# **RESEARCH ARTICLE**

# Association Between ERCC2 Polymorphisms and Glioma Risk: a Meta-analysis

# Li-Ming Huang<sup>1</sup>, Xi Shi<sup>1</sup>, Dan-Fang Yan<sup>2</sup>, Min Zheng<sup>1</sup>, Yu-Jie Deng<sup>1</sup>, Wu-Cha Zeng<sup>1</sup>, Chen Liu<sup>1</sup>, Xue-De Lin<sup>1\*</sup>

# Abstract

ERCC2 is an essential component of the nucleotide excision repair pathway which is involved in the effective maintenance of genome integrity. Association studies on ERCC2 polymorphisms and glioma risk have yielded inconclusive results. This meta-analysis was performed to gain a better insight into the relationship between ERCC2 polymorphisms and glioma risk. A systematic literature search updated to December 2, 2013 was performed in the Pubmed and EMBASE databases. Crude pooled odds ratios (ORs) with their corresponding 95% confidence intervals (95% CIs) were used to estimate the association between ERCC2 polymorphisms and glioma risk under a suitable effect model according to heterogeneity. All analyses were performed using Review Manager 5 (version 5.2) and STATA (version 12.0). The combined results demonstrated rs13181 to be significantly associated with glioma risk (G allele versus T allele: OR=1.15, 95% CI=1.05-1.26, P=0.002; dominant model: OR=1.22, 95% CI=1.07-1.39, P=0.002; recessive model: OR=1.18, 95% CI=0.98-1.41, P=0.070). We also found that rs13181 acts in an allele dose-dependent manner (GG versus TT: OR=1.30, 95% CI=1.07-1.57, P=0.009; TG versus TT: OR=1.20, 95% =CI 1.05–1.37, P=0.009; trend test, P=0.004). However, no evidence was found in analyses for the association between other 3 ERCC2 polymorphisms (rs238406, rs1799793, and rs1052555) and susceptibility to glioma development. Our meta-analysis suggests that rs13181 is significantly associated with glioma risk in an allele dose-dependent manner, whereas, 3 other ERCC2 polymorphisms (rs238406, rs1799793, and rs1052555) may have no influence.

Keywords: ERCC2 - polymorphism - glioma - meta-analysis

Asian Pac J Cancer Prev, 15 (11), 4417-4422

## Introduction

Glioma is the most common type of primary brain tumors. Although some advances have been made in the detection and management of glioma in the past decades, most of the patients with glioma still have a poor prognosis (Schwartzbaum et al., 2006). Numerous studies have been conducted to uncover the etiology of glioma, but only ionizing radiation has so far been established firmly to be an environmental risk factor for glioma (Ohgaki et al., 2005). An important carcinogenic mechanism of ionizing radiation for glioma is inducing various types of DNA damage including single- and double-strand breaks. The accumulation of DNA damages will finally result in tumor occurrence if there is the existence of DNA repair defects. Thus, DNA repair genes are supposed to be the potential susceptibility genes of glioma.

Four major DNA repair pathways are involved in the effective maintenance of genome integrity, including the nucleotide excision repair (NER), base excision repair (BER), double strand break repair (DSBR), and mismatch repair (MMR) pathways (Wood et al., 2001).

ERCC2 is an essential component of the ubiquitous NER pathway. Defects of this gene were established to result in the cancer-prone syndrome xeroderma pigmentosum group D (Lehmann, 2001). Furthermore, polymorphisms located in ERCC2 have been reported to be associated with the susceptibility of several cancers (Duan et al., 2012; Guo et al., 2012). ERCC2 is located in the chromosome 9q13.3 which has been reported to be frequently abnormal in glioma, and decreased copy number of it was also observed to be a common occurrence in glioma (Liang et al., 1995; Yong et al., 1995). Compared with normal brain tissue, down-regulated expression of ERCC2 was also observed in astrocytoma (Smith et al., 2000). Therefore, ERCC2 is considered as an important candidate tumor suppressor gene of glioma naturally.

