Meta-Analysis of Prognostic Significance of Cancer Stem Cell Markers in Oral Squamous Cell Carcinoma

Abhilasha Tripathi¹, Mohit Singh², Prabhaker Mishra³, Naseem Fatima⁴, Vijay Kumar^{2*}

Abstract

Objective: Researchers have shown significant interest in cancer stem cells in recent years. *CD44, CD24, CD133*, and *ALDH* serve as indicators of cancer stem cell-like cells in Oral Squamous Cell Carcinoma. However, the prognostic significance of these Cancer Stem Cell markers in Squamous Cell Carcinoma is still debated. This study employed metaanalysis to evaluate the prognostic significance of cancer stem cells about Oral Squamous Cell Carcinoma. **Methods:** *CD44, CD133, CD24*, and *ALDH* markers were analyzed in 19 retrospective studies to determine their relationship with prognosis and clinicopathological parameters. Risk ratios (RRs) and odds ratios (ORs) were calculated for 3-years survival rates and clinicopathological parameters, respectively, using a fixed-effects model. **Result:** The finding of our study based on extracted survival rates showed that cancer stem cell markers, *CD133* expression was related with the poor prognosis (RR= 1.62, 95% CI = 1.08-2.44, P= 0.02). *ALDH* expression significantly correlated with lymph node metastasis (OR= 4.13, 95% CI= 1.88-9.10, P<0.001) and clinical staging (OR= 2.26, 95% CI= 1.05-4.88, P= 0.04). **Conclusion:** The findings indicate that CSC markers could be used to predict oral cancer prognosis. Our study contributes to the literature on survival outcomes of Oral Squamous Cell Carcinoma. These findings offer a structure for the advancement of cancer treatments that specifically target cancer stem cells. Conducting additional studies with a broader group of patients will help confirm the role of cancer stem cells as dependable predictors of prognosis.

Keywords: oral squamous cell carcinoma- cancer stem cell marker- prognosis- clinicopathological parameters

Asian Pac J Cancer Prev, 25 (10), 3597-3607

Introduction

Oral squamous cell carcinoma (OSCC) ranks as the sixth most prevalent malignancy globally [1]. The global incidence of oral cancer in 2020 was expected to be 377,713 cases, resulting in 177,757 deaths [2]. There has been a continuous increase in new cases of OSCC worldwide every year. Oral cavity cancer is quite prevalent in India, representing approximately 30% of the total global cases [3].

Smoking and alcohol consumption are the primary risk factors for oral cancer in developed countries, while chewing tobacco is the predominant cause of oral cancer in South Asia, particularly India [4]. Conventionally, lymph node metastasis (number and size of lymph nodes and extra nodal extension) is the furthermost significant prognostic factor, resulting in a 50% decrease in survival [5]. Additional characteristics that influence the prognosis include the size of the tumor, the degree of differentiation, the presence of perineural invasion (PNI), and lymphovascular invasion (LVI) in the initial lesion [6, 7]. Over the years, improvements in surgery, radiotherapy techniques, new chemotherapeutic agents, and monoclonal antibodies have improved the prognosis of this cancer.

Despite current treatment modalities, the prognosis for OSCC remains poor, as more than half of the patients die within five years [8]. A subset of patients with earlystage disease has poor outcomes, and some advanced stages of cancer have relatively better outcomes in terms of recurrence and survival. A small minority of patients do not exhibit a response to chemotherapy and radiotherapy, or they acquire resistance to these treatments. CSC exhibits strong tumorigenicity, self-renewal, and differentiation capabilities which maintain cancer. Additionally, these cells are capable of drug resistance and immune evasion. Owing to their rapid immunophenotypic changes, CSCs are difficult to identify using specific markers in tissue and blood [9]. These cells may serve as important targets for the oral cancer therapy, considering their critical roles in tumor induction, progression & metastasis [10].

