

RESEARCH ARTICLE

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The Expression of *PDL-1* and *PD1* in the Microenvironment of Oral Squamous Cell Carcinoma

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Abstract

Objective: Immune checkpoint proteins, especially *PD-1/PD-L1*, play a vital role in controlling the intensity and duration of the immune response. However, cancer cells often over-express *PDL-1* on their surfaces, which leads to the permanent activation of the *PD-1/PDL-1* pathway and exhaustion of T cells and creates a resistant tumor microenvironment. This study aimed to analyze *PD-1*, *PD-L1* expression in tumor cells (*PDL-1/TC*) and in Tumor-Infiltrating Lymphocytes (*PDL-1/TILS*) of OSCC patients and associated with and to correlate it with histologic grade of malignancy and clinicopathologic parameters. **Methods:** The sample consisted of 43 archived specimens of 43 patients of OSCC with Clinical features (gender, age, smoking, clinical stage) collected from medical records between 2014-2021. The intensity of *PD-1*, *PDL-1/TC*, *PDL-1/TILS*, positive cells were assessed by immunohistochemistry. **Result:** *PDL-1/TILS* and *PDL-1/TC* were observed in all specimens except two cases in well-differentiated were negative. *PDL-1/TILS* was significant between histological grades ($P=0.004<0.05$). There was no significant between *PDL-1/TC* and *PDL-1/TILS* and between poorly differentiated and moderately differentiated groups' ROC P values ($P=0.133, 0.340>0.05$) respectively. There is a difference between *PDL-1/TC* and *PDL-1/TILS* between poorly differentiated and well-differentiated groups ROC P value (0.005, 0.028), Sensitivity (0.857, 0.857), specificity (0.765, 0.824) respectively and there is a difference between and *PDL-1/TILS* moderate differentiated and well-differentiated groups ROC P value (0.133,) Sensitivity (0.737), specificity (0.765). No differences between *PDL-1/TC* and moderate and well-differentiated groups P value (0.173). There is a significant correlation between *PDL-1/TC* and *PDL-1/TILS* with an age $P>65$ value (0.032) in the well-differentiated group. *PDL-1/TC* and *PDL-1/TILS* with the depth of invasion (DOI) in the well-differentiated group. No significant correlation was obtained between *PDL-1/TC* and *PDL-1/TILS* and smoking, clinical stage, and gender. No significant correlation was obtained between PD1 and the histological grades or clinicopathologic characteristics. **Conclusion:** *PDL-1/TILS* and *PDL-1/TC* are independent prognostic factors in OSCC and *PDL-1/TILS* has an important role.

Keywords: *PD1- PDL-1- TILS- OSCC- Tumor-Infiltrating Lymphocytes- Immune checkpoints*

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Introduction

Oral Squamous Cell Carcinoma accounts for 90% of all oral malignancies [1]. It was estimated that about 300,000 new cases were diagnosed as OSCC and about 150,000 cases have died from OSCC in 2020 [2, 3]. OSCC predominantly occurs in males who are in their fifth to seventh decade of life [4]. Recent studies also found an association of squamous cell carcinoma with females under the age of 45 with new cases in young patients [5, 6].

Tumor microenvironment plays a key role in tumor growth and progression [7]. The TME contains many components including immune cells (Tumor-Infiltrating Lymphocytes [TILs], macrophages, and dendritic cells); cancer-associated fibroblasts, and endothelial cells, fibronectin and collagen fibers soluble factors [8, 9]. It has been shown that immune checkpoints play an

important role in the tumor microenvironment and can be manipulated as a mechanism for tumor immune evasion [10]. the most recent studied - is *PD-1/PD-L1* pathway. which control The duration and intensity of the immune response and prevents autoimmune diseases [11, 12]. However, in the tumor microenvironment (TME), the *PD-1/PD-L1* axis is involved in cancer cells immune escape [13]. And the binding of *PD-1* to programmed death ligand 1 (PDL1), negatively regulates T-cell-mediated immune responses, T cell activation [14] and can be manipulated as a mechanism for tumor immune evasion [15].

PD-1 (CD279) a type I transmembrane protein composed of 288 amino acid and belong to the immunoglobulin CD28 family [16]. It was first described in the early 1990s given its expression during induction of apoptosis in a T-cell hybridoma [17] *PD-1* is expressed

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in many immune cells, including peripherally activated T cells, B cells, monocytes, natural killer (NK) cells, and certain DCs [18]. and the recent studies showed that *PD1* is expressed in tumor cells in some cancers, such as melanoma and liver cancers [19, 20] while its ligands *PD-L1* and *PD-L2* are mainly expressed in antigen-presenting cells and tumor cells [21].

programmed death ligand 1 (PDL1/CD274) is a 290-amino-acid type I transmembrane glycoprotein belonging to B7-CD28 family of the immunoglobulin superfamily [22]. It is expressed in tumor cells and (APCs) various types of immune cells, including activated B cells and T cells, macrophages, and dendritic cells [18, 20] and the engagement of PD-L1 with *PD-1* of T cell creates T cell exhaustion, , and interleukin-10 (IL-10) production [24]. Another interacting molecule such as B7-1 (CD80), a protein expressed on activated T cells and APCs, interacts with the PD-L1 of tumor cells causing the negative regulation of activated T-cell [25].

The interaction between tumor cells and T cells and immune checkpoint is complex in tumor microenvironment of OSCC. We aim in this study to evaluate the correlation between the expression of *PD1/PDL-1* pathway and T cells in OSCC microenvironment.

Materials and Methods

The sample

43 resection specimens of 43 patients of OSCC treated from 2014 to 2021 were taken from the archives of Damascus hospital. clinical data from medical records of patients were obtained (e.g., gender, age, smoking, clinical stage,...). Tumor stage and clinical stage were classified as initial (I and II) or advanced (III and IV). (Table 1). Histologic grades were classified according to WHO criteria into three groups well- differentiated, moderately differentiated, and poorly differentiated tumors [26]. clinical UICC-stage (I–IV) and TNM classification of OSCC were documented in the histologic reports according to the guidelines of the most recent World Health Organization classification of tumors of the head and neck and the International Union Against Cancer [27].

Immunohistochemistry Stain

Paraffin-embedded specimens were cut into 4 µm thickness and mounted on immunological slides and then dried by autoclave. Sections were deparaffinized in xylene for 3 minutes followed by rehydrated with an alcohol gradient (100%, 95%, ,70%) then washed with water.

