# **RESEARCH ARTICLE**

# **Relapse and Survival in Bladder Cancer Patients Undergoing microRNA-129 and microRNA-145 Assays**

# Mostafa M M ELfieky<sup>1</sup>, Mohamed Abd El Rahman<sup>2,3\*</sup>, Aysam M Fayed<sup>1,4</sup>, Zahraa Haleem Al-Qaim<sup>5</sup>, Ahmed Khalid Aldhalmi<sup>2</sup>, Eman AE Badr<sup>6</sup>, Amal Abdel Aziz<sup>1</sup>, Gehan M A Ibrahim<sup>1</sup>

## Abstract

**Objective:** The lack of indicators to measure tumor's invasive biological behavior is an important issue. The aim of this study was to examine the effect of miRNAs 129 and 145 on tumor progression as well as patient survival. **Method:** Seventy five breast cancer (BC) patients and 75 controls were included in this research. Two miRNA expressions were estimated using real-time PCR. Biomarkers for BC detection was tested using ROC curves and AUC. **Result:** *miR-129* and *miR-145* expressions were significant. *miR-129* and miR-145 classifiers (AUC = 0.943 and 0.748, respectively) help diagnose BC. Unlike *miR-145*, *miR-129* did not affect the Kaplan–Meier survival curve analysis for progression-free survival at the end of the trial. The development of transitional cell carcinoma disease was found to have a strong correlation with *miR-145* in both univariate and multivariate Cox regression analyses. Additionally, infiltrating + invasive urothelial carcinoma was also found to be correlated with miR-145. Conversely, elevated *miR-129* expression in BC patients did not lead to an increase in cancer-specific recurrence or mortality, as observed in both univariate and multivariate Cox regression studies. **Conclusion:** The miRNA signature can help detect survival-associated miRNAs and develop BC miRNA therapeutics.

Keywords: Bladder cancer- microRNA 129- microRNA 145- relapse- survival

Asian Pac J Cancer Prev, 25 (6), 2113-2121

# Introduction

Bladder cancer (BC) ranks 7<sup>th</sup> and 14<sup>th</sup> in men and women worldwide, respectively [1]. In industrialized countries, men develop BC. It remains the most frequent urinary cancer [2]. Most BCs (stages Ta and T1) are nonmuscle invasive and successful. T2–T4 muscleinvasive bladder tumors are distantly invasive [3,4]. BC is primarily hereditary and environmental [5]. Despite advancements in surgery, radiation, and chemotherapy, breast cancer remains highly fatal [6]. Established BC biomarkers have many diagnostic limitations [7-10]. Novel markers are required to identify and predict BC at an early stage.

Posttranscriptionally, microRNAs (miRNAs) influence gene expression and become tumor suppressors or oncogenes [11,12]. BC miRNA dysregulation was associated with BC [13-15]. Thus, miRNAs can detect cancer. Prior studies demonstrated that miRNAs alter BC cell motility, invasion, proliferation, and survival.

Several studies reported *miR-129-5p* to be the most downregulated in spinal tissues. *miR-129-5p* overexpression improved spinal tissue inflammation, apoptosis, and functional recovery in injured animals [16,17]. Tumor growth and progression involve miR-145-5p. Few studies explored BC and miR-145-5p. miR-145-5p was found in BC cell proliferation and migration [18,19]. The aim of this study was to examine the effect of miRNAs 129 and 145 on tumor progression as well as patient survival.

## **Materials and Methods**

In this study, 150 participants were included: 75 with BC and 75 healthy volunteers confirmed using cystoscopy. This research was conducted at the Clinical Urology Department, Faculty of Medicine, Menoufia University, from November 2021 to December 2022.

<sup>1</sup>Molecular Biology Department, Genetic Engineering and Biotechnology Research Institute (GEBRI), University of Sadat City, Egypt. <sup>2</sup>College of Pharmacy, Al-Mustaqbal University, Babylon, 51001, Iraq. <sup>3</sup>Clinical Pharmacy Department, Badr University Hospital, Faculty of Medicine, Helwan University, Egypt. <sup>4</sup>Medical Laboratory Techniques Department, College of Health and Medical Technique, Al-Mustaqbal University, Babylon, 51001, Iraq. <sup>5</sup>Department of Anesthesia Techniques, Al-Mustaqbal University College, Iraq. <sup>6</sup>Medical Biochemistry and Molecular Biology, Faculty of Medicine, Menoufia University, Egypt. \*For Correspondence: mohamedmahmoud@mustaqbal-college.edu.iq

#### Mostafa M M ELfieky et al

Menoufia University Faculty of Medicine Medical Research Ethics Committee approved the experiment. Radiology, cystoscopy, histology, and clinical tests confirm the diagnosis of BC. Cancer was staged and graded using TNM and WHO standards. Patients with BC had isolated lesions on ultrasound and CT imaging. Sex, age, domicile, socioeconomic level, pathological subtype, grade, smoking, hematuria, relapse or progression use status, and other confounding risk markers were collected in structured personal interviews. All patients consented, knowing the risks.

In total, 3 mL of blood was collected from each participant via aseptic venipuncture. Sample division: 2 mL of whole blood was put into a simple sterile tube and centrifuged at 3500 rpm for 15 min. The serum was used to measure creatinine, urea, and Hb. RT-qPCR RNA Extraction. RNeasy Mini Kit and Qiazol Reagent (Qiagen, Germany) were used to extract total RNA containing miRNA to quantify miR-145 and miR-129 in BC tissues. RNA purity was confirmed using a NanoDrop spectrophotometer (Thermo Scientific, USA). Per instructions, the miScript II First Strand cDNA kit (Qiagen, Germany) reverse-transcribed RNA. The reactions were incubated at 25°C for 5 min, 42°C for 60 min, and 70°C for 5 min in a 2720 Applied Biosystems thermal cycler. RT-qPCR was performed using Singapore SYBR Green PCR Master Mix (Life Technologies, USA) and a thermal Real-time PCR system (Applied Biosystems, Thermo Fisher Scientific, Inc.). After a 10 min incubation at 94°C, the reactions went through 45 cycles of 15 s at 95°C and 30 s at 60°C. Primers were designed as follows:

miR-145- 5p	Forward	5'-CCTTGTCCTCACGGTCCAGT-3'
	Reverse	5'-AACCATGACCTCAAGAACAGTATTT-3
miR-129-5p	Forward	'- CTTTTTGCGGTCTGGGCTTGC -3'
	Reverse	5'- AGCAAGCCCAGACCGCAAAAA -3'
U6	Forward	5'-CTCGCTTCGGCAGCACA-3'
	Reverse	5'-AACGCTTCACGAATTTGCGT-3'
18SrRNA	Forward	5'-GCGGTTCTATTTTGTTGGTTT-3'
	Reverse	5'-ATCGCCGGTCGGCATCGTTT -3'

U6 controlled *miR-145* and miR129. The 2\_DDCt technique (Livak and Schmittgen, 2001) evaluated miR-145-5p and miR129-5p levels on an Applied Biosystems 7500 Real-Time PCR System (Foster City, CA, USA).