Although the treatment of oral cancer has been extensively investigated, some questions remain

¹Department of Pharmacology, King George's Medical University, Lucknow, India. ²Department of Surgical Oncology, King George's Medical University, Lucknow, Uttar Pradesh, India. ³Department of Biostatistics and Health Informatics, Sanjay Gandhi Postgraduate Institute of Medical Sciences, Lucknow, Uttar Pradesh, India. ⁴Department of Biochemistry, Era's Lucknow Medical College & Hospital, Lucknow, Uttar Pradesh, India. *For Correspondence: drvsonco75@gmail.com

Abhilasha Tripathi et al

unanswered. It is challenging to predict OSCC behavior using standard clinical and histological indicators. CSC is one such area that has recently received attention from researchers and is believed to be an important driver of tumor biology, which can predict metastasis, recurrence, and survival. There has been growing interest in these CSC markers for oral cancer in recent years, such as *ALDH*, *CD133*, *CD44*, and *CD24* [11].

Therefore, the present study targeted to identify the association amongst CSC marker expression (*CD133*, *CD24*, *CD44*, and *ALDH*), survival and tumor characteristics. This meta-analysis is expected to provide insights into cancer stem cells as promising prognosticators and quantitatively explore which of these CSC markers are more clinically relevant.

Materials and Methods

Data sources and search strategy

A literature search of studies on samples of human subjects was conducted through online databases, including PubMed, ClinicalTrials.gov, Science Direct, Embase and Scopus, using a web-based search in Google Scholar until the 1st of August 2021. Two independent authors (AT and MS) separately searched titles and abstracts based on predefined inclusion criteria to identify eligible studies. The search was run using both MeSH thesaurus and the combination of the following main keywords: "Oral Squamous Cell Carcinoma," "CD24", "Prognosis," and "Survival." The search strategy is summarized in the figure (Figure S1). Preferred Reporting Items for Systematic Reviews and Meta-analysis (PRISMA) were followed to select studies for the proposed meta-analysis.

Criteria for inclusion and exclusion of articles

The studies incorporated in this meta-analysis fulfilled the subsequent inclusion criteria : [i] original articles published in English with full texts available;[ii] patients should be histologically confirmed cases of OSCC;[iii] the expression of oral cancer CSC markers should be detected in a primary cell or surgically dissected primary tumour using immunohistochemistry (IHC) or fluorescence-activated single-cell sorting (FACS);[iv] studies demonstrating the relationship of expression of ALDH, CD133, CD24, and CD44 with survival, i.e., overall survival (OS), disease-free survival (DFS), and clinical and pathological parameter of OSCC;[v] Studies should provide sufficient data for the calculation of risk ratio (RR), odds ratio (OR), disease-free survival (DFS), hazard ratio, or overall survival at 95% confidence interval (CI). The exclusion criteria were as follows: [i] studies involving animal specimens or other cell lines; [ii] reviews, meta-analyses, systematic reviews, case reports, letters, editorials, book chapters, conference abstracts, or expert opinions; [iii] studies not focusing on the risk of OSCC; [iv] duplicate publications; [v] studies not related to the topic of interest; and [vi] studies that did not provide enough information about prognosis.

Data extraction

Literature review table for the initial screening of articles was prepared and discrepancies were resolved through mutual discussion. Two authors (AT and PM) isolated the prognostic and tumour characteristic data from relevant literature and compiled them in a table. For each of the included studies, the following descriptive data were collected: author name, year of publication, period of sample collection, sample size, disease specified, cancer stem cell markers included, the technique used for isolation of markers, clinicopathological features measured, survival data (Kaplan-Meier curves), and the total number of cancer stem cell marker-positive and negative populations. PRISMA was used for the study selection process (Figure 1). Using the criteria for inclusion and exclusion, a comprehensive evaluation was conducted on a total of 88 papers that were deemed relevant. Ultimately, a total of 19 papers were incorporated into the meta-analysis. All included studies had an observational study design. Web Plot Digitizer software v.4.4 was used to extract data from KM curves. The 3-year survival rate was calculated by digitizing the Kaplan-Meier (KM) curves reported in each study. The extracted data were statistically analyzed. Events were calculated based on the survival rates obtained. PM, MS, and AT cross-checked all extracted data.