Based on the hypothesis that polymorphisms located in ERCC2 may affect its DNA repair capacity by multiple mechanisms, several studies were conducted to investigate the association between ERCC2 polymorphisms and glioma risk. Four coding polymorphisms, rs13181 (K751Q), rs238406 (R156R), rs1799793 (D312N), and rs1052555 (D711D) were widely investigated (Caggana

<sup>1</sup>Department of Chemotherapy, The First Affiliated Hospital, Fujian Medical University, Fuzhou, <sup>2</sup>Department of Radiation Oncology, The First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China \*For correspondence: linxdfj2088@ aliyun.com

#### Li-Ming Huang et al

et al., 2001; Wrensch et al., 2005; Yang et al., 2005; Liu et al., 2009; McKean-Cowdin et al., 2009; Rajaraman et al., 2010; Chen et al., 2012; Luo et al., 2013; Salnikova et al., 2013). However, the results from these studies are to some extent divergent, which may be attributed partly to the limited power of individual study. Thus, we performed a meta-analysis to have a better insight into the relationship between these ERCC2 polymorphisms and glioma risk. To our knowledge, this is the most comprehensive meta-analysis conducted to date for the association between ERCC2 polymorphisms and glioma risk.

#### **Materials and Methods**

#### Search strategy and selection criteria

To ensure the rigour of this current meta-analysis, we designed it according to the guidelines of Preferred Reporting Items for Systemic Reviews and Meta-Analyses statement (PRISMA) (Moher et al., 2009). Systematic literature search updated to December 2, 2013 was performed in the Pubmed and EMBASE databases, using the search terms: (ERCC2 OR "Excision repair cross-complementing group 2" OR "DNA repair gene") AND (polymorphism OR variant OR variation) AND (glioma OR "brain tumor"). References of reviews and retrieved studies were also scanned to search for additional relevant studies. The following criteria were used for the study selection: (1) case-control or cohort study design; (2) assessment of the association between ERCC2 polymorphisms and glioma risk; (3) sufficient data were provided for estimating an odds ratio (OR) with 95% confidence interval (CI); (4) the genotype distribution of controls must be in Hardy-Weinberg equilibrium (HWE). Studies with overlapping subjects were also included if they focused on different polymorphisms. If studies had overlapping data on the same polymorphism, we just extracted data from the largest study for final analysis.

#### Data extraction

Data were extracted by two investigators independently complying with the selection criteria listed above. In case of discrepancies, the group discussion was conducted until a consensus was reached. The following data were extracted for each study: first author's name, publication year, origin country, ethnicity (categorized as Caucasian, Asian, or mixed descent), control source, genotyping method, total number of cases and controls, the HWE for controls, and genotype or allele frequency of cases and controls.

#### Statistical Analysis

Statistical analyses were performed using Review Manager 5 (version 5.2; The Cochrane Collaboration, Oxford, United Kingdom) and the STATA statistical software package (version 12.0; StataCorp, College Station, Tex). Crude pooled ORs with their corresponding 95% CIs were used to estimate the association between ERCC2 polymorphisms and glioma risk. Heterogeneity assumption was checked using the Q test and I<sup>2</sup> statistics. P>0.10 for the Q-test indicated a lack of heterogeneity among studies, while I<sup>2</sup>>50% was considered a measure

of severe heterogeneity. According to heterogeneity, pooled ORs were calculated using a fixed-effects model (the Mantel–Haenszel method) in this study. The potential publication bias was estimated by the funnel plot and Egger's linear regression test. All statistical tests were two-sided. P<0.05 was used as the criterion of statistical significance.

#### Results

#### Study selection and characteristics in the meta-analysis

The process of study selection was shown in Figure 1. Based on our search terms, a total of 53 articles were retrieved through Pubmed (24 articles) and EMBASE (29 articles) databases. After an initial screening of the titles and abstracts, 44 of them were excluded based on inclusion and exclusion criteria. Thus, 9 potential articles were remained for full-text view. Moreover, 1 additional study was identified from retrieved articles. After carefully reading the full articles, 2 articles were further excluded because of insufficient data or deviation from HWE. Finally, 8 eligible articles whose characteristics are listed in Table 1 were included in this meta-analysis. All eligible articles were published after 2000. Six of them were conducted in the United States (Caggana et al., 2001; Wrensch et al., 2005; Yang et al., 2005; Liu et al., 2009; McKean-Cowdin et al., 2009; Rajaraman et al., 2010), 1 in China (Chen et al., 2012), and 1 in Russian (Salnikova et al., 2013). Many kinds of genotyping methods were used, including polymerase chain reactionrestriction fragment length polymorphism (PCR-RFLP), allele-specific oligonucleotide hybridization (ASOH), Pyrosequencing, MassARRAY, TaqMan, and allelespecific tetraprimer PCR. Overall, there were 7 articles on rs13181 (Caggana et al., 2001; Wrensch et al., 2005; Liu et al., 2009; McKean-Cowdin et al., 2009; Rajaraman et al., 2010; Chen et al., 2012; Salnikova et al., 2013), 4 articles on rs238406 (Caggana et al., 2001; Wrensch et al., 2005; Yang et al., 2005; Liu et al., 2009), 4 articles on rs1799793 (Caggana et al., 2001; Rajaraman et al., 2010; Chen et al., 2012; Salnikova et al., 2013), and 2 articles