#### Statistical Analysis

Computer data were analyzed using IBM SPSS 20.0. A percentage and number represented quantitative data. Only BC deaths were examined. At the last follow-up, censored patients did not attain the endpoint. The survival curves were generated using the Kaplan–Meier method. Survival distribution tests were conducted on subgroups using the log-rank method. Multivariate Cox proportional hazards regression analysis examined miRNA levels' predictive power after controlling for other factors. Significant differences between populations were observed at confidence levels above 95% (p < 0.05) in all two-sided tests. The survival study used Fisher's Exact or Monte Carlo correction, Student's t-test, Mann–Whitney test, and

Chi-square test. Data significance was at the level of 5%.

#### Results

The participants included 75 BC patients (mean:  $68.93 \pm 8.31$  years), with 51 males and 24 females. Of 75 healthy volunteers, 47 males and 28 females were free of chronic diseases and had a mean age of  $67.19 \pm 6.14$  years. Moreover, 75 BC cases were grouped by age (45–73 years) and smoking (25 negative and 50 positive). BC patients and controls shared gender, age, and smoking rates.

# Clinicopathological characteristics of the patient's population

The clinical characteristics of BC are listed in Table 2. Pathological subtypes were transitional cell carcinoma (89.3%), urothelial carcinoma (6.7%), and infiltrating + invasive carcinoma (4.0%). The grades found were as follows: Grades I (22.7%), II (41.3%), and III (36.0%) appear. Hematuria indicated a 40% tumor size compared to that revealed by the CT (60%,  $\leq$ 3 vs. 40%, >3). Prevalence of no recurrence (85.3%) was more significant than that of advancement (14.7%) and duration per month (mean: 11.41 ± 1.53) (Table 1).

Significant variations in urea, creatinine, and Hb levels were found between patients and controls ( $p \le 0.001$ ). Table 2 shows that the case group had significantly lower *miR-129* and *miR-145* expressions than the control group (p = 0.001).

Correlation between the miR-129 and miR-145 expressions with personal, clinicopathological, and biochemical parameters

Hb, pathological subtype, and grade were positively linked with *miR-129* (r = 0.320, p = 0.005). In the cases group, miRNA 145 was strongly correlated with CT tumor size and Hb (r = 0.241, p = 0.037; r = 0.306, p = 0.008). Higher grade stage and CT tumor size decreased *miR-129* and *miR-145* expressions (p = 0.003 and 0.008, respectively; table not supplied).

# miR-129 and miR-145 bladder cancer diagnostic effectiveness

ROC curve analysis revealed that *miR-129* and *miR-145* differentiated patients from controls. The ROC curve analysis for *miR-129* showed an AUC of 0.943 (95% CI = 0.907–0.98; p < 0.001). The values were as follows: diagnostic sensitivity of 94.67%, specificity of 85.5%, PPV of 85.5%, and NPV of 94.0% with maximum at a threshold of  $\leq$ 1.287 (p < 0.001). ROC curve analysis for *miR-145* showed an AUC of 0.748 (95% CI = 0.665–0.831; p < 0.001), with a diagnostic sensitivity of 88.0%, specificity of 62.67%, PPV of 70.2%, and NPV of 83.9%, peaking at  $\leq$ 0.987 (p < 0.001), as shown in Figure 1.

#### Progression-free survival data of the cases

Research indicated no significant difference in progression-free survival between patients with low and high miRNA 129 expression (p > 0.246; mean = 11.250, 77.5%; mean = 11.429, 88.6%). Concurrently, there was a significant difference in progression-free survival between

Table 1. Clinical-Pathological Features of Patients					
	No.	%			
Pathological subtype					
Transitional cell carcinoma	67	89.3			
Urothelial carcinoma	5	6.7			
Infiltrating + Invasive urothelial carcinoma	3	4			
Grade					
Grade I	17	22.7			
Grade II	31	41.3			
Grade III	27	36			
CT tumor size					
≤3	45	60			
>3	30	40			
Min. – Max.	1.50 - 7.50				
Mean $\pm$ S.D.	$3.20 \pm 1.31$				
Median (IQR)		5 – 3.95)			
Hematuria					
Negative	45	60			
Positive	30	40			
Relapse or progression status					
No relapse	64	85.3			
pt. relapsed	11	14.7			
Duration (months)					
Min. – Max.	6.0 - 12.0				
Mean $\pm$ S.D.	$11.41\pm1.53$				
Median (IQR)	12.0 (12.	0 - 12.0)			

1 1

CD /

patients with low and high miRNA 145 expressions at the end of the study (p < 0.001; mean = 10.718, 66.7%; mean = 12.0, 100%), as shown in Figure 2.

The relationship between bladder cancer patients' clinicopathological traits and relapse

Univariate Cox regression analysis showed that parametric data were nonsignificant as disease progression predictors: age (p = 0.375), sex (female; p = 0.284), smoking (p = 0.781), tumor size (p = 0.772), hematuria (p = 0.735), urea (p = 0.801), creatinine (p = 0.864), and Hb (%) (p = 0.914). Regarding the pathological subtypes, transitional cell carcinoma and infiltrating + invasive urothelial carcinoma were significant predictors of disease progression (p = 0.007 and p = 0.030, respectively), whereas urothelial carcinoma and Grade III were not significant predictors of disease progression (p = 0.030and p = 0.121, respectively). As for miR-145, it was an important predictor of disease progression (p < 0.001), whereas miR-129 was not (p = 0.211). Transitional cell carcinoma (p = 0.007) and *miR-145* were significant disease progression predictors in multivariate Cox regression analysis (p<0.001), whereas infiltrating + invasive urothelial carcinoma was not a significant predictor (p = 0.357). Concurrently, all other parameters were nonsignificant (Table 3).