Reporting quality assessment

The assessment of the reporting quality of all eligible studies was conducted using the Reporting Recommendations for Tumor Marker Prognostic Studies (REMARK) criteria (Table S1). Articles with a reporting quality score of six or higher were considered high-quality studies and included in a meta-analysis. Disagreements were resolved by discussion.

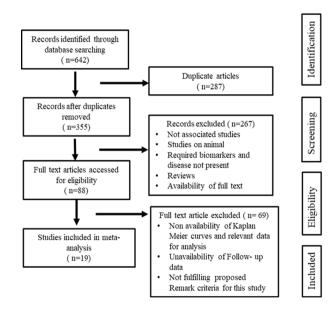


Figure 1. 642 Records Identified through Database Searching. After removing duplicates, 355 records were screened, and 267 were excluded. 88 full-text articles were assessed, with 69 excluded. 19 studies were included in the meta-analysis.

Statistical analysis

The primary endpoint of this study was the relationship with CSC marker expression and Survival (OS and DFS) in subjects with OSCC. Secondary endpoints included the association between tumour characteristics (tumour differentiation, lymph node metastases, clinical TNM staging, and tumour size) and CSC marker expression. Review Manager software v.5.4.1 was used to calculate the forest plots. The heterogeneity of the odds ratios (ORs) for tumor characteristics and relative risks (RRs) for survival rates was determined using a fixed-effects model at a 95% confidence interval, and a random-effects model was processed when the heterogeneity was significant. The threshold for statistical significance was established at a p-value of less than 0.05. The Egger test with a funnel plot was conducted to evaluate publication bias using JASP software v.0.14.1. Furthermore, a p-value of less than 0.10 indicated publication bias and asymmetry.

Results

Study characteristics

Among the 19 research that were part of the metaanalysis, 9 studies were on CD44 [12-20], 6 studies were on ALDH [20-25], 5 studies were on CD24 [19,23,26-28], and 3 studies were on CD133 [16, 29, 30]. All studies were published between 2000 and 2020. The total sample size of the studies was 1410, with a range of 20-150, and the mean was 74.21. Out of the 19 studies, 17 expressed overall survival, one expressed disease-free survival, and one expressed disease-specific survival. Geographically, most studies were conducted in China and India, including four (21.05%) from each country, three from Japan (15.78%), three from Brazil (15.78%), one from the United States of America (5.26%), Germany (5.26%), Iran (5.26%), Sri Lanka (5.26 %), and South Korea (5.26%). Ten studies expressed tumor differentiation (52.6%) and lymph node metastasis (52.6%), five studies expressed clinical TNM staging (26.3%), and four studies expressed tumor size (21.05%). Flow cytometry and immunohistochemistry (IHC) were used to characterize cancer stem cell markers. There were 2 studies on tongue squamous cell carcinoma (TSCC) and 17 studies on OSCC. The characteristics of the studies are displayed in Table 1.

Correlation of cancer stem cell marker expression and tumour characteristics

Lymph node Metastasis

The expression of *CD24* (OR= 1.32, 95% CI= 0.55-3.15, P= 0.53) and *CD44* (OR= 1.19, 95% CI= 0.31-4.60, P= 0.80) was not associated with Lymph node metastasis. Moreover, expression of *ALDH* (OR= 4.13, 95% CI= 1.88-9.10, P< 0.001) was related with the presence of lymph node metastasis (N+) with no heterogeneity. The combined pooled OR of all the studies expressing cancer stem cell markers was not associated (OR=2.02, 95% CI= 0.98-4.15, P= 0.06) with the presence of lymph node metastasis (N+). However, considerable heterogeneity (I²= 74 %, P_h < 0.001) was observed for lymph node metastases (Figure 2a).