For antigen retrieval, and according to manufacturer’s instruction of the Ventana company @the sections were boiled in Immune DNA Retriever by microwave for 30 minutes followed by washing with (wash buffer) for 5mints followed with blocked by incubation with Poly Detector Peroxidase Blocker for 10 mints. then tissue sections were washed in TBS (wash buffer)for 5 minutes 3 times followed by incubation with the primary antibodies *PDL-1*(VENTANA, SP263) *PD1* VENTANA, for 1h at 4 °C overnight and then washed by TBS(Tris -Buffered Saline) for 15 mints After that the sections were incubation with the secondary antibody, Poly Detector plus link

Table 1. Clinicopathologic Features of Oral Squamous Cell Carcinoma (n = 43)

Clinicopathologic	Number	Percentage
Features	n=(43)	%
Age	43-82	64.7
Gender		
Male	24	55.8
Female	19	44.2
Smoking		
Yes	19	44.2
No	24	55.8
Clinical stage		
I/II	21	48.8
III/IV	22	51.2
Lymph node		
N0	27	62.8
N1	9	20.9
N2	7	16.3
Metastasis		
M0	0 (0)	0
M1	0 (0)	0
Location		
Toung	22	51.2
Other sites	21	48.8

for 15 mints followed by washing with TBS and then incubation with Poly Detector HRP label for 15 mints followed by washing with TBS(Tris-buffered saline)for 15 mints followed by (DAB) for 15 min and then washed with H₂O. Finally the samples were counterstained with Meyer’s hematoxylin and mounted.

Evaluation of *PD1* and *PDL-1*

For each slide, five representative fields were selected for counted the numbers of *PD1* and *PDL-1* at high power (400 ×) The final density of each section was calculated as the average number of five high-power fields (HPFs). Only membranous *PD-L1* expression was considered positive in tumor cells, whereas cytoplasmic and/or membranous reactions were considered positive in TILs the cutoff is *PD-L1* ≥5 [28, 29]. for *PD1* cytoplasmic and/or membranous any staining considered positive in TILs. All slides were evaluated by two investigators blinded to the clinical data.

Statistical analysis

One-Way ANOVA test was used to associate expression of *PDL-1*/TC and *PDL-1*/TILS with histologic grade of malignancy. and Student’s t-test was used for associations expression of *PDL-1*/TC and *PDL-1*/TILS clinicopathological parameters. operating characteristic (ROC) and area under curve (AUC) analyses were used to estimate the predictive value of *PDL-1*/TC and *PDL-1*/TILS + in the prognosis of OSCC. Sensitivity and specificity were estimated. Analyses were conducted with SPSS 13.0

Results

Patient Characteristics

The patients were 24 men (55.8) and 19 women (44.2) and ranged in age from 43 to 82 year (mean 64.7, standard deviation 9.4). The tumor sub site was classified as the tongue (n =22= 51.2%), and other sites of oral cavity (n =21= 48.8) is reported in 19 (44.2), 21 (51.2) had stage I/ II, 22 (51.2%) had stage III /IV. table (1), OSCC grades were 17(39.5%) well-differentiated, 19(44.2%) moderately and 7 (16.3%) poorly-differentiated (Table 1).

Associations between PD1, PDL-1/TC and PDL-1/TILS density and clinicopathologic features

No significant correlation was obtained between the expression of (PDL-1/TC, PDL-1/TILS and PD-1) with smoking, T stage and gender. There is a significant correlation between PDL-1/TC and PDL-1/TILS with ages \geq 65 p value (0.032) in well differentiated group (Table 2). And PDL-1/TC with the depth of invasion (DOI) in well differentiated group (Table 3).

Associations between PD1, PDL-1/TC and PDL-1/TILS and histological grade of OSCC

The expression of PD1, PDL-1/TILS and PDL-1/TC were observed in all specimens. Figures (1, 2) except two cases in well differentiated were negative. There was a significant difference between PDL-1/TILS and the histological grades(P=0.004<0.05) Table 4. we noticed that PD1 was expressed in some tumor cells Figure (1). And most expression was in poor- differentiated group with no significant differences with histological grades.

The prognostic value of PDL-1/TILS and PDL-1/TC for poorly-differentiated OSCC between the three groups

There was no significant between PD1, PDL-1/TC and PDL-1/TILS and between poorly differentiated and moderate differentiated groups ROC P value (P=0.236,0.133, 0.340>0.05) respectively. there is a difference between PD1, PDL-1/TC and PDL-1/TILS between poorly differentiated and well differentiated groups ROC P value (0.024,0.005, 0.028) Sensitivity (0.857,0.857, 0.857), specificity (0.765,0.765, 0.824)

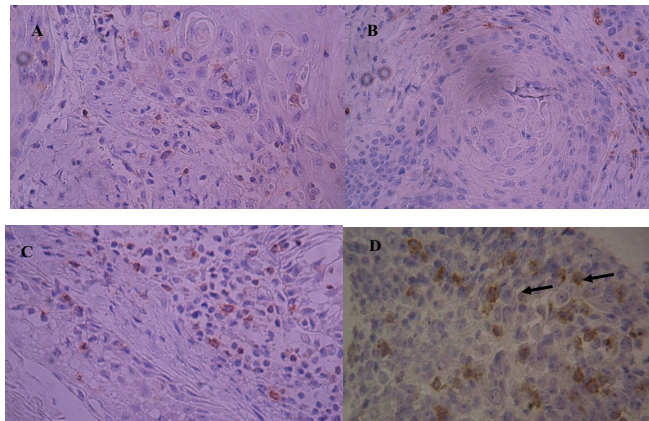


Figure 1. PD1 Expression in OSCC A: well-differentiated samples, B: Moderately-differentiated samples, and C: poorly-differentiated samples. D: PD1 expression in some tumor cells (the arrows)

Table 2. Average Numbers of PD1, PDL-1/TILS and PDL-1/TC and coleration with clinicopathologic features of OSCC according to histological grades of OSCC. P and r indicate P-value and correlation coefficient from T student test Asterisk (*) indicates P-value and correlation coefficient from Pearson test.