# The relationship between bladder cancer patients' clinicopathological traits and mortality

Univariate Cox regression analysis for case group mortality parameters showed that parametric data were not associated with disease progression: age (p = 0.409), sex (female) (p = 0.268), smoking (p = 0.806), tumor size (P=0.837), hematuria (p = 0.687), urea (p = 0.897), creatinine (p = 0.870), and Hb (%) (p = 0.939). Transitional cell carcinoma (p = 0.014) and *miR-145* were associated with disease progression (p < 0.001) in univariate Cox regression analysis for death in the cases group.



Figure 1. ROC Curve for miRNA 145 and miRNA 129 to Discriminate Patients (n = 75) from Control (n = 75). ROC curve analysis found that miR-129 and miR-145 discriminated patients from control. The ROC curve analysis for miR-129 showed an AUC of 0.943 (95% CI=0.907-0.98, P <0.001). Diagnostic sensitivity (94.67%), specificity (85.5%), PPV (85.5%), and NPV (94.0%) maximum at a threshold of  $\leq 1.287$  (P<0.001). ROC curve analysis for miR-145 showed an AUC of 0.748 (95% CI=0.665-0.831, P <0.001), with diagnostic sensitivity (88.0%), specificity (62.67%), PPV (70.2%), and NPV (83.9%) peaking at  $\leq 0.987$  (P<0.001).

Table 2. Lab	Tests and	miRNA L	Levels in	the Two	Groups
--------------	-----------	---------	-----------	---------	--------

	Patients $(n = 75)$	Control $(n = 75)$	t	р
Urea (mg/dl)				ł
Min. – Max.	47.50 - 69.0	22.0 - 34.0		
Mean $\pm$ S.D.	$53.36 \pm 4.27$	$27.20\pm3.07$	43.099*	< 0.001*
Median (IQR)	52.0 (50.40 - 56.45)	28.0 (25.0 - 30.0)		
Creatinine (mg/dl)				
Min. – Max.	1.60 - 2.40	0.80 - 1.10		
Mean $\pm$ S.D.	$1.97\pm0.19$	$0.96\pm0.09$	41.950*	< 0.001*
Median (IQR)	1.90 (1.80 - 2.10)	0.97 (0.90 - 1.0)		
Hb (gm/dl)				
Min. – Max.	8.90 - 12.0	9.40 - 13.20		
Mean $\pm$ S.D.	$10.49\pm0.80$	$11.99 \pm 1.03$	9.999*	< 0.001*
Median (IQR)	10.50 (9.85 - 11.0)	12.0 (11.60–12.85)		
MiRNA129-5p				
Min. – Max.	0.44 - 2.23	0.76 - 2.80		
Mean $\pm$ S.D.	$0.88\pm0.33$	$1.91\pm0.62$	323.0*	< 0.001*
Median (IQR)	0.85 (0.66 - 0.95)	1.75 (1.40 - 2.60)		
MiRNA145-5p				
Min. – Max.	0.22 - 2.23	0.08 - 3.83		
Mean $\pm$ S.D.	$0.66\pm0.37$	$1.54 \pm 1.07$	1416.0*	< 0.001*
Median (IQR)	0.55 (0.46 - 0.78)	1.37 (0.59 – 2.30)		

IQR, Interquartile range; SD, Standard deviation; t, Student's t-test; p, p value for comparing the two studied groups; \*, Statistically significant at  $p \le 0.05$ 

Urothelial carcinoma was not significant (p = 0.102), nor was infiltrating + invasive urothelial carcinoma (p = 0.069), Grade (III) (p = 0.151), or miRNA 129 (p = 0.168). Transitional cell carcinoma (p = 0.004) and *miR-145* (p < 0.001) were significant predictors of disease progression in multivariate Cox regression analysis for case mortality, whereas all the other parameters were nonsignificant (Table 4).

## Discussion

According to the latest data from the Egyptian National Cancer Institute registry, the prevalence of BC is 10.1% [20]. BC, which is the sixth most common cancer in the

Table 3. Cox Regressior	Univariate and	l Multivariate for	Case Grou	p Relapse	Parameters
-------------------------	----------------	--------------------	-----------	-----------	------------

		Univariate	#Multivariate		
	Р	H.R. (L.L. – U.L. 95%C.I)	р	H.R. (L.L. – U.L. 95%C.I)	
Age (years)	0.375	1.033(0.961 - 1.111)			
Sex (female)	0.284	1.815(0.610 - 5.402)			
Smoking	0.781	1.182(0.364 - 3.839)			
Pathological subtype					
Transitional cell carcinoma	0.007*	0.198(0.061 - 0.647)	0.036*	0.160(0.029 - 0.888)	
Urothelial carcinoma	0.098	3.577(0.789 - 16.207)			
Infiltrating + Invasive urothelial carcinoma	0.030*	5.404(1.179 - 24.764)	0.357	2.682(0.328 - 21.929)	
Grade (III)	0.121	2.371(0.796 - 7.065)			
C.T. tumor size	0.772	1.061(0.710 - 1.587)			
Hematuria	0.735	1.208(0.406 - 3.594)			
Urea	0.801	1.016(0.898 - 1.149)			
Creatinine	0.864	0.775(0.043 - 14.109)			
H.B. (%)	0.914	1.038(0.525 - 2.056)			
miRNA 129	0.211	0.237(0.025 - 2.265)			
miRNA 145	0.001*	0.005(0.0 - 0.113)	0.001*	0.001(0.0 - 0.057)	

HR, Hazard ratio; CI, Confidence interval; LL, Lower limit; UL, Upper limit; #, All variables with p < 0.05 were included in the multivariate; \*, Statistically significant at  $p \le 0.05$ .



Figure 2. Kaplan-Meier Survival Curve for Progression-Free Survival with MiRNA 129 and 145 a and b. No significant difference in progression-free survival between patients with low and high micro-RNA 129 expression was found at the end of the study (P > 0.246, Mean= 11.250, 77.5% & 11.429, 88.6%), while there was a significant difference in progression-free survival between patients with low and high micro-RNA 145 expression at the end of the study (P<0.001, mean 10.718, 66.7% & 12.0, 100%).