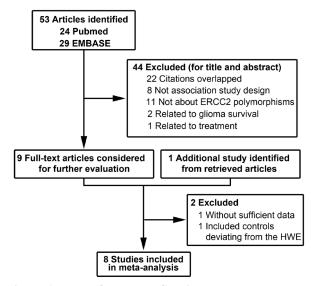


Figure 1. Flow of Included Studies

Table 1. Characteristics of Studies Included in the Meta-analysis

First author	Year	Country	Ethnicity	Cases	Controls	Control sour	ce Genotyping method	SNPs	HWE
Caggana M	2001	USA	Caucasian	161	164	population	PCR-RFLP	rs238406	>0.05
							ASOH	rs1799793	>0.05
							ASOH	rs1052555	>0.05
							PCR-RFLP	rs13181	>0.05
Wrensch M	2005	USA	Mixed	472	558	population	PCR-RFLP	rs238406	0.72
							PCR-RFLP	rs13181	0.40
Yang P	2005	USA	Caucasian	141	108	hospital	Pyrosequencing	rs1052555	>0.05
							Pyrosequencing	rs238406	>0.05
Liu Y	2009	USA	Caucasian	373	365	population	MassARRAY	rs238406	0.34
							MassARRAY	rs13181	0.45
McKean-Cowdin R	2009	USA	Caucasian	1015	1994	mixed	TaqMan/MassARRAY	rs13181	>0.05
Rajaraman P	2010	USA	Caucasian	362	495	hospital	TaqMan	rs1799793	>0.05
							TaqMan	rs13181	>0.05
Chen DQ	2012	China	Chinese	393	410	hospital	TaqMan	rs1799793	0.36
							TaqMan	rs13181	0.18
Salnikova LE	2013	Russian	Caucasian	161	464	hospital	allele-specific tetraprimer PCR	rs1799793	0.07
							allele-specific tetraprimer PCR	rs13181	0.92

SNPs, single nucleotide polymorphisms; HWE, Hardy-Weinberg equilibrium; PCR-RFLP, polymerase chain reaction-restriction fragment length polymorphism; ASOH, allele-specific oligonucleotide hybridization

	Glion	าล	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup					Weight	M-H. Fixed, 95% CI	M-H, Fixed, 95% Cl
G vs T	Literites	10101	LTOING	10101	Treight	M 11. 1 1. CO. 0070 OF	M 11. 1 1400. 0070 01
McKean-Cowdin R 2009	766	1998	1403	3940	68.0%	1.12 [1.01, 1.26]	
Chen DQ 2012	310	786	284	820	19.7%	1.23 [1.00, 1.51]	<b>—</b> —
Salnikova LE 2013	140	322	359	918	12.3%	1.20 [0.93, 1.55]	
Total (95% CI)	140	3106	328	5678	100.0%	1.15 [1.05, 1.26]	•
Total events	1216	3100	2046	3078	100.0 %	1.15 [1.05, 1.20]	•
Heterogeneity: Chi <sup>2</sup> = 0.66,		- 0.72					
Test for overall effect: Z = 3			), 1 0%				
Test for overall effect. Z = 3	5.00 (P =	0.002)					
GG+TG vs TT							
McKean-Cowdin R 2009	623	999	1147	1970	68.8%	1.19 [1.02, 1.39]	
Chen DQ 2012	254	393	235	410	19.3%	1.36 [1.02, 1.81]	
Salnikova LE 2013	107	161	235	459	11.9%	1.18 [0.81, 1.72]	
Total (95% CI)	107	1553	288	2839	100.0%	1.18 [0.81, 1.72]	-
Total events	984	1555	1670	2039	100.0%	1.22 [1.07, 1.39]	-
		0.70					
Heterogeneity: Chi <sup>2</sup> = 0.71,			); 1* = 0%				
Test for overall effect: Z = 3	3.03 (P =	0.002)					
GG vs TG+TT							
McKean-Cowdin R 2009	143	999	256	1970	67.7%	1.12 [0.90, 1.39]	
Chen DQ 2012	56	393	230	410	18.9%	1.22 [0.81, 1.85]	
Salnikova LE 2013	33	393 161	49	410	18.9%	1.22 [0.81, 1.85]	
Total (95% CI)	33	1553	/1	2839	100.0%	1.18 [0.98, 1.41]	
Total events	232	1555	376	2035	100.078	1.10 [0.50, 1.41]	-
		- 0.00					
Heterogeneity: Chi <sup>2</sup> = 0.83,			); 1* = 0%				
Test for overall effect: Z = '	1.79 (P =	0.07)					
GG vs TT							
McKean-Cowdin R 2009	143	519	256	1079	68.3%	1.22 [0.96, 1.55]	+-∎
Chen DQ 2012	56	195	49	224	18.4%	1.44 [0.92, 2.24]	
Salnikova LE 2013	33	87	71	242	13.2%	1.47 [0.88, 2.46]	
Total (95% CI)	00	801		1545	100.0%	1.30 [1.07, 1.57]	-
Total events	232		376				-
Heterogeneity: Chi <sup>2</sup> = 0.68,		= 0.71					
Test for overall effect: Z = 2			,,				
		,					
TG vs TT							
McKean-Cowdin R 2009	480	856	891	1714	68.6%	1.18 [1.00, 1.39]	- <b>-</b> -
Chen DQ 2012	198	337	186	361	19.5%	1.34 [0.99, 1.81]	
Salnikova LE 2013	74	128	217	388	11.9%	1.08 [0.72, 1.62]	
Total (95% CI)		1321		2463	100.0%	1.20 [1.05, 1.37]	•
Total events	752		1294				
Heterogeneity: Chi <sup>2</sup> = 0.83.		= 0.66					
Test for overall effect: Z = 2			,,				
							0.5 0.7 1 1.5 2