Parameter	OSCC grades	PDL-1/TILS		PDL-1/TC		PD1	
		P-value	The difference between the two averages	P-value	The difference between the two averages	P-value	The difference between the two averages
Sex	well-differentiated	3.86	0.559	6.18	0.693	4.85	0.264
	Moderately-differentiated	4.61	0.524	10.61	0.426	6.19	0.293
	poorly-differentiated	14.5	0.246	0.9	0.96	0.58	0.58
Age \geq 65	well-differentiated	16.29	0.023	41.33	0.013	9.23	0.06
	Moderately-differentiated	0.89	0.905	7.18	0.601	6.68	0.267
	poorly-differentiated	9.58	0.418	2.97	0.824	6.67	0.552
Smoking	well-differentiated	5.21	0.448	5.87	0.305	1.89	0.682
	Moderately-differentiated	5.87	0.411	16.52	0.92	2.51	0.671
	poorly-differentiated	9.58	0.418	2.97	0.824	6.67	0.552
stage I/II AND III/IV	well-differentiated	4.4	0.505	20.97	0.168	2.24	0.612
	Moderately-differentiated	5.58	0.294	4.01	0.771	0.11	0.986
	poorly-differentiated	24.33	0.107	1	0.966	4.5	0.78

Table 3. The Result of Pearson Test, which Present the Correlation of *PDL-1/TILS*, *PDL-1/TC* and *PD1* with Depth of Invasion(DOI).

Parameter	OSCC grades	<i>PDL-1/TILS</i>		<i>PDL-1/TC</i>		<i>PD1</i>	
		P-value*	Correlation coefficient value	P-value*	Correlation coefficient value	P-value*	Correlation coefficient value
Depth of invasion(DOI) *	well-differentiated	0.172	0.347	0.043	0.495	0.297	0.269
	Moderately-differentiated	0.606	0.126	0.601	4.01	0.687	0.099
	poorly-differentiated	0.95	0.029	0.07	0.716	0.199	0.552

respectively and there is a significant difference between *PDL-1/TILS* between moderately- differentiated and well- differentiated groups ROC P value (0.0103,). Specificity (0.765) Sensitivity (0.737) Table 5, Figure 3.

The value of Sensitivity and specificity of PDL-1/TILS and PDL-1/TC as a prognostic for poorly-differentiated OSCC in the group of well-differentiated and poorly differentiated

The best value between sensitivity and specificity was at the value of *PD1*, *PDL-1/TILS* and *PDL-1/TC* 40 /74 where the sensitivity value was equal to 0.714,0.857and the specificity value was equal to 0.941, 0.824and therefore we conclude that the value 40 /74 can be determined as a standard value for *PDL-1/TILS* and *PDL-1/TC* rate as a predictor of moderately-differentiated OSCC in a sample search. Table 6, Figure 4.

The value of Sensitivity and specificity of *PDL-1/TILS* and *PDL-1/TC* as a prognostic for poorly-differentiated OSCC in the group of Well-differentiated and Moderately

differentiated. The best value between sensitivity and specificity was at the value of *PDL-1/TILS* 32.5 where the sensitivity value was equal to 0.737 and the specificity value was equal to 0.765 and therefore we conclude that the value 32.5 can be determined as a standard value for *PDL-1/TILS* rate as a predictor of moderately-differentiated OSCC in a sample search Table (6). The correlation of *PDL-1/TILS* and *PDL-1/TC* and the expression of T cells (*CD4*, *CD8*). From our previous study. (Wahbi and Mandili , 2022) we found a correlation between *PDL-1/TILS* with the immune expression of *CD4* Table (7)

Discussion

Cancer cells express high levels of *PD-L1* witch activate the *PD-L1/PD1* pathway as an immune escape mechanism, that increases their survival rate [30]. The overexpressed of *PD-L1* on tumor cells binds to the *PD-1* on tumor-infiltrating lymphocytes (TILs), which counteracts the TCR-signaling cascade through

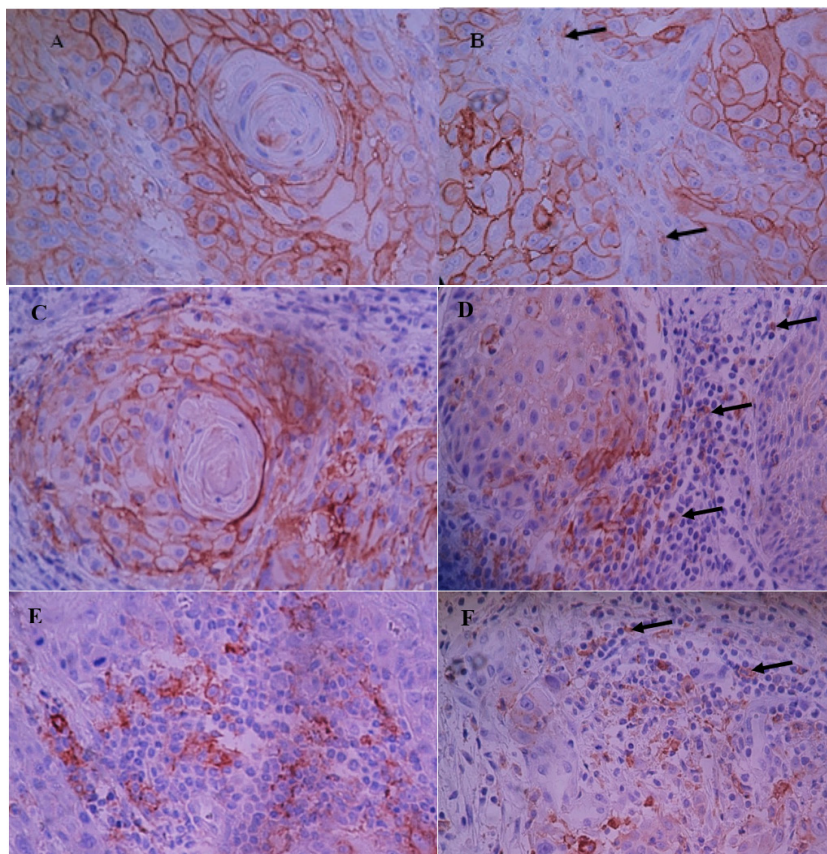


Figure 2. *PDL-1* and Expression in Tumor Cells and TILS (the arrows) OSCC A, B well-differentiated samples, C, D Moderately-differentiated samples, and E, F poorly-differentiated samples.