Clinical TNM staging

Clinical staging (Stage III+IV) was not associated with CD 44 expression (OR= 1.70, 95% CI= 0.54-5.38, P=0.37). However, considerable heterogeneity (I²= 76%, P_h= 0.005) was observed. In contrast, *ALDH* expression (OR= 2.26, 95% CI= 1.05-4.88, P= 0.04) was considerably related with higher clinical TNM staging (stage III + IV), with non-significant heterogeneity (I²= 0%, P_h= 0.93). Statistically, no association was found among the expression of CSC markers and clinical staging in the combined pooled OR of all the studies expressing CSC markers (OR= 1.81, 95% CI=0.84-3.91, P= 0.13) with a significant heterogeneity (I²= 64%, P_h = 0.02) (Figure 2b).

Tumor Differentiation

The ORs for tumor differentiation for individual cancer stem cell biomarkers are depicted in Figure 3a. The pooled OR for *CD24* (OR= 0.65, 95% CI= 0.11-3.65, P= 0.62), *CD44* (OR= 0.94, 95% CI= 0.40-2.22, P= 0.89), *CD133* (OR= 0.81, 95% CI= 0.17-3.91, P= 0.79) and *ALDH* (OR= 1.93, 95% CI= 0.86-4.34, P= 0.11) showed that their expression was not connected with tumor differentiation. Considerable heterogeneity was observed in the studies for the markers, *CD24* (I²= 76%, P_h= 0.02) and *CD44* (I²= 72%, P_h=0.001) respectively. In the combined pooled OR of the studies, CSC marker expression and tumor differentiation (OR=0.99, 95% CI=0.57-1.69, P= 0.96) were not statistically related, despite of considerable heterogeneity (I²= 63%, P_h= 0.0005).

Tumor size

No significant association was found between *CD24* (OR=0.73, 95% CI=0.29-1.84, P=0.50) and *ALDH* (OR= 1.45, 95% CI=0.58-3.60, P=0.43) expression and tumour size. No heterogeneity was observed between these studies expressing *CD24* and *ALDH* (I²=13 %, P_h=0.28 and I²= 0%, P_h=0.64), respectively. The combined pooled analysis of all studies showed that CSC marker expression was not related with tumor size (OR= 1.04, 95% CI= 0.55-1.95, P=0.91). No significant heterogeneity (I²=0%, P_h=0.50) was observed between the studies related to tumour size (Figure 3b).

Association between the expression of cancer stem cell markers and the overall survival rate after 3 years

The pooled risk ratio (RR) of the studies expressing CD44 was 1.32 (95% CI= 0.81-2.13, P= 0.26), CD24 was 0.83 (95% CI= 0.53-1.28, P= 0.39), CD133 was 1.62 (95% CI= 1.08-2.44, P= 0.02), and *ALDH* was 1.20 (95%) CI= 0.95-1.52, P= 0.12). Accordingly, our observation displayed the over expression of CD44, CD24, and ALDH was not significantly connected with overall survival. Though, a statistically significant association was only observed for CD133 expression. Heterogeneity was significant between studies expressing CD44 (I²= 60%, $P_{\mu} = 0.01$) but not between those expressing CD24 $(I^2 = 52\%, P_h = 0.10), CD133 (I^2 = 56\%, P_h = 0.10), and$ ALDH (I²=48%, P_b=0.10). Although, as per the forest plot illustration, three out of the four markers supported a poor prognosis, CD24 expression was associated with a good prognosis. ALDH, CD44, and CD133 expressions were

(a)