Figure 2. Forest Plots of ORs with 95% CI for rs13181 and Glioma Risk

on rs1052555 (Caggana et al., 2001; Yang et al., 2005). After carefully reading the full articles, we found that 4 studies (Caggana et al., 2001; Wrensch et al., 2005; Liu et al., 2009; Rajaraman et al., 2010) might contain partial overlapping data on rs13181 with the study by McKean-Cowdin et al (McKean-Cowdin et al., 2009). Moreover, the study by Caggana M et al (Caggana et al., 2001) may also contain partial overlapping data on rs238406 with the study by Wrensch M et al (Wrensch et al., 2005). The largest study was selected for analysis. Thus, 3 studies were included respectively for the meta-analysis on rs13181 (McKean-Cowdin et al., 2009; Chen et al., 2012; Salnikova et al., 2013) and rs238406 (Wrensch et al., 2005; Yang et al., 2005; Liu et al., 2009). For rs238406, the study by Yang et al (Yang et al., 2005) did not provide sufficient genotype data. We just extracted the allele frequency of cases and controls from this study for further analyses.

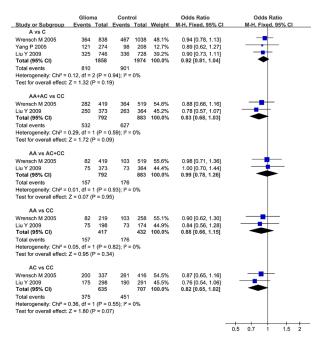


Figure 3. Forest Plots of ORs with 95% CI for rs238406 and Glioma Risk

Because the studies by Rajaraman et al and Salnikova et al focused on glioma and other cancers, we just extracted the data on glioma (Rajaraman et al., 2010; Salnikova et al., 2013).

#### Meta-analysis results

We pooled all eligible studies together for each polymorphism. Since no heterogeneity obviously existed (P>0.10 and I<sup>2</sup><50 % for all polymorphisms), we used fixed-effects model to evaluate the overall association between each polymorphism and susceptibility of glioma. No evidence of publication bias was found in our study even though the number of eligible studies for each polymorphism was less than 10 (data not shown). As shown in Figure 2, a total of 1, 553 glioma cases and 2, 839 healthy controls were included in the meta-analysis on rs13181, and significant association was observed