Table 4. Average Numbers of PD-1, PDL-1/TILS, PD-L1/TC According to Histological Grades of OSCC

OSCC grades	Number of patients	PD1		PDL-1/TILS		PDL-1/TC	
		Mean± SD	P-value	Mean± SD	P-value	Mean± SD	P-value
Well-differentiated	17	12.06± 8.69	0.056	21.71± 13.03	0.004	50.35± 30.79	0.102
Moderately- differentiated	19	18.21± 12.34	3.098	34.42± 15.02	6.429	64.89± 27.75	2.415
Poorly-differentiated	7	23.86± 12.99		42.14± 13.96		75.86± 18.77	

Table 5. The Results of (ROC) and Area under Curve (AUC) Test

Parameter	poorly-differentiated & Moderately-differentiated		well-differentiated & poorly-differentiated		well-differentiated & Moderately -differentiated	
	(ROC) and area under curve (AUC)	P.value	(ROC) and area under curve (AUC)	P.value	(ROC) and area under curve (AUC)	P.value
PDL-1/TILS	0.305	0.133	0.803	0.005	0.751	0.0103
PDL-1/TC	0.376	0.34	0.374	0.028	0.633	0.173
PD-1	0.376	0.34	0.79	0.028	0.644	0.141

Table 6. The Sensitivity and Specificity Values of PD1, PDL-1/TILS and PDL-1/TC between the Groups of Well-Differentiated and Poorly-Differentiated and the Groups of Well-Differentiated and Moderately -differentiated

	well-differentiated and poorly-differentiated			well-differentiated and Moderately -differentiated	
	PDL-1/TILS	PDL-1/TC	PD-1	PDL-1/TILS	
Sensitivity	0.714	0.857	0.857	0.737	
specificity	0.941	0.824		0.765	
Ratio is equal to or greater than	40	74	16.5	32.5	

phosphorylating SHP-2 [32], and represents the main inhibitory signal on T cell activity [10, 11]. This study is a part of our research for Master's degree and we evaluated the expression of CD4+, CD8 in the same sample of this study [33]. In this study we evaluated the expression of PD1 in TILS only, we noticed some positive tumor cells, indicating that tumor cells may express PD1; most studies on PD-1 expression have focused on immune cells, making its expression and functions in cancer cells unclear [31, 32]. Recent studies have shown that cancer cells can express PD1; such as the study of Yao et al, [19], which included a re-analysis of PD-1 expression in different cancer types. Immunohistochemical staining for

PD-1 within this study also showed that hepatocellular carcinoma, renal carcinoma, urothelial carcinoma, testicular cancer, and melanoma have subsets of tumor cells staining positive for PD-1 [18]. Other studies have reported the presence of PD-1 in melanoma [34, 35], breast cancer [36] and liver cancer [37] and lung cancer [38]. However, the mechanism of PD-1 expression in cancer cells is unknown, and may be explained by several mechanisms such as gene copy number modifications, epigenetic modifications, microenvironment, etc. According to the CCLE dataset, PD-1 is expressed in some cancer cell lines, with potential effects on PD-1 mRNA expression. In addition, cytokines and immune

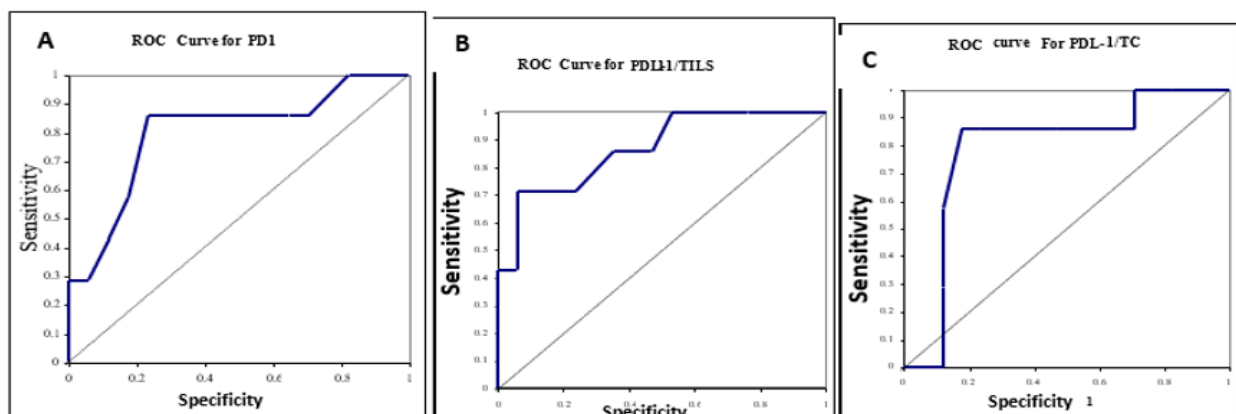


Figure 3. The ROC Curve between the Groups of Well-Differentiated and Poorly-Differentiated. (A) represent the ROC Curve for PD1, (B) represent the ROC Curve for PDL-1/TILS and C represent the ROC Curve for PDL-1/TC

Table 7. The Correlation of *PDL-1/TILS* and *PDL-1/TC* and the Expression of T Cells (*CD4*, *CD8*)

Parameter	Histological grade	<i>PDL-1/TC</i>		<i>PDL-1/TILS</i>	
		Correlation coefficient value	P. value	Correlation coefficient value	P. value
CD4	Well -differentiated	0.302	0.239	0.279	0.277
	Moderately -differentiated	0.093	0.706	0.289	0.231
	poorly -differentiated	0.49	0.316	0.003	0.436
CD8	Well-differentiated	-0.353	0.164	0.416	0.097
	Moderately -differentiated	0	1	0.09	0.715
	Poorly -differentiated	0.094	0.841	0.61	0.146