	Experime		Contr			Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% C	I M-H, Random, 95% CI
2.1.1 CD24							
Todoroki et al 2016	2	11	5	39	6.7%	1.51 [0.25, 9.11]	
de Moraes et al 2016	19	36	3	6	6.9%	1.12 [0.20, 6.30]	
Tamatani et al 2018	9	41	5	29	8.6%	1.35 [0.40, 4.55]	
Subtotal (95% CI)		88		74	22.2%	1.32 [0.55, 3.15]	
Total events	30		13				
Heterogeneity: Tau ² = 0.			= 2 (P = 0	.97); l²	= 0%		
Test for overall effect: Z	= 0.63 (P =	0.53)					
2.1.2 CD44							
Todoroki et al 2016	6	23	1	27	5.5%	9.18 [1.01, 83.11]	
de Moraes et al 2016	14	23	8	19	8.5%	2.14 [0.62, 7.37]	
Boxberg et al 2018	5	33	15	56	8.9%	0.49 [0.16, 1.50]	
Lee at al 2015	15	28	8	29	9.0%	3.03 [1.01, 9.12]	
Sato et al 2000	8	53	34	67	9.7%	0.17 [0.07, 0.42]	
Subtotal (95% CI)		160		198	41.6%	1.19 [0.31, 4.60]	
Total events	48		66				
Heterogeneity: Tau ² = 1.	91; Chi² = 2	4.85, d	f = 4 (P <	0.0001); I ² = 84%	6	
Test for overall effect: Z	= 0.26 (P =	0.80)					
2.1.3 ALDH							
Rao et al 2020	20	26	0	14	3.9%	91.46 [4.77, 1754.50]	
de Moraes et al 2016	5	6	17	36	5.4%	5.59 [0.59, 52.73]	
Filho et al 2020	12	16	17	39	8.3%	3.88 [1.06, 14.20]	
Tamatani et al 2018	7	18	7	52	8.5%	4.09 [1.19, 14.11]	
Jayasooriya et al 2016 Subtotal (95% CI)	28	78 144	12	62 203	10.0% 36.2%	2.33 [1.07, 5.10] 4.13 [1.88, 9.10]	
Total events	72	144	53	200	00.270	4.10 [1.00, 0.10]	
Heterogeneity: Tau ² = 0.	• =	26 df		18): l ²	= 36%		
Test for overall effect: Z					00,0		
Total (95% CI)		392		475	100.0%	2.02 [0.98, 4.15]	
Total events	150		132				-
Heterogeneity: Tau ² = 1.		5.67, d		< 0.000	001); l ² = 7	4%	
Test for overall effect: Z			(0.000	. ,, .		
Test for subgroup differe	•	,	df = 2 (P :	= 0.10)	$I^2 = 56.1^{\circ}$	%	Favours [Negative] Favours [Positive]
(1)							
(b)	Experim	ental	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events		Events	Total	Weight	M-H, Random, 95% CI	M-H, Random, 95% CI
1.1.1 CD44	LYCING	10141			mangint		

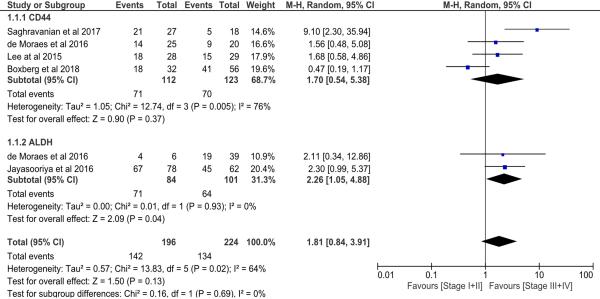


Figure 2. The Forest Plot of OR was assessed for the association between Cancer Stem Cell Marker and Tumour Characteristics, Analysis of *CD 24, CD44*, and *ALDH* expression and Lymphnode metastasis in OSCC patients (a); Analysis of *CD44*, and *ALDH* expression and clinical staging in OSCC patients (b).

associated with lower overall survival rates (Figure 4).

Assessment of Publication Bias

Egger's regression tests revealed non-significant publication bias for each CSC marker in terms of tumour size; however, in terms of clinical staging and tumour differentiation, *CD44* expression showed a significant