Table 4. Univariate and Multivariate Cox Regression Analysis for Case Group Mortality Parameters

		Univariate		#Multivariate
	Р	H.R. (L.L. – U.L. 95%C.I)	Р	H.R. (L.L. – U.L. 95%C.I)
Age (years)	0.409	1.031(0.959 - 1.108)		
Sex (female)	0.268	1.851(0.622 - 5.510)		
Smoking	0.806	1.159(0.357 - 3.764)		
Pathological subtype				
Transitional cell .carcinoma	0.014*	0.228(0.070 - 0.742)	0.004*	0.142(0.038 - 0.533)
Urothelial carcinoma	0.102	3.530(0.779 - 15.992)		
Infiltrating + Invasive urothelial carcinoma	0.069	4.045(0.896 - 18.254)		
Grade (III)	0.151	2.225(0.748 - 6.623)		
C.T. tumor size	0.837	1.043(0.696 - 1.563)		
Hematuria	0.687	1.251(0.420 - 3.727)		
Urea	0.897	1.008(0.892 - 1.140)		
Creatinine	0.87	0.784(0.043 - 14.340)		
H.B. (%)	0.939	1.027(0.516 - 2.043)		
miRNA 129	0.168	0.199(0.020 - 1.971)		
miRNA 145	< 0.001*	0.003(0.0 - 0.083)	< 0.001*	0.142(0.038 - 0.533)

HR, Hazard ratio; CI, Confidence interval; LL, Lower limit; UL, Upper Limit; #, All variables with p < 0.05 were included in the multivariate; \*, Statistically significant at  $p \le 0.05$ 

United States, is uncommon among individuals under the age of 40 [21]. Complexity and limited therapeutic efficacy make BC, the most common urinary system cancer, worse [22]. Thus, BC mechanisms and new treatments must be understood. Ta, T1, and T2-4 are muscle-invasive cancers, and malignancies exist. Long-term follow-up shows that benign, noninvasive papillary tumors seldom become muscle-invasive, up to 60% [23]. Many high-throughput studies examined genetic alterations and gene expression in BC progression [3].

miRNAs regulate mammalian gene expression and physiology [24,25]. Numerous studies demonstrated that improperly produced miRNAs disrupt well-controlled cellular RNA networks, promoting cancer cell growth, progression, and metastasis. Cancer cells' aberrant miRNAs and RNA network modifications explain growth and metastasis. BC cell growth requires dysregulated miRNAs [14]. In the above scenario, we examined *miR-129* and *miR-145* expression levels as diagnostic, prognostic, and therapeutic biomarkers.

UBC is predominantly male, according to the textbook [26]. Smoking and cancer-causing substances may increase in men [27]. Pakistan [28], Netherlands [29], Malaysia [30], and Nigeria [31] investigations support this. UBC instances were 5:1 M: F; however, Rambau et al. [32] from Tanzania showed a female prevalence.

#### Mostafa M M ELfieky et al

Men accounted for 68% of bladder cancer cases in 2019 [33]. Again, Lavery et al. [34] discovered 102 men and 27 females in 129 instances.

Transitional cell carcinoma (TCC) was the most prevalent histological pattern (89.3%), followed by urothelial carcinoma (6.7%) and infiltrating + invasive (4.0%). Similar high TCC rates also exist elsewhere. TCC is the most prevalent BC, according to textbooks and WHO data [26]. However, rural Africans may have lower bladder TCC rates due to lesser chemical exposure [35]. Nepalese Vaidya [36] reported 97.6% TCC. Mubarak et al. [28] found 94.3% TCC in Pakistan. In 2010, the Netherlands cancer registry reported over 90% of TCC urothelial tumors [29]. Kong et al. [30] found 90.4% TCC in Malaysia. According to El-Siddig et al. [37], 70% of TCC cases were superficial and 30% muscle-invasive.

According to Cohen and Brown [38], 40% hematuria was found. Rafique and Javed [39] found 78.6% hematuria, while Ragab et al. [40] reported 72.5%. Gupta et al. [41] found painless hematuria in 40% of BC patients. The majority of BC patients who undergo cystoscopy and provide sufficient urine samples have microhematuria. The tumors were histologically classified using the WHO (1999)/ISUP urothelial neoplasm grading system [42]. First-line testing includes cystoscopy and urine cytology. Cystoscopy is the most effective method for staging and diagnosing BC. Cancers can be classified into high-grade and low-grade categories [43]. Based on aggressiveness, low-grade cancer cells grow slowly, seem normal, and operate like healthy cells, whereas high-grade cells expand quickly, look disordered, and are more likely to go into the bladder muscle layer.

In a study conducted in Nepal, it was shown that 52.2% (n = 120) of the masses were classified as highgrade. Out of these masses, 77.3% were categorized as II (31 cases, 41.3%), III (27 cases, 36.0%), and I (17 cases, 22.7%). However, El-Siddig et al. [37] discovered that two-thirds of the cases were low-grade and one-third were high-grade. Researchers from Malaysia documented that 32.5% of cases of high-grade superficial TCC were observed [30]. In Southern Pakistan, research found a ratio of 3/4 low to 1/4 excellent quality [28]. Low-grade papillary urothelial carcinoma was 53.85% more common than high-grade (34.61%), according to Laishram et al. [44,45], with percentages of 44% and 29.5%, respectively. In Amman city, Al Khader et al. [46] found 57 (58.2%) and 5 (5.1%) high-grade cancers in 65-84-year-old and >84-year-old individuals.

CT helps in determining the localization of tumors [47]. CT displays BC diagnostics, tumor stage, and therapeutic selection vascular structure. Liu et al. [48] reported tumor CT size ranging from  $\leq 3$  cm for 60% to >3 cm for 40%. CT accuracy depends on bladder tumor lesion size. Many studies revealed that microRNAs (miRNAs/miRs) promote oncological and nononcological illnesses through biological signaling networks. miRNAs are now of interest to BC researchers [49]. Small, noncoding RNAs impede translation, lowering target gene expression. In this study, we investigated the effects of *miR-129* on gene expression [50]. The study found that BC patients had less *miR-129* and *miR-145* expression than those in controls.

Volinia et al. observed that miR-143 (2.6-fold) and *miR-145* (7.0-fold) were most downregulated. Most *miR-145* was found in lymphocytes and connective tissue. *miR-145* and miR-143 were downregulated in various malignancies and scarcely expressed in carcinoma and normal urothelial cells [51,52]. The *miR-129*-upregulated genes in advancing samples were target site-rich. TP53INP1 and *miR-129* targets are involved in melanoma [53]. *miR-145* and *miR-129* predict BC prognosis [54,55].