	Glion	19	Contr			Odds Ratio	Odds Ratio
Study or Subaroup					Weight	M-H. Fixed, 95% CI	M-H, Fixed, 95% Cl
A vs G	Erente	10101	Eterno	10101	mengin	11 11 11 11 10 10 10 10 10	
Caggana M 2001	85	270	98	274	13.0%	0.83 [0.58, 1.18]	
Rajaraman P 2010	240	678	311	936	33.0%	1.10 [0.89, 1.36]	
Chen DQ 2012	294	786	280	820	33.6%	1.15 [0.94, 1.41]	+
Salnikova LE 2013	133	322	337	902	20.4%	1.18 [0.91, 1.53]	
Total (95% CI)	133	2056	331		100.0%	1.10 [0.97, 1.24]	•
Total events	752	2050	1026	2002	100.070	1.10 [0.37, 1.24]	•
Heterogeneity: Chi <sup>2</sup> = 2.9		(D = 0 3		97.			
Test for overall effect: Z				70			
rest for overall effect. Z	= 1.04 (F	- 0.12)					
AA+AG vs GG							
Caggana M 2001	68	135	81	137	14.6%	0.70 [0.43, 1.13]	
Rajaraman P 2010	199	339	254	468	32.2%	1.20 [0.90, 1.59]	
Chen DQ 2012	238	393	233	410	32.9%	1.17 [0.88, 1.55]	
Salnikova LE 2013	97	161	265	451	20.3%	1.06 [0.74, 1.54]	
Total (95% CI)		1028		1466	100.0%	1.09 [0.92, 1.28]	-
Total events	602		833				
Heterogeneity: Chi <sup>2</sup> = 3.9	91. df = 3	(P = 0.2	27); l <sup>2</sup> = 2	3%			
Test for overall effect: Z							
AA vs AG+GG							
Caggana M 2001	17	135	17	137	11.7%	1.02 [0.50, 2.09]	
Rajaraman P 2010	41	339	57	468	33.5%	0.99 [0.65, 1.52]	
Chen DQ 2012	56	393	47	410	31.4%	1.28 [0.85, 1.94]	
Salnikova LE 2013	36	161	72	451	23.4%	1.52 [0.97, 2.37]	
Total (95% CI)		1028		1466	100.0%	1.21 [0.96, 1.53]	
Total events	150		193				
Heterogeneity: Chi <sup>2</sup> = 2.7				%			
Test for overall effect: Z	= 1.59 (P	= 0.11)					
AA vs GG							
Caggana M 2001	17	84	17	73	13.3%	0.84 [0.39, 1.79]	
Rajaraman P 2010	41	181	57	271	32.4%	1.10 [0.70, 1.73]	
Chen DQ 2012	56	211	47	224	30.7%	1.36 [0.87, 2.12]	
Salnikova LE 2013	36	100	72	258	23.6%	1.45 [0.89, 2.37]	
Total (95% CI)	00	576			100.0%	1.23 [0.96, 1.58]	-
Total events	150		193				
Heterogeneity: Chi <sup>2</sup> = 1.8	37. df = 3	(P = 0.6	$50):  ^2 = 0$	%			
Test for overall effect: Z	= 1.60 (P	= 0.11)					
AG vs GG							
Caggana M 2001	51	118	64	120	14.7%	0.67 [0.40, 1.11]	
Rajaraman P 2010	158	298	197	411	31.7%	1.23 [0.91, 1.65]	
Chen DQ 2012	182	337	186	363	33.6%	1.12 [0.83, 1.50]	
Salnikova LE 2013	61	125	193	379	20.0%	0.92 [0.61, 1.38]	
Total (95% CI)		878		1273	100.0%	1.05 [0.88, 1.25]	-
Total events	452		640				
Heterogeneity: Chi <sup>2</sup> = 4.6				6%			
Test for overall effect: Z	= 0.50 (P	= 0.62)					
							0.5 0.7 1 1.5 2

Figure 4. Forest Plots of ORs with 95% CI for rs1799793 and Glioma Risk

	Glion	na	Conti	rol		Odds Ratio		Odd	ls Rati	io	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% CI		M-H, Fi	xed, 9	5% CI	
T vs C											
Caggana M 2001	69	228	91	280	52.5%	0.90 [0.62, 1.31]				-	
Yang P 2005	79	274	64	210	47.5%	0.92 [0.62, 1.37]		_		_	
Total (95% CI)		502		490	100.0%	0.91 [0.69, 1.20]					
Total events	148		155								
Heterogeneity: Chi <sup>2</sup> =	0.01, df =	1 (P = 0	0.93); l² =	0%							
Test for overall effect:	Z = 0.66 (	P = 0.5	1)								
						-	-	0.7	+	45	+
							0.5	0.7	1	1.5	- 2

Figure 5. Forest Plots of ORs with 95% CI for rs1052555 and Glioma Risk

(G allele versus T allele: OR=1.15, 95% CI=1.05–1.26, P=0.002; dominant model: OR=1.22, 95% CI=1.07–1.39, P=0.002; recessive model: OR=1.18, 95% CI=0.98–1.41, 75.0 P=0.070). Furthermore, compared with subjects having the TT genotype, subjects having the GG or TG genotype had an OR of 1.30 (95% CI=1.07–1.57, P=0.009) or 1.20 (95% CI=1.05–1.37, P=0.009) for developing glioma50.0 respectively. The results suggest that rs13181 acts in an allele dose–dependent manner (trend test; P=0.004).