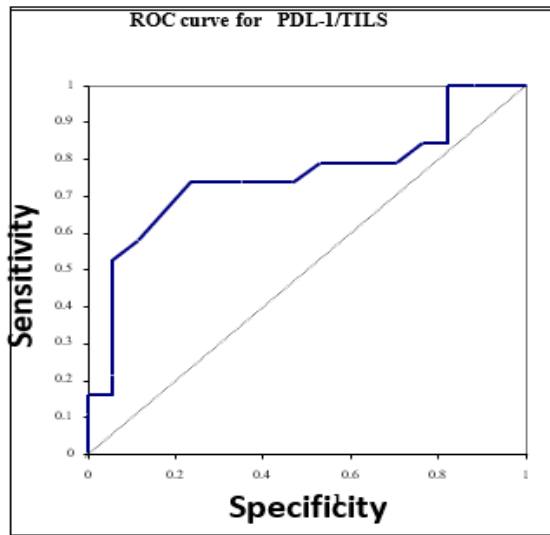


Figure 4. The ROC Curve for *PDL-1/TILS* between the Groups of Well-Differentiated and Moderately -Differentiated

cells present in the tumor microenvironment may also be involved in stimulating tumor *PD-1* expression [29] According to what was mentioned above, head and neck cancers did not express *PD-1* in cancer cells, and our study can be considered the first study to indicate the presence of *PD-1* in OSCC cells, but in our study we did not include *PD-1* expression within the statistical analysis. We noticed that in well-differentiated cancer samples, *PD-1* expression in TILS was positive, with two samples were having negative staining. This is considered a high expression rate compared to the study of Maruse et al. [39] that showed *PD-1* expression in 97 oral squamous cell carcinoma samples was positive in (61.9%) of cases. In our study, the highest expression was in poorly differentiated group, with no statistically significant differences in histological grades. We found a prognostic value between well-differentiated and poorly differentiated groups, the presence of negative samples in well-differentiated group can be explained by the presence of an effective immune response, while the increase in expression in poorly differentiated group indicates that cancer cells use the *PD-1* pathway as an immune escape mechanism, and Creates an immune-resistant environment [15]. The expression of *PD-L1* in cells Cancer in OSCC ranged from 10 to 90% of cases [40, 41]; In our study, the tumor cells showed positive expression for *PDL-1* in the all samples,

between 1 and 98% in various histological grades, except two negative cases in the well-differentiated group without any statistically significant differences with the histological grades. The expression was heterogeneous. For *PDL-1/TILS* expression, we found statistically significant differences, where the expression was highest in the poorly differentiated group. We may explain that T cells during tumor growth are constantly stimulated by tumor antigens which lead to T cell exhaustion [42], which express immunosuppressive receptors such as *PD-1* and its ligand *PDL-1* [43]. The study of Lechner [44] showed increased expression of *PD-L1* in tumor-infiltrating lymphocytes in OSCC. These results indicate that *PD-L1* that the expression of *PD-L1* in immune cells may be of equal or even greater biological importance than the expression in tumor cells, and this was suggested by a study by Gibbons Johnson [45]. In the study of Kim [46] on 402 patients with head and neck cancers, 204 of them OSCC. That study found that the *PD-L1* expression in TILS, not in tumor cells, has a better prognosis. Adamski [47] studied *PDL-1* expression in 159 patients with OSCC in the tongue and floor of the mouth. The expression of *PD-L1/TC* or *PDL-1/TILS* was not associated with age, gender, clinical stage, histological grade, or smoking and alcohol habits [1]. This is similar to our results in this study; that we found that the expression of *PD-1*, *PD-L1/TILS* and *PDL-1/TC* was not associated with gender, clinical stage, or smoking, except for the association of *PD-L1/TILS* with poorly differentiated cancer, and the association of expression of *PD-L1/TILS* and *PDL-1/TC* with ages older than 65 years, and we explain this by weak host immunity in the elderly, which may be affected by chronic diseases, with the presence of chronic infections, which may activate immunosuppressive signals, most notably *PD-L1*, and this contradicts the Mortin [48]; He compared the expression of *PD-L1* in 175 cases of OSCC ,There was an association between *PD-L1* expression with clinical stage, and age. In the study of Ahn, which was conducted on 68 patients with OSCC, he found no correlation between *PD-L1* expression *PD-L1* expression and clinical characteristics, while *PD-1+* expression was associated with older patients, and with advanced clinical stage, as well as in the study of Kouketsu ; He compared the expression of *PD-1* and *PDL-1* between patients with OSCC (106 samples) and premalignant lesions (79 samples), and found that positive expression of *PDL-1* in OSCC was higher without differences with histological grades. the expression of *PDL-1* and *PD-1* were associated

with tumor size, advanced clinical stage, lymphocytic infiltration, and depth of invasion [49]. and we agreed with his study only with the association of *PDL-1* expression and the depth of invasion. we explain our results by tumor development as a result of overexpressing *PDL-1* in tumor cells, thus suppressing immunity, and promoting tumor growth.

In our study, the expression of *PDL-1*/TC and the expression of *PDL-1*/TILS was not associated with gender, and this is consistent with the study of Admasi [47]. In contrast, some studies found a correlation between *PDL-1* expression with females, such as the Hanna study. [50], as well as Satgunaseelan found an association of *PD-L1* expression with females and these studies explained their results to higher estrogen levels after menopause [6]. In contrast, Lin found an association of *PD-L1* expression in males and smokers [51].

The association of *PDL-1* expression in tumor cells with T cell expression has been evaluated in several studies such as Sales de Sá's study [9]; The expression of *PDL-1* was associated with the expression of CD3+ and CD8+, and was not associated with the expression of CD4+. Takahashi also found that patients with high expression of *PD-L1* combined with high infiltration of CD4+ had better outcomes than cases in which infiltration of CD4+ was low [52]. However, in the study of Lequerica and her colleagues, they showed that high infiltration of CD4+ and CD8+ and CD8+/FOXP3+ were associated with higher positive expression of *PD-L1* on cancer cells [40]. In a study by Cho [53] *PDL-1* expression was associated with low CD8+ expression. but in the study of Mattox [54] they found no correlation between the expression of *PDL-1* and the expression of CD4, and they found a correlation with the expression of CD8, Conversely in our study there was no correlation between the expression of *PDL-1* in tumor cells and the expression of CD8+ and CD4 while we found a correlation between the expression of *PDL-1* in TILS and the expression of CD4+. This can be explained by the fact that CD4 T cells are composed of heterogeneous subsets that perform functions. Promoting or anti-tumor depending on the secreted cytokines [55, 56]. And the expression of *PDL-1*/ TILS was associated with poorly differentiated group, we infer from this the predominance of the regulatory pattern of T cells [57]. We also note that all previous studies focused on the expression of PDL-1 in tumor cells with T cells without mentioning its expression in TILS, so there are not enough studies for comparison.