Prostate cancer tissues have less miR-129-5p, according to Gao et al. [56]. In BC, miR-129 had the largest overexpression compared to those in previous studies [55,57]. Prior work indicated that miR-129-5p overexpression decreases ZIC2 expression, impacting the Wnt/ $\beta$ -catenin pathway. E-cadherin expression increased, whereas phosphorylated Wnt,  $\beta$ -catenin, N-cadherin, and vimentin decreased. By targeting ZIC2, miR-129-5p may diminish EMT, angiogenesis, tumorigenesis, and migration. Our findings complemented those of Zabolotneva et al. [57], who found that miR-145 and miR-133a were downregulated in cancer tissues and could distinguish cancer cells from noncancer cells with >70% sensitivity and >75% specificity.

Hb, pathogenic subtype, and grade positively correlated with *miR-129* (r = 0.320; p = 0.005). Low *miR-129* expression in peripheral blood mononuclear cells was linked to aggressive clinical-pathological features like histological grade (p = 0.010), high preoperative PSA level (p = 0.002), pathological stage (p = 0.011), high Gleason score (p = 0.005), lymph node metastasis, angiolymphatic invasion, and biochemical recurrence [58]. Hb, sex, and CT tumor size were strongly correlated with miRNA 145 (r = 0.241, p = 0.037; r = 0.306, p = 0.008). Researchers identified a favorable connection between the high tumor stage and low miR-145. Other studies discovered that linked *miR-145* underexpression to cancer and high-grade urothelial carcinomas downregulated *miR-145* but not low-grade ones [59,60].

Ichimi et al. [13] detected *miR-145* underexpression in BC using microarray and qRT-PCR. They did not link *miR-145* expression to histological grading, staging, or tumor behavior. Dip et al. [61] showed no effect of *miR-145* expression on histological grade, tumor stage, angiolymphatic neoplastic invasion, or recurrence. Research revealed that miRNA 129 may distinguish patients from controls (p < 0.001), with an AUC of 0.943 (95% CI: 0.907–0.978) at cutoff  $\leq 1.287$ , resulting in 94.67% sensitivity, 84.0% specificity, and 85.5% PPV.

MicroRNA 145 can distinguish patients from controls (p < 0.001), with an AUC of 0.748 (95% CI: 0.665–0.831), at cutoff  $\leq$ 0.987, achieving 88% sensitivity, 62.67% specificity, and 70.2% PPV. However, low and high levels of miRNAs 129 and 145 did not affect survival time (mean = 11.500, 87.5%; mean = 11.590, 89.7%) or high levels (mean 11.314, 82.9%; mean = 11.222, 80.6%). The multivariate study showed that prostate cancer patients severely downregulated *miR-129*, a novel independent prognostic factor [58,62]. The BC miRNAs [63-66]. miRNAs may diagnose, prognose, and treat BC [64,65]. Significant indicators for disease progression in univariate

and multivariate Cox regression analysis include TCC (p = 0.007), infiltrating + invasive urothelial carcinoma (p = 0.030), and *miR-145* (p < 0.001). In Table 4, univariate and multivariate Cox regression analysis revealed TCC (p = 0.014; p = 0.004) and *miR-145* as significant disease development predictors (p < 0.001).

In contrast, Kaplan–Meier survival analysis of the miRNA signature showed that six miRNAs, hsa-miR-652-5p, hsa-miR-193b-5p, hsa-*miR-129*-5p, hsa-miR-143-5p, has-miR-496, and hsa-miR-7-1-3p, were good predictors of overall survival in bladder urothelial carcinoma. BC T-stages 1–4 imply cancer progression in surrounding tissue layers, from connective tissue underlying the bladder (T1) to TCC (which were 90% of BC cases), 75%–85% of which are nonmuscle invasive (Tis/CIS, Ta, and T1).

In prostate cancer patients, *miR-145* deletion increased disease progression and poor survival, according to [67]. High miR-143/145 levels independently predicted superficial tumor growth and poor muscle-invasive patient survival [68]. In most cancers, *miR-145* is downregulated. *miR-145* may affect Akt and KRas pathway tumorigenicity in the cell microenvironment. Moreover, *miR-145* is upregulated by P53 and FoxO transcription factors [69-71]. Then, *miR-145* silences c-Myc, causing apoptosis and cell cycle arrest. *miR-145* increased pi53 pathway activation and p53 transcriptional targets BBC3 (PUMA) and CDKN1A (P21), suggesting a tumor-suppressor loop [72]. RREB1 represses *miR-145* for KRas-induced oncogenic cell transformation [73].

Screening and therapeutic follow-up should be replaced with routine *miR-129* and *miR-145* deployment. Urine or plasma tests can detect noninvasive symptoms. Furthermore, we need more participants to confirm our findings. The miRNA signature may improve survival-associated miRNA research and BLC miRNA target-based therapies.

In conclusion, human BC malignancies depended on *miR-145* and *miR-129* for carcinogenesis, progression, histological pattern, grade, CT tumor size, recurrence, survival, and mortality. It may also be a future BC biomarker used for screening, prognosis, and identifying treatment targets.

# **Author Contribution Statement**

All authors contributed to the study idea and design, experimental work, paper writing, editing, and revision.

# Acknowledgements

## Funding statement

The study was self-funded. The authors did not receive any external funds. The study is part of an approved student thesis.

## Ethical approval and consent to participate

Studies Including Human Subjects. The Declaration of Helsinki was followed in the conduct of the study. The Menoufia University Faculty of Medicine Ethics Committee approved the study procedure, and all subjects supplied written informed permission.

## Conflicts of interest

The study authors affirm that no conflicts of interest could influence the findings.