Unfortunately, there was no evidence for the association25. between other polymorphisms and susceptibility of glioma. The results were shown in Figure 3 for rs238406 (A allele versus C allele: OR=0.92, 95% CI=0.81-1.04, P=0.190; dominant model: OR=0.83, 95% CI=0.68-1.03, P=0.090; recessive model: OR=0.99, 95% CI=0.78-1.26, P=0.950; AA versus CC: OR=0.88, 95% CI=0.66-1.15, P=0.340; AC versus CC: OR=0.82, 95% CI=0.65-1.02, P=0.070), Figure 4 for rs1799793 (A allele versus G allele: OR=1.10,95% CI=0.97-1.24, P=0.120; dominant model: OR=1.09,95% CI=0.92-1.28, P=0.320; recessive model: OR=1.21, 95% CI=0.96-1.53, P=0.110; AA versus GG: OR=1.23, 95% CI=0.96-1.58, P=0.110; AC versus GG: OR=1.05, 95% CI=0.88-1.25, P=0.620), and Figure 5 for rs1052555 (T allele versus C allele: OR=0.91, 95% CI=0.69-1.20, P=0.510).

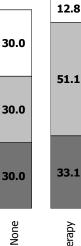
### Discussion

Although glioma accounts for almost 80% of primary brain tumors (Schwartzbaum et al., 2006), it is still a relatively rare entity. Thus, it is difficult for any single study to recruit enough patients for powerful genetic association analyses, which may partly explain the inconclusive results from previous association studies on ERCC2 polymorphisms and glioma risk. To avoid this issue, we pooled all eligible studies together to assess the association between ERCC2 polymorphisms and glioma risk in the present study. We found that rs13181 was significantly associated with glioma risk. However, no evidence was found in analyses for the association between other 3 ERCC2 polymorphisms (rs238406, rs1799793, and rs1052555) and susceptibility of glioma even though positive trend was seen for rs238406.

As an accredited environmental risk factor for glioma, ionizing radiation induces several kinds of DNA damage, including single- and double-strand breaks. Defects in cellular DNA repair pathways will lead to an accumulation of deleterious mutations in genomic DNA that result from non-repair or mis-repair DNA damage induced by endogenous or exogenous agents including ionizing radiation, and then results in the development of cancer (Hoeijmakers, 2009). NER is a ubiquitous sophisticated DNA repair mechanism which plays a predominant role in recognizing and repairing a wide range of structurally unrelated lesions such as bulky adducts and thymidine dimmers (de Laat et al., 1999). ERCC2 is an essential component of the NER pathway. It functions as an ATPdependent DNA helicase which is an integral member of the basal transcription factor BTF2/TFIIH complex (Sung et al., 1993). Mutations in ERCC2 have been reported to reduce the activity of TFIIH complex, which may lead 100.0to repair and transcription\_defects (Coin et al. 1000)

uo repa	ur and	tra	nscripi	.1011	delects (Com et al., 1999).				
Decrea	6.3	pres	10.1	f EF	20.2	nas I	been re	ported to be	
associa		th t		urre	20.3	f gli		Liang et al.,	
0 <sup>1995; Y</sup>	F6 2	al.,		Smi	54.2	,20	25.0	theory, some	
polymo		hs lo		n ez		cha		e amino acid	
sequen		infl	40.0 SI	ene		ons.		ore, ERCC2	
functio		lyn		sms		ossi		lated to the	
Osuscept		of g		In o		y, w		cted the data	
from a		f 1,		ion		nts		839 healthy	
control		le n		alys		s13		d found that	
0 <sup>a signi</sup>		sso	38.0 <sup>e</sup>	bet		ERC	31.3	lymorphism	
rs1318		lioi		e no		hym		lymorphism	
rs1318	31.3	es i		exoi	23.7	EF		It gives rise	
to a Ly		n sı		ion		no a		sidue 751 of	

**CERCC2** protein, which might affect protein functions. Moreover it was observed that the ERC 2 mRNA with rs131810 allele was less stable than the mRNA with rs131811 allele (Masan et al 2012). Thus, the T to G change at rs13181 may induce the downregulation of ERCC2 expression. The association of rs13181 with glioma risk is in correspondence with the above mentioned phenometron that lower expression of ERCC2 is related to the susceptibility of glioma. Previous studies also found that individuals carrying rs13181 TT genotype are likely to have more enhanced protection ability against oxidative or UV-indiced DNA damage than those with GG genotype