In conclusion, we conclude that *PDL-1*/TILS, *PDL-1*/TC are independent prognostic factors in OSCC and *PDL-1*/TILS has an important role, since it was associated to the histological grade and CD4 infiltration.

Author Contribution Statement

Collecting data, research and writing the manuscript: Hanan Wahbi. Manuscript revision: Ahmad AL Manadili.

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Approval

It was approved by scientific committee, Faculty of dentistry, Damascus university Study Registration: number 2650 on the date of 2/8/2021, Scientific Research Committee /Damascus university

Ethical Declaration

The authors declare that they have no conflict of interest

Conflict of Interest

All patients have consented to all clinical data included in this study according to the ethical approval of the research (number 2650 on the date of 2/8/2021, Scientific Research Committee /Damascus university)

References

- Ahn H, Yang JM, Kim H, Chung JH, Ahn SH, Jeong WJ, Paik JH. Clinicopathologic implications of the miR-197/*PD-L1* axis in oral squamous cell carcinoma. *Oncotarget*. 2017;8(39):66178–66194. <https://doi.org/10.18632/oncotarget.19842>
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin*. 2021;71(3):209–249. <https://doi.org/10.3322/caac.21660>
- Wu J, Zhang T, Xiong H, Zeng L, Wang Z, Peng Y, et al. Tumor-Infiltrating CD4(+) Central Memory T Cells Correlated with Favorable Prognosis in Oral Squamous Cell Carcinoma. *J Inflamm Res*. 2022;15:141-152. <https://doi.org/10.2147/JIR.S343432>
- Jayasooriya PR, Pitakotuwege TN, Mendis BR, Lombardi T. Descriptive study of 896 Oral squamous cell carcinomas from the only University based Oral Pathology Diagnostic Service in Sri Lanka. *BMC Oral Health*. 2016;16:1. <https://doi.org/10.1186/s12903-015-0139-y>
- Ng JH, Iyer NG, Tan MH, Edgren G. Changing epidemiology of oral squamous cell carcinoma of the tongue: A global study. *Head Neck*. 2017;39(2):297–304. <https://doi.org/10.1002/hed.24589>
- Satgunaseelan L, Gupta R, Madore J, Chia N, Lum T, Palme CE, et al. Programmed cell death-ligand 1 expression in oral squamous cell carcinoma is associated with an inflammatory phenotype. *Pathology*. 2016;48(6):574–580. <https://doi.org/10.1016/j.pathol.2016.07.003>
- Alves A, Diel L, Ramos G, Pinto A, Bernardi L, Yates J, Lamers M. Tumor microenvironment and Oral Squamous Cell Carcinoma: A crosstalk between the inflammatory state and tumor cell migration. *Oral Oncol*. 2021;112:105038. <https://doi.org/10.1016/j.oraloncology.2020.105038>
- Mao X, Xu J, Wang W, Liang C, Hua J, Liu J, et al. Crosstalk between cancer-associated fibroblasts and immune cells in the tumor microenvironment: new findings and future perspectives. *Molecular Cancer*. 2021;20(1):131. <https://doi.org/10.1186/s12943-021-01428-1>
- Sales de Sá R, Miranda Galvis M, Mariz BALA, Leite AA, Schultz L, Almeida OP, et al. Increased Tumor Immune Microenvironment CD3+ and CD20+ Lymphocytes Predict a Better Prognosis in Oral Tongue Squamous Cell Carcinoma. *Front Cell Dev Biol*. 2021;8:622161. <https://doi.org/10.3389/fcell.2021.622161>