# References

- Leiblich A. Recent developments in the search for urinary biomarkers in bladder cancer. Curr Urol Rep. 2017;18(12):100. https://doi.org/10.1007/s11934-017-0748-x.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209-49. https://doi.org/10.3322/caac.21660.
- Dyrskjot L, Zieger K, Orntoft TF. Recent advances in highthroughput molecular marker identification for superficial and invasive bladder cancers. Front Biosci. 2007;12:2063-73. https://doi.org/10.2741/2211.
- 4. Tan WS, Tan WP, Tan MY, Khetrapal P, Dong L, deWinter P, et al. Novel urinary biomarkers for the detection of bladder cancer: A systematic review. Cancer Treat Rev. 2018;69:39-52. https://doi.org/10.1016/j.ctrv.2018.05.012.
- Ye F, Wang L, Castillo-Martin M, McBride R, Galsky MD, Zhu J, et al. Biomarkers for bladder cancer management: Present and future. Am J Clin Exp Urol. 2014;2(1):1-14.
- Jiang QQ, Liu B, Yuan T. Microrna-16 inhibits bladder cancer proliferation by targeting cyclin d1. Asian Pac J Cancer Prev. 2013;14(7):4127-30. https://doi.org/10.7314/ apjcp.2013.14.7.4127.
- Xu X, Li S, Lin Y, Chen H, Hu Z, Mao Y, et al. Microrna-124-3p inhibits cell migration and invasion in bladder cancer cells by targeting rock1. J Transl Med. 2013;11:276. https:// doi.org/10.1186/1479-5876-11-276.
- Lenis AT, Lec PM, Chamie K, Mshs MD. Bladder cancer: A review. Jama. 2020;324(19):1980-91. https://doi. org/10.1001/jama.2020.17598.
- Lokeshwar VB, Schroeder GL, Selzer MG, Hautmann SH, Posey JT, Duncan RC, et al. Bladder tumor markers for monitoring recurrence and screening comparison of hyaluronic acid-hyaluronidase and bta-stat tests. Cancer. 2002;95(1):61-72. https://doi.org/10.1002/cncr.10652.
- Lokeshwar SD, Lopez M, Sarcan S, Aguilar K, Morera DS, Shaheen DM, et al. Molecular oncology of bladder cancer from inception to modern perspective. Cancers (Basel). 2022;14(11). https://doi.org/10.3390/cancers14112578.
- 11. Zhang M, Zhuang Q, Cui L. Mir-194 inhibits cell proliferation and invasion via repression of rap2b in bladder cancer. Biomed Pharmacother. 2016;80:268-75. https://doi.org/10.1016/j.biopha.2016.03.026.
- Grimaldi AM, Lapucci C, Salvatore M, Incoronato M, Ferrari M. Urinary mirnas as a diagnostic tool for bladder cancer: A systematic review. Biomedicines. 2022;10(11). https://doi. org/10.3390/biomedicines10112766.
- Ichimi T, Enokida H, Okuno Y, Kunimoto R, Chiyomaru T, Kawamoto K, et al. Identification of novel microrna targets based on microrna signatures in bladder cancer. Int J Cancer. 2009;125(2):345-52. https://doi.org/10.1002/ijc.24390.
- Yoshino H, Seki N, Itesako T, Chiyomaru T, Nakagawa M, Enokida H. Aberrant expression of micrornas in bladder cancer. Nat Rev Urol. 2013;10(7):396-404. https://doi. org/10.1038/nrurol.2013.113.
- 15. Wang L, Fu D, Qiu Y, Xing X, Xu F, Han C, et al. Genomewide screening and identification of long noncoding rnas

and their interaction with protein coding rnas in bladder urothelial cell carcinoma. Cancer Lett. 2014;349(1):77-86. https://doi.org/10.1016/j.canlet.2014.03.033.

- Guo X, Piao H, Zhang Y, Sun P, Yao B. Overexpression of microrna-129-5p in glioblastoma inhibits cell proliferation, migration, and colony-forming ability by targeting zfp36l1. Bosn J Basic Med Sci. 2020;20(4):459-70. https://doi. org/10.17305/bjbms.2019.4503.
- 17. Wan Y, Yang ZQ. Lncrna neat1 affects inflammatory response by targeting mir-129-5p and regulating notch signaling pathway in epilepsy. Cell Cycle. 2020;19(4):419-31. https:// doi.org/10.1080/15384101.2020.1711578.
- Xu WX, Liu Z, Deng F, Wang DD, Li XW, Tian T, et al. Mir-145: A potential biomarker of cancer migration and invasion. Am J Transl Res. 2019;11(11):6739-53.
- Zhang H, Jiang M, Liu Q, Han Z, Zhao Y, Ji S. Mir-145-5p inhibits the proliferation and migration of bladder cancer cells by targeting tagln2. Oncol Lett. 2018;16(5):6355-60. https://doi.org/10.3892/ol.2018.9436.
- 20. Xu Y, Huo R, Chen X, Yu X. Diabetes mellitus and the risk of bladder cancer: A prisma-compliant meta-analysis of cohort studies. Medicine (Baltimore). 2017;96(46):e8588. https:// doi.org/10.1097/md.00000000008588.
- 21. Babjuk M, Burger M, Compérat EM, Gontero P, Mostafid AH, Palou J, et al. European association of urology guidelines on non-muscle-invasive bladder cancer (tat1 and carcinoma in situ) - 2019 update. Eur Urol. 2019;76(5):639-57. https://doi.org/10.1016/j.eururo.2019.08.016.
- 22. Aghaalikhani N, Rashtchizadeh N, Shadpour P, Allameh A, Mahmoodi M. Cancer stem cells as a therapeutic target in bladder cancer. J Cell Physiol. 2019;234(4):3197-206. https://doi.org/10.1002/jcp.26916.
- 23. Cookson MS, Herr HW, Zhang ZF, Soloway S, Sogani PC, Fair WR. The treated natural history of high risk superficial bladder cancer: 15-year outcome. J Urol. 1997;158(1):62-7. https://doi.org/10.1097/00005392-199707000-00017.
- Di Leva G, Croce CM. Roles of small rnas in tumor formation. Trends Mol Med. 2010;16(6):257-67. https://doi. org/10.1016/j.molmed.2010.04.001.
- Garzon R, Marcucci G, Croce CM. Targeting micrornas in cancer: Rationale, strategies and challenges. Nat Rev Drug Discov. 2010;9(10):775-89. https://doi.org/10.1038/ nrd3179.
- Bray F, Ren JS, Masuyer E, Ferlay J. Global estimates of cancer prevalence for 27 sites in the adult population in 2008. Int J Cancer. 2013;132(5):1133-45. https://doi.org/10.1002/ ijc.27711.
- Halaseh SA, Halaseh S, Alali Y, Ashour ME, Alharayzah MJ. A review of the etiology and epidemiology of bladder cancer: All you need to know. Cureus. 2022;14(7):e27330. https://doi.org/10.7759/cureus.27330.
- Mubarak M, Kazi J, Hashmi A, Hussain M, Naqvi SA, Rizvi SAH. Urinary bladder tumors in southern pakistan: A histopathological perspective. Middle East J Cancer. 2014;5(3):167-73.
- Ploeg M, Aben KK, Hulsbergen-van de Kaa CA, Schoenberg MP, Witjes JA, Kiemeney LA. Clinical epidemiology of nonurothelial bladder cancer: Analysis of the netherlands cancer registry. J Urol. 2010;183(3):915-20. https://doi. org/10.1016/j.juro.2009.11.018.
- 30. Kong CH, Singam P, Hong GE, Cheok LB, Azrif M, Tamil AM, et al. Clinicopathological features of bladder tumours in a single institution in malaysia. Asian Pac J Cancer Prev. 2010;11(1):149-52.
- Anunobi CC, Banjo AA, Abdulkareem FB, Daramola AO, Akinde OR, Elesha SO. Bladder cancer in lagos: A 15 year histopathologic review. Niger Postgrad Med J.

