the DNA damage response (Momand et al., 1992). Cooking oil fume exposure, fuel smoke exposure, and passive smoking all induce DNA damage and may consequently increase lung cancer risk (Tokiwa et al., 1994; Mooney et al., 1995; Zhang et al., 2002). Therefore, we analyzed the combined effects of p53 and MDM2 polymorphisms and environmental factor exposure. We found that exposure to cooking oil fumes or fuel smoke, as well as passive smoking, increases the risk of lung adenocarcinoma in female non-smokers who carry p53 or MDM2 mutant alleles, indicating that both MDM2 and p53 polymorphisms interact with all three environmental factors.

Our analysis of cooking oil fume exposure or passive smoking combined with MDM2 SNP309 revealed an interesting phenomenon: ORs for individuals carrying thee GG genotype and exposed to cooking oil fumes or passive smoking were lower than expected. This phenomenon may be linked to (1) previous reports that the MDM2 GG genotype leads to high MDM2 protein expression (Bond et al.,2004) and (2) the ability of MDM2 to negatively regulate estrogen receptor (ER) expression (Duong et al., 2007) because the estrogen signaling pathway is thought to be associated with an increased risk of developing lung cancer, especially adenocarcinoma (Karlsson et al., 2012). In addition, polycyclic aromatic hydrocarbons, which are important component of both cooking oil fumes and cigarette smoke, are reported to decrease estrogen levels (Siegfried, 2010). Thus, it is possible that a reduction in both ER expression and estrogen levels leads to a decreased lung adenocarcinoma risk in female non-smokers.

In conclusion, we have shown that p53 Arg72Pro and MDM2 SNP309, either alone or combination, are associated with an increased lung adenocarcinoma risk in Chinese female non-smokers. Moreover, cooking oil fumes, fuel smoke, and passive smoking may increase the risk of lung adenocarcinoma in Chinese female non-smokers who carry p53 or MDM2 mutant alleles.

Acknowledgements

This study was supported by grant no.81102194 and no.81272293 from National Natural Science Foundation of China. The authors are most grateful to all the participants in the present study. We would also thank all the personnel at the hospitals for their help with sample collection. The author(s) declare that they have no competing interests.

References

- Ara S, Lee PS, Hansen MF, et al (1990). Codon 72 polymorphism of the T*p53* gene. Nucleic acids research, **18**, 4961.
- Bond GL, Hu W, Bond EE, et al (2004). A single nucleotide polymorphism in the MDM2 promoter attenuates the *p53* tumor suppressor pathway and accelerates tumor formation in humans. Cell, **119**, 591-602.
- Bond GL, Hu W,Levine A (2005). A single nucleotide polymorphism in the MDM2 gene: from a molecular and cellular explanation to clinical effect. Cancer Res, **65**, 5481-4.
- Brennan P (2002). Gene-environment interaction and aetiology of cancer: what does it mean and how can we measure it? *Carcinogenesis*, **23**, 381-7.
- Caceres DD, Quinones LA, Schroeder JC, et al (2009). Association between *p53* codon 72 genetic polymorphism and tobacco use and lung cancer risk. *Lung*, **187**, 110-5.
- Chene P (2003). Inhibiting the *p53*-MDM2 interaction: an important target for cancer therapy. *Nat Rev Cancer*, **3**, 102-9.
- Chua HW, Ng D, Choo S, et al (2010). Effect of MDM2 SNP309 and *p53* codon 72 polymorphisms on lung cancer risk and survival among non-smoking Chinese women in Singapore. *BMC Cancer*, **10**, 88.
- Dulic V, Kaufmann WK, Wilson SJ, et al (1994). *p53*-dependent inhibition of cyclin-dependent kinase activities in human fibroblasts during radiation-induced G1 arrest. *Cell*, **76**, 1013-23.
- Dumont P, Leu JI, Della Pietra AC, et al (2003). The codon 72 polymorphic variants of *p53* have markedly different apoptotic potential. *Nat Genet*, **33**, 357-65.
- Duong V, Boulle N, Daujat S, et al (2007). Differential regulation of estrogen receptor alpha turnover and transactivation by Mdm2 and stress-inducing agents. *Cancer Res*, **67**, 5513-21.
- Eymin B, Gazzeri S, Brambilla C, et al (2002). Mdm2 overexpression and p14(ARF) inactivation are two mutually exclusive events in primary human lung tumors. *Oncogene*, **21**, 2750-61.
- Haupt Y, Maya R, Kazaz A, et al (1997). Mdm2 promotes the rapid degradation of *p53. Nature*, **387**, 296-9.
- Hu Z, Ma H, Lu D, et al (2006). Genetic variants in the MDM2 promoter and lung cancer risk in a Chinese population. International journal of cancer. *Int J Cancer*, **118**, 1275-8.
- Jassem J, Szymanowska A, Jassem E, et al (2005). Relation between *p53* codon 72 polymorphism and somatic *p53* gene mutation in non-small cell lung cancer (NSCLC). *Ejc Supplements*, **3**, 336-.
- Jemal A, Bray F,Center MM (2011). Global Cancer Statistics. CA Cancer J Clin, **61**, 69-90.
- Jin S, Levine AJ (2001). The *p53* functional circuit. *J Cell Sci*, **114**, 4139-40.
- Karlsson C, Helenius G, Fernandes O, et al (2012). Oestrogen receptor beta in NSCLC - prevalence, proliferative influence, prognostic impact and smoking. APMIS, 120, 451-8.
- Kastan MB, Onyekwere O, Sidransky D, et al (1991). Participation of *p53* protein in the cellular response to DNA damage. *Cancer Res*, **51**, 6304-11.

Yang-Wu Ren et al

- Lain S,Lane D (2003). Improving cancer therapy by nongenotoxic activation of p53. Eur J Cancer, 39, 1053-60.
- Li M, Yin Z, Guan P, et al (2008). XRCC1 polymorphisms, cooking oil fume and lung cancer in Chinese women nonsmokers. *Lung Cancer*, **62**, 145-51.
- Li Y, Qiu LX, Shen XK, et al (2009). A meta-analysis of T*p53* codon 72 polymorphism and lung cancer risk: evidence from 15,857 subjects. *Lung Cancer*, **66**, 15-21.
- Lind H, Zienolddiny S, Ekstrom PO, et al (2006). Association of a functional polymorphism in the promoter of the MDM2 gene with risk of nonsmall cell lung cancer. International journal of cancer. *Int J Cancer*, **119**, 718-21.
- Liu D, Wang F, Guo X, et al (2013). Association between *p53* codon 72 genetic polymorphisms and tobacco use and lung cancer risk in a Chinese population. *Mol Biol Rep*, **40**, 645-9.
- Momand J, Zambetti GP, Olson DC, et al (1992). The mdm-2 oncogene product forms a complex with the *p53* protein and inhibits *p53*-mediated transactivation. *Cell*, **69**, 1237-45.
- Mooney LA, Santella RM, Covey L, et al (1995). Decline of DNA damage and other biomarkers in peripheral blood following smoking cessation. *Cancer Epidemiol Biomarkers Prev*, 4, 627-34.
- Pesch B, Kendzia B, Gustavsson P, et al (2012). Cigarette smoking and lung cancer-relative risk estimates for the major histological types from a pooled analysis of casecontrol studies. *Int J Cancer*, **131**, 1210-9.
- Piao JM, Kim HN, Song HR, et al (2011). p53 codon 72 polymorphism and the risk of lung cancer in a Korean population. Lung Cancer, 73, 264-7.
- Sakiyama T, Kohno T, Mimaki S, et al (2005). Association of amino acid substitution polymorphisms in DNA repair genes Tp53, POLI, REV1 and LIG4 with lung cancer risk. Int J Cancer, 114, 730-7.
- Siegfried JM (2010). Early changes in pulmonary gene expression following tobacco exposure shed light on the role of estrogen metabolism in lung carcinogenesis. *Cancer Prev Res (Phila)*, **3**, 692-5.
- Smyth JF (1996). Cancer genetics and cell and molecular biology. Is this the way forward? *Chest*, **109**, 125S-9S.
- Thomas M, Kalita A, Labrecque S, et al (1999). Two polymorphic variants of wild-type *p53* differ biochemically and biologically. *Mol Cell Biol*, **19**, 1092-100.
- Tokiwa H, Sera N, Nakashima A, et al (1994). Mutagenic and carcinogenic significance and the possible induction of lung cancer by nitro aromatic hydrocarbons in particulate pollutants. *Environ Health Perspect*, **102**, 107-10.
- Wu X, Zhao H, Amos CI, et al (2002). *p53* genotypes and haplotypes associated with lung cancer susceptibility and ethnicity. *J Natl Cancer Inst*, **94**, 681-90.
- Yin Z, Su M, Li X, et al (2009). ERCC2, ERCC1 polymorphisms and haplotypes, cooking oil fume and lung adenocarcinoma risk in Chinese non-smoking females. J Exp Clin Cancer Res, 28, 153.
- Zang EA,Wynder EL (1996). Differences in lung cancer risk between men and women: examination of the evidence. J Natl Cancer Inst, 88, 183-92.
- Zhang H, Wang G, Tan W (2002). [Study on the effects of cooking oil fume condensate on the DNA integrity]. Wei Sheng Yan Jiu, 31, 238-40.
- Zhang X, Miao X, Guo Y, et al (2006). Genetic polymorphisms in cell cycle regulatory genes MDM2 and Tp53 are associated with susceptibility to lung cancer. *Human mutation*, 27, 110-7.
- Zhuo W, Zhang L, Zhu B, et al (2012). Association of MDM2 SNP309 variation with lung cancer risk: evidence from 7196 cases and 8456 controls. *PLoS One*, **7**, e41546.