- doi.org/10.3389/fcell.2020.622161
10. Wei G, Zhang H, Zhao H, Wang J, Wu N, Li L, et al. Emerging immune checkpoints in the tumor microenvironment: Implications for cancer immunotherapy. *Cancer Lett*;511:68–76. <https://doi.org/10.1016/j.canlet.2021.04.021>
 11. Salmaninejad A, Khoramshahi V, Azani A, Soltaninejad E, Aslani S, Zamani MR, et al. *PD-1* and cancer: molecular mechanisms and polymorphisms. *Immunogenetics*;70(2):73–86. <https://doi.org/10.1007/s00251-017-1015-5>
 12. Han Y, Liu D, Li L. *PD-1/PD-L1* pathway: current researches in cancer. *Am J Cancer Res*. 2020;10(3):727–742.
 13. Yi M, Niu M, Xu L, Luo S, Wu K. Regulation of *PD-L1* expression in the tumor microenvironment. *J Hematol Oncol*. 2021;14(1):10. <https://doi.org/10.1186/s13045-020-01027-5>
 14. Jiang Y, Chen M, Nie H, Yuan Y. *PD-1* and *PD-L1* in cancer immunotherapy: clinical implications and future considerations. *Hum Vaccines Immunother*. 2019;15(5):1111–1122. <https://doi.org/10.1080/21645515.2019.1571892>
 15. Ramsay AG. Immune checkpoint blockade immunotherapy to activate anti-tumour T-cell immunity. *Br J Haematol*. 2013;162(3):313–325. <https://doi.org/10.1111/bjh.12380>
 16. Ibañez-Vega J, Vilchez C, Jimenez K, Guevara C, Burgos PI, Naves R. Cellular and molecular regulation of the programmed death-1/programmed death ligand system and its role in multiple sclerosis and other autoimmune diseases. *J Autoimmun*. 2021;123:102702. <https://doi.org/10.1016/j.jaut.2021.102702>
 17. Gong J, Chehrazhi-Raffle A, Reddi S, Salgia R. Development of *PD-1* and *PD-L1* inhibitors as a form of cancer immunotherapy: a comprehensive review of registration trials and future considerations. *J Immunother Cancer*. 2018;6(1):8. <https://doi.org/10.1186/s40425-018-0316-z>
 18. Sharpe AH, Pauken KE. The diverse functions of the *PD1* inhibitory pathway. *Nat Rev Immunol*. 2018;18(3):153–167. <https://doi.org/10.1038/nri.2017.108>
 19. Yao H, Wang H, Li C, Fang JY, Xu J. Cancer Cell-Intrinsic *PD-1* and Implications in Combinatorial Immunotherapy. *Front Immunol*. 2018;9. <https://doi.org/10.3389/fimmu.2018.01774>
 20. Wang J, Tian S, Sun J, Zhang J, Lin L, Hu C. The presence of tumour-infiltrating lymphocytes (TILs) and the ratios between different subsets serve as prognostic factors in advanced hypopharyngeal squamous cell carcinoma. *BMC cancer*. 2020;20(1):731. <https://doi.org/10.1186/s12885-020-07234-0>
 21. Wang X, Yang L, Huang F, Zhang Q, Liu S, Ma L, You Z. Inflammatory cytokines IL-17 and TNF- α up-regulate *PD-L1* expression in human prostate and colon cancer cells. *Immunol Lett*. 2017;184:7–14. <https://doi.org/10.1016/j.imlet.2017.02.006>
 22. Sanmamed MF, Chen L. Inducible expression of B7-H1 (*PD-L1*) and its selective role in tumor site immune modulation. *Cancer J*. 2014;20(4):256–261. <https://doi.org/10.1097/PPO.0000000000000061>
 23. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer*. 2012;12(4):252–264. <https://doi.org/10.1038/nrc3239>
 24. Sun Z, Fourcade J, Pagliano O, Chauvin JM, Sander C, Kirkwood JM, Zarour HM. IL10 and *PD-1* Cooperate to Limit the Activity of Tumor-Specific *CD8+* T Cells. *Cancer Res*. 2015;75(8):1635–1644. <https://doi.org/10.1158/0008-5472.CAN-14-3016>
 25. Butte MJ, Keir ME, Phamduy TB, Sharpe AH, Freeman GJ. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity*. 2007;27(1):111–122. <https://doi.org/10.1016/j.immuni.2007.05.016>
 26. El-Naggar AK, Chan JKC, Takata T, Grandis JR, Sliotweg PJ. The fourth edition of the head and neck World Health Organization blue book: editors' perspectives. *Hum Pathol*. 2017;66:10–12. <https://doi.org/10.1016/j.humpath.2017.05.014>
 27. Almagush A, Mäkitie AA, Triantafyllou A, de Bree R, Strojanc P, Rinaldo A, et al. Staging and grading of oral squamous cell carcinoma: An update. *Oral Oncol*. 2020;107:104799. <https://doi.org/10.1016/j.oraloncology.2020.104799>
 28. Ngamphaiboon N, Chureemas T, Siripoon T, Arsa L, Trachu N, Jiarpinitnun C, et al. Characteristics and impact of programmed death-ligand 1 expression, *CD8+* tumor-infiltrating lymphocytes, and p16 status in head and neck squamous cell carcinoma. *Med Oncol*. 2019;36(2):21. <https://doi.org/10.1007/s12032-018-1241-1>
 29. Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-*PD-L1* antibody MPDL3280A in cancer patients. *Nature*. 2014;515(7528):563–567. <https://doi.org/10.1038/nature14011>
 30. De Meulenaere A, Vermassen T, Aspeslagh S, Huvenne W, Van J, Dorpe, Ferdinande L, Rottey S. Turning the tide: Clinical utility of *PD-L1* expression in squamous cell carcinoma of the head and neck. *Oral Oncol*. 2017;70:34–42. <https://doi.org/10.1016/j.oraloncology.2017.05.002>
 31. Ghiringhelli F, Bibeau F, Greillier L, Fumet JD, Ilie A, Monville F, Laugé C, et al. Immunoscore immune checkpoint using spatial quantitative analysis of *CD8* and *PD-L1* markers is predictive of the efficacy of anti- *PD1/PD-L1* immunotherapy in non-small cell lung cancer. *EBioMedicine*. 2023;92:104633. <https://doi.org/10.1016/j.ebiom.2023.104633>
 32. Bardhan K, Anagnostou T, Boussiotis VA. The *PD1:PD-L1/2* Pathway from Discovery to Clinical Implementation. *Front Immunol*. 2016;7:550. <https://doi.org/10.3389/fimmu.2016.00550>
 33. Wahbi HS, Manadili A. Immunoexpression of Tumor Infiltrating Lymphocytes(TILs) *CD4+* and *CD8+* in Oral Squamous Cell Carcinoma (OSCC) in Correlations with Clinicopathological Characteristics and Prognosis. *Asian Pac J Cancer Prev*. 2022;23(12):4177–4183. <https://doi.org/10.31557/APJCP.2022.23.12.4177>
 34. Kleffel S, Posch C, Barthel SR, Mueller H, Schlapbach C, Guenova E, et al. Melanoma Cell-Intrinsic *PD-1* Receptor Functions Promote Tumor Growth. *Cell*. 2015;162(6):1242–1256. <https://doi.org/10.1016/j.cell.2015.08.052>
 35. Schatton T, Schütte U, Frank NY, Zhan Q, Hoerning A, Robles SC, et al. Modulation of T-cell activation by malignant melanoma initiating cells. *Cancer Res*. 2010;70(2):697–708. <https://doi.org/10.1158/0008-5472.CAN-09-1592>
 36. Aceto N, Sausgruber N, Brinkhaus H, Gaidatzis D, Martiny-Baron G, Mazarrol G, et al. Tyrosine phosphatase SHP2 promotes breast cancer progression and maintains tumor-initiating cells via activation of key transcription factors and a positive feedback signaling loop. *Nat Med*. 2012;18(4):529–537. <https://doi.org/10.1038/nm.2645>
 37. Li H, Li X, Liu S, Guo L, Zhang B, Zhang J, Ye Q. Programmed cell death-1 (*PD-1*) checkpoint blockade in combination with a mammalian target of rapamycin inhibitor restrains hepatocellular carcinoma growth induced by hepatoma cell-intrinsic *PD-1*. *Hepatology*. 2017;66(6):1920–1933. <https://doi.org/10.1002/hep.29360>
 38. Du S, McCall N, Park K, Guan Q, Fontina P, Ertel A, et al. Blockade of Tumor-Expressed *PD-1* promotes lung cancer growth. *Oncoimmunology*. 2018;7(4):e1408747. <https://doi.org/10.1080/21624022.2018.1408747>