2010;17(1):40-4.

- 32. Rambau PF, Chalya PL, Jackson K. Schistosomiasis and urinary bladder cancer in north western tanzania: A retrospective review of 185 patients. Infect Agent Cancer. 2013;8(1):19. https://doi.org/10.1186/1750-9378-8-19.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin. 2019;69(1):7-34. https://doi.org/10.3322/ caac.21551.
- 34. Lavery HJ, Zaharieva B, McFaddin A, Heerema N, Pohar KS. A prospective comparison of urovysion fish and urine cytology in bladder cancer detection. BMC Cancer. 2017;17(1):247. https://doi.org/10.1186/s12885-017-3227-3.
- 35. Heyns CF, van der Merwe A. Bladder cancer in africa. Can J Urol. 2008;15(1):3899-908.
- 36. Vaidya S, Lakhey M, K CS, Hirachand S. Urothelial tumours of the urinary bladder: A histopathological study of cystoscopic biopsies. JNMA J Nepal Med Assoc. 2013;52(191):475-8.
- 37. El-Siddig AA, Albasri AM, Hussainy AS, Alhujaily AS. Urinary bladder cancer in adults: A histopathological experience from madinah, saudi arabia. J Pak Med Assoc. 2017;67(1):83-6.
- Cohen RA, Brown RS. Clinical practice. Microscopic hematuria. N Engl J Med. 2003;348(23):2330-8. https://doi. org/10.1056/NEJMcp012694.
- Rafique M, Javed AA. Clinico-pathological features of bladder carcinoma: Experience from a tertiary care hospital of pakistan. Int Urol Nephrol. 2006;38(2):247-50. https:// doi.org/10.1007/s11255-006-6676-1.
- Ragab HH, El-badry MS, Abdel ghani MM, Aboelhassan MH. Urinary bladder carcinoma pattern at urology minia university hospital. MJMR. 2021;32(1):36-43.
- 41. Gupta P, Jain M, Kapoor R, Muruganandham K, Srivastava A, Mandhani A. Impact of age and gender on the clinicopathological characteristics of bladder cancer. Indian J Urol. 2009;25(2):207-10. https://doi.org/10.4103/0970-1591.52916.
- 42. Epstein JI, Amin MB, Reuter VR, Mostofi FK. The world health organization/international society of urological pathology consensus classification of urothelial (transitional cell) neoplasms of the urinary bladder. Bladder consensus conference committee. Am J Surg Pathol. 1998;22(12):1435-48. https://doi.org/10.1097/00000478-199812000-00001.
- Park JC, Citrin DE, Agarwal PK, Apolo AB. Multimodal management of muscle-invasive bladder cancer. Curr Probl Cancer. 2014;38(3):80-108. https://doi.org/10.1016/j. currproblcancer.2014.06.001.
- 44. Laishram RS, Kipgen P, Laishram S, Khuraijam S, Sharma DC. Urothelial tumors of the urinary bladder in manipur: A histopathological perspective. Asian Pac J Cancer Prev. 2012;13(6):2477-9. https://doi.org/10.7314/ apjcp.2012.13.6.2477.
- 45. Ahmed Z, Muzaffer S, Khan M, Kayani N, Pervez S, Husseini AS, et al. Transitional cell carcinomas of the urinary bladder. A histopathological study. J Pak Med Assoc. 2002;52(9):396-8.
- 46. Al Khader AM, Abu Shahin NI, Obeidat FN, Al-Chalabi MA. Urinary bladder cancer in jordanian adults: A histopathological and epidemiological study from a tertiary care center in amman. J Pak Med Assoc. 2019;69(3):415-7.
- 47. Li L, Na R, Mi T, Cheng H, Ma L, Chen G. Medical image diagnostic value of computed tomography for bladder tumors. Comput Math Methods Med. 2021;2021:3781028. https://doi.org/10.1155/2021/3781028.
- 48. Liu M, Li M, Liu J, Wang H, Zhong D, Zhou H, et al. Elevated urinary urea by high-protein diet could be one of the

Relapse and Survival in Bladder Cancer Patients Undergoing microRNA-129 and microRNA-145 Assays inducements of bladder disorders. J Transl Med. 2016;14:53. 2011 update. Eur Urol. 2011;59(6):997-1008. https://doi.

https://doi.org/10.1186/s12967-016-0809-9.