(Qiao et al., 2002; Wlodarczyk et al., 2012). These studies on the biologic function of rs13181 all indicates that the G allele may be a risk allele for glioma, which are consistent with our findings that rs13181G allele was significantly associated with increased risk of glioma. Furthermore, our study also found that subjects having the GG genotype had a higher OR than those having the TG genotype, and the P value of trend test is 0.004. The results reveal that the association of rs13181 with glioma risk is likely in an allele dose–dependent manner. Further experiments are needed to verify this phenomenon.

In published articles, there is only one meta-analysis about the relationship between rs13181 and susceptibility of glioma, in which Xu et al found no significant association in either overall population or subgroup populations (Xu et al., 2013). The discrepancy between our results and their analysis may be caused by following reasons. Data from 4 studies by Caggana et al (Caggana et al., 2001), Wrensch et al (Wrensch et al., 2005), McKean-Cowdin et al (McKean-Cowdin et al., 2009) and Rajaraman et al (Rajaraman et al., 2010) were pooled together for the meta-analysis by Xu et al (Xu et al., 2013). Subjects of the study by McKean-Cowdin et al (McKean-Cowdin et al., 2009) were enrolled from 4 centers, the National Cancer Institute (NCI), the National Institute for Occupational Safety and Health (NIOSH), the University of Texas M. D. Anderson Cancer Center (MDA), and the University of California at San Francisco (UCSF). Subjects of the other 3 studies were also from UCSF, UCSF, and NCI respectively. Thus, overlapping data existed in the study by Mckean-Cowdin et al (McKean-Cowdin et al., 2009) and the other 3 studies. They were not excluded from the meta-analysis by Xu et al (Xu et al., 2013). Furthermore, the meta-analysis by Xu et al (Xu et al., 2013) included the study by Luo et al (Luo et al., 2013) in which the genotype frequencies for rs13181 do not conform to HWE. Deviations from HWE in control subjects may bias the estimates of genetic effects in genetic association studies and meta-analysis (Zintzaras, 2010), which should be avoided when extracting the data for meta-analysis. In our study, we excluded the overlapping data from the studies by Caggana et al (Caggana et al., 2001), Wrensch et al (Wrensch et al., 2005) and Rajaraman et al (Rajaraman et al., 2010), as well as the study by Luo et al (Luo et al., 2013). Our results demonstrate that there is a significant association between rs13181 and susceptibility of glioma.

Our study also has some limitations which should be considered in interpreting the results. First, because only published studies indexed by the selected database were included for meta-analysis, publication bias may occur due to missing the unpublished studies or some relevant published studies. Second, because of the limited sample size for glioma in Asian and insufficient information on histologic types extracted from the included studies, data were not stratified by ethnicity and histologic type.

In conclusion, our current results indicate that rs13181 in ERCC2 is a genetic susceptibility factor for developing glioma. The results are consistent with the biologic function of the polymorphism and support the hypothesis that defects of ERCC2 may play a pivotal role in cancer development.

# Acknowledgements

This work was supported by the National Natural Science Foundation of China (Grants No. 81301772).