- org/10.1080/2162402X.2017.1408747
39. Maruse Y, Kawano S, Jinno T, Matsubara R, Goto Y, Kaneko N, Sakamoto T, et al. Significant association of increased *PD-L1* and *PD-1* expression with nodal metastasis and a poor prognosis in oral squamous cell carcinoma. *Int J Oral Maxillofac Surg.* 2018;47(7):836–845. <https://doi.org/10.1016/j.ijom.2018.01.004>
 40. Lequerica-Fernández P, Suárez-Canto J, Rodríguez-Santamarta T, Rodrigo JP, Suárez-Sánchez FJ, Blanco-Lorenzo V, et al. Prognostic Relevance of *CD4+*, *CD8+* and *FOXP3+* TILs in Oral Squamous Cell Carcinoma and Correlations with *PD-L1* and Cancer Stem Cell Markers. *Biomedicines.* 2021;9(6):653. <https://doi.org/10.3390/biomedicines9060653>
 41. Oliveira-Costa JP, de Carvalho AF, da Silveira daGG, Amaya P, Wu Y, Park KJ, et al. Gene expression patterns through oral squamous cell carcinoma development: *PD-L1* expression in primary tumor and circulating tumor cells. *Oncotarget.* 2015;6(25):20902. <https://doi.org/10.18632/oncotarget.3939>
 42. Philip M, Schietinger A. *CD8+* T cell differentiation and dysfunction in cancer. *Nat Rev Immunol.* 2022;22:209–223. <https://doi.org/10.1038/s41577-021-00574-3>
 43. Philip M, Fairchild L, Sun L, Horste EL, Camara S, Shakiba M, et al. Chromatin states define tumour-specific T cell dysfunction and reprogramming. *Nature.* 2017;545(7655):452–456. <https://doi.org/10.1038/nature22367>
 44. Lechner A, Schlößer H, Rothschild SI, Thelen M, Reuter S, Zentis P, et al. Characterization of tumor-associated T-lymphocyte subsets and immune checkpoint molecules in head and neck squamous cell carcinoma. *Oncotarget.* 2017;8(27):44418–44433. <https://doi.org/10.18632/oncotarget.17901>
 45. Gibbons Johnson RM, Dong H. Functional Expression of Programmed Death-Ligand 1 (B7-H1) by Immune Cells and Tumor Cells. *Front Immunol.* 2017;8:961. <https://doi.org/10.3389/fimmu.2017.00961>
 46. Kim HR, Ha SJ, Hong MH, Heo SJ, Koh YW, Choi EC, et al. *PD-L1* expression on immune cells, but not on tumor cells, is a favorable prognostic factor for head and neck cancer patients. *Scientific Rep.* 2016;6:36956. <https://doi.org/10.1038/srep36956>
 47. Adamski LJ, Starzyńska A, Adamska P, Kunc M, Sakowicz-Burkiewicz M, Marvaso G, et al. High *PD-L1* Expression on Tumor Cells Indicates Worse Overall Survival in Advanced Oral Squamous Cell Carcinomas of the Tongue and the Floor of the Mouth but Not in Other Oral Compartments. *Biomedicines.* 2021;9(9):1132. <https://doi.org/10.3390/biomedicines9091132>
 48. Moratin J, Metzger K, Safaltin A, Herpel E, Hoffmann J, Freier K, et al. Upregulation of *PD-L1* and *PD-L2* in neck node metastases of head and neck squamous cell carcinoma. *Head Neck.* 2018;41(8):2484–2491. <https://doi.org/10.1002/hed.25713>
 49. Kouketsu A, Sato I, Oikawa M, Shimizu Y, Saito H, Takahashi T, Kumamoto H. Expression of immunoregulatory molecules *PD-L1* and *PD-1* in oral cancer and precancerous lesions: A cohort study of Japanese patients. *J Craniomaxillofac Surg.* 2019;47(1):33–40. <https://doi.org/10.1016/j.jems.2017.04.013>
 50. Hanna GJ, Woo SB, Li YY, Barletta JA, Hammerman PS, Lorch JH. Tumor *PD-L1* expression is associated with improved survival and lower recurrence risk in young women with oral cavity squamous cell carcinoma. *Int J Oral Maxillofac Surg.* 2018;47(5):568–577. <https://doi.org/10.1016/j.ijom.2017.09.006>
 51. Lin YM, Sung WW, Hsieh MJ, Tsai SC, Lai HW, Yang SM. High *PD-L1* Expression Correlates with Metastasis and Poor Prognosis in Oral Squamous Cell Carcinoma. *PLoS One.* 2015;10(11):e0142656. <https://doi.org/10.1371/journal.pone.0142656>
 52. Takahashi H, Sakakura K, Arisaka Y, Tokue A, Kaira K, Tada H, et al. Clinical and Biological Significance of *PD-L1* Expression Within the Tumor Microenvironment of Oral Squamous Cell Carcinoma. *AntiCancer Res.* 2019;39:3039–3046. <https://doi.org/10.21873/anticancer.13437>
 53. Cho YA, Yoon HJ, Lee JI, Hong SP, Hong SD. Relationship between the expressions of *PD-L1* and tumor-infiltrating lymphocytes in oral squamous cell carcinoma. *Oral Oncol.* 2011;47(12):1148–53. <https://doi.org/10.1016/j.oraloncology.08.007>
 54. Mattox AK, Lee J, Westra WH, Pierce RH, Ghossein R, Faquin WC, Diefenbach TJ, et al. *PD-1* Expression in Head and Neck Squamous Cell Carcinomas Derives Primarily from Functionally Anergic *CD4+* TILs in the Presence of *PD-L1+* TAMs. *Cancer Res.* 2017;77(22):6365–6374. <https://doi.org/10.1158/0008-5472.CAN-16-3453>
 55. Huang W, August A. The signaling symphony: T cell receptor tunes cytokine-mediated T cell differentiation. *J Leukoc Biol.* 2015;97(3):477–485. <https://doi.org/10.1189/jlb.1RI0614-293>
 56. Courtney AH, Lo WL, Weiss A. TCR Signaling: Mechanisms of Initiation and Propagation. *Trends Biochem Sci.* 2017;43(2):108–123. <https://doi.org/10.1016/j.tibs.11.008>
 57. Alonso R, Flament H, Lemoine S, Sedlik C, Bottasso E, Péguillet I, Prémel V, et al. Induction of anergic or regulatory tumor-specific *CD4+* T cells in the tumor-draining lymph node. *Nat Commun.* 2018;9(1):2113. <https://doi.org/10.1038/s41467-018-04524-x>



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