- Braicu C, Cojocneanu-Petric R, Chira S, Truta A, Floares A, Petrut B, et al. Clinical and pathological implications of mirna in bladder cancer. Int J Nanomedicine. 2015;10:791-800. https://doi.org/10.2147/ijn.S72904.
- Deng B, Tang X, Wang Y. Role of microrna-129 in cancer and non-cancerous diseases (review). Exp Ther Med. 2021;22(3):918. https://doi.org/10.3892/etm.2021.10350.
- 51. Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, et al. A microrna expression signature of human solid tumors defines cancer gene targets. Proc Natl Acad Sci U S A. 2006;103(7):2257-61. https://doi.org/10.1073/pnas.0510565103.
- 52. Slaby O, Svoboda M, Fabian P, Smerdova T, Knoflickova D, Bednarikova M, et al. Altered expression of mir-21, mir-31, mir-143 and mir-145 is related to clinicopathologic features of colorectal cancer. Oncology. 2007;72(5-6):397-402. https://doi.org/10.1159/000113489.
- Bonazzi VF, Irwin D, Hayward NK. Identification of candidate tumor suppressor genes inactivated by promoter methylation in melanoma. Genes Chromosomes Cancer. 2009;48(1):10-21. https://doi.org/10.1002/gcc.20615.
- 54. Villadsen SB, Bramsen JB, Ostenfeld MS, Wiklund ED, Fristrup N, Gao S, et al. The mir-143/-145 cluster regulates plasminogen activator inhibitor-1 in bladder cancer. Br J Cancer. 2012;106(2):366-74. https://doi.org/10.1038/ bjc.2011.520.
- 55. Dyrskjøt L, Ostenfeld MS, Bramsen JB, Silahtaroglu AN, Lamy P, Ramanathan R, et al. Genomic profiling of micrornas in bladder cancer: Mir-129 is associated with poor outcome and promotes cell death in vitro. Cancer Res. 2009;69(11):4851-60. https://doi.org/10.1158/0008-5472. Can-08-4043.
- 56. Gao G, Xiu D, Yang B, Sun D, Wei X, Ding Y, et al. Mir-129-5p inhibits prostate cancer proliferation via targeting etv1. Onco Targets Ther. 2019;12:3531-44. https://doi. org/10.2147/ott.S183435.
- Zabolotneva AA, Zhavoronkov A, Garazha AV, Roumiantsev SA, Buzdin AA. Characteristic patterns of microrna expression in human bladder cancer. Front Genet. 2012;3:310. https://doi.org/10.3389/fgene.2012.00310.
- 58. Xu S, Yi XM, Zhou WQ, Cheng W, Ge JP, Zhang ZY. Downregulation of mir-129 in peripheral blood mononuclear cells is a diagnostic and prognostic biomarker in prostate cancer. Int J Clin Exp Pathol. 2015;8(11):14335-44.
- Shi B, Sepp-Lorenzino L, Prisco M, Linsley P, deAngelis T, Baserga R. Micro rna 145 targets the insulin receptor substrate-1 and inhibits the growth of colon cancer cells. J Biol Chem. 2007;282(45):32582-90. https://doi.org/10.1074/ jbc.M702806200.
- Fujii T, Shimada K, Tatsumi Y, Hatakeyama K, Obayashi C, Fujimoto K, et al. Microrna-145 promotes differentiation in human urothelial carcinoma through down-regulation of syndecan-1. BMC Cancer. 2015;15:818. https://doi. org/10.1186/s12885-015-1846-0.
- Dip N, Reis ST, Srougi M, Dall'Oglio MF, Leite KR. Expression profile of microrna-145 in urothelial bladder cancer. Int Braz J Urol. 2013;39(1):95-101; discussion 2. https://doi.org/10.1590/s1677-5538.Ibju.2013.01.12.
- 62. Xu S, Yi XM, Zhang ZY, Ge JP, Zhou WQ. Mir-129 predicts prognosis and inhibits cell growth in human prostate carcinoma. Mol Med Rep. 2016;14(6):5025-32. https://doi. org/10.3892/mmr.2016.5859.
- 63. Babjuk M, Oosterlinck W, Sylvester R, Kaasinen E, Böhle A, Palou-Redorta J, et al. Eau guidelines on nonmuscle-invasive urothelial carcinoma of the bladder, the

2011 update. Eur Urol. 2011;59(6):997-1008. https://doi. org/10.1016/j.eururo.2011.03.017.

- 64. Prasad SM, Decastro GJ, Steinberg GD. Urothelial carcinoma of the bladder: Definition, treatment and future efforts. Nat Rev Urol. 2011;8(11):631-42. https://doi.org/10.1038/ nrurol.2011.144.
- 65. Sexton WJ, Wiegand LR, Correa JJ, Politis C, Dickinson SI, Kang LC. Bladder cancer: A review of non-muscle invasive disease. Cancer Control. 2010;17(4):256-68. https://doi. org/10.1177/107327481001700406.
- 66. Dip N, Reis ST, Timoszczuk LS, Viana NI, Piantino CB, Morais DR, et al. Stage, grade and behavior of bladder urothelial carcinoma defined by the microrna expression profile. J Urol. 2012;188(5):1951-6. https://doi.org/10.1016/j.juro.2012.07.004.
- Avgeris M, Stravodimos K, Fragoulis EG, Scorilas A. The loss of the tumour-suppressor mir-145 results in the shorter disease-free survival of prostate cancer patients. Br J Cancer. 2013;108(12):2573-81. https://doi.org/10.1038/ bjc.2013.250.
- 68. Avgeris M, Mavridis K, Tokas T, Stravodimos K, Fragoulis EG, Scorilas A. Uncovering the clinical utility of mir-143, mir-145 and mir-224 for predicting the survival of bladder cancer patients following treatment. Carcinogenesis. 2015;36(5):528-37. https://doi.org/10.1093/carcin/bgv024.
- 69. Sachdeva M, Zhu S, Wu F, Wu H, Walia V, Kumar S, et al. P53 represses c-myc through induction of the tumor suppressor mir-145. Proc Natl Acad Sci U S A. 2009;106(9):3207-12. https://doi.org/10.1073/pnas.0808042106.
- Suzuki HI, Yamagata K, Sugimoto K, Iwamoto T, Kato S, Miyazono K. Modulation of microrna processing by p53. Nature. 2009;460(7254):529-33. https://doi.org/10.1038/ nature08199.
- Gan B, Lim C, Chu G, Hua S, Ding Z, Collins M, et al. Foxos enforce a progression checkpoint to constrain mtorc1activated renal tumorigenesis. Cancer Cell. 2010;18(5):472-84. https://doi.org/10.1016/j.ccr.2010.10.019.
- 72. Spizzo R, Nicoloso MS, Lupini L, Lu Y, Fogarty J, Rossi S, et al. Mir-145 participates with tp53 in a death-promoting regulatory loop and targets estrogen receptor-alpha in human breast cancer cells. Cell Death Differ. 2010;17(2):246-54. https://doi.org/10.1038/cdd.2009.117.
- 73. Kent OA, Chivukula RR, Mullendore M, Wentzel EA, Feldmann G, Lee KH, et al. Repression of the mir-143/145 cluster by oncogenic ras initiates a tumor-promoting feedforward pathway. Genes Dev. 2010;24(24):2754-9. https:// doi.org/10.1101/gad.1950610.



This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.