## References

- Caggana M, Kilgallen J, Conroy JM, et al (2001). Associations between ERCC2 polymorphisms and gliomas. *Cancer Epidemiol Biomarkers Prev*, **10**, 355-60.
- Chen DQ, Yao DX, Zhao HY, Yang SJ (2012). DNA repair gene ERCC1 and XPD polymorphisms predict glioma susceptibility and prognosis. *Asian Pac J Cancer Prev*, **13**, 2791-4.
- Coin F, Bergmann E, Tremeau-Bravard A, Egly JM (1999). Mutations in XPB and XPD helicases found in xeroderma pigmentosum patients impair the transcription function of TFIIH. *EMBO J*, **18**, 1357-66.
- de Laat WL, Jaspers NG, Hoeijmakers JH (1999). Molecular mechanism of nucleotide excision repair. *Genes Dev*, 13, 768-85.
- Duan XL, Gong H, Zeng XT, et al (2012). Association between XPD Asp312Asn polymorphism and esophageal cancer susceptibility: a meta-analysis. *Asian Pac J Cancer Prev*, 13, 3299-303.
- Guo LY, Jin XP, Niu W, et al (2012). Association of XPD and XRCC1 genetic polymorphisms with hepatocellular carcinoma risk. *Asian Pac J Cancer Prev*, **13**, 4423-6.
- Hoeijmakers JH (2009). DNA damage, aging, and cancer. N Engl J Med, 361, 1475-85.
- Lehmann AR (2001). The xeroderma pigmentosum group D (XPD) gene: one gene, two functions, three diseases. *Genes Dev*, **15**, 15-23.
- Liang BC, Ross DA, Reed E (1995). Genomic copy number changes of DNA repair genes ERCC1 and ERCC2 in human gliomas. *J Neurooncol*, **26**, 17-23.
- Liu Y, Scheurer ME, El-Zein R, et al (2009). Association and interactions between DNA repair gene polymorphisms and adult glioma. *Cancer Epidemiol Biomarkers Prev*, 18, 204-14.
- Luo KQ, Mu SQ, Wu ZX, et al (2013). Polymorphisms in DNA repair genes and risk of glioma and meningioma. *Asian Pac J Cancer Prev*, **14**, 449-52.
- McKean-Cowdin R, Barnholtz-Sloan J, Inskip PD, et al (2009). Associations between polymorphisms in DNA repair genes and glioblastoma. *Cancer Epidemiol Biomarkers Prev*, 18, 1118-26.
- Moher D, Liberati A, Tetzlaff J, Altman DG (2009). Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *J Clin Epidemiol*, **62**, 1006-12.
- Moisan F, Laroche-Clary A, Auzanneau C, et al (2012). Deciphering the role of the ERCC2 gene polymorphism on anticancer drug sensitivity. *Carcinogenesis*, **33**, 962-8.
- Ohgaki H, Kleihues P (2005). Epidemiology and etiology of gliomas. *Acta Neuropathol*, **109**, 93-108.
- Qiao Y, Spitz MR, Shen H, et al (2002). Modulation of repair of ultraviolet damage in the host-cell reactivation assay by polymorphic XPC and XPD/ERCC2 genotypes. *Carcinogenesis*, 23, 295-9.
- Rajaraman P, Hutchinson A, Wichner S, et al (2010). DNA repair gene polymorphisms and risk of adult meningioma, glioma, and acoustic neuroma. *Neuro Oncol*, **12**, 37-48.
- Salnikova LE, Belopolskaya OB, Zelinskaya NI, Rubanovich AV (2013). The potential effect of gender in CYP1A1 and GSTM1 genotype-specific associations with pediatric brain tumor. *Tumour Biol*, 34, 2709-19.

Schwartzbaum JA, Fisher JL, Aldape KD, Wrensch M (2006).

Asian Pacific Journal of Cancer Prevention, Vol 15, 2014 4421

#### Li-Ming Huang et al

Epidemiology and molecular pathology of glioma. *Nat Clin Pract Neurol*, **2**, 494-503; quiz 1 p following 16.

- Smith JS, Tachibana I, Pohl U, et al (2000). A transcript map of the chromosome 19q-arm glioma tumor suppressor region. *Genomics*, **64**, 44-50.
- Sung P, Bailly V, Weber C, et al (1993). Human xeroderma pigmentosum group D gene encodes a DNA helicase. *Nature*, 365, 852-5.
- Wlodarczyk M, Nowicka G (2012). XPD gene rs13181 polymorphism and DNA damage in human lymphocytes. *Biochem Genet*, **50**, 860-70.
- Wood RD, Mitchell M, Sgouros J, Lindahl T (2001). Human DNA repair genes. *Science*, **291**, 1284-9.
- Wrensch M, Kelsey KT, Liu M, et al (2005). ERCC1 and ERCC2 polymorphisms and adult glioma. *Neurol Oncol*, 7, 495-507.
- Xu Z, Ma W, Gao L, Xing B (2013). Association between ERCC1 C8092A and ERCC2 K751Q polymorphisms and risk of adult glioma: a meta-analysis. *Tumour Biol*, [Epub ahead of print].
- Yang P, Kollmeyer TM, Buckner K, et al (2005). Polymorphisms in GLTSCR1 and ERCC2 are associated with the development of oligodendrogliomas. *Cancer*, **103**, 2363-72.
- Yong WH, Chou D, Ueki K, et al (1995). Chromosome 19q deletions in human gliomas overlap telomeric to D19S219 and may target a 425 kb region centromeric to D19S112. J Neuropathol Exp Neurol, 54, 622-6.
- Zintzaras E (2010). Impact of Hardy-Weinberg equilibrium deviation on allele-based risk effect of genetic association studies and meta-analysis. *Eur J Epidemiol*, **25**, 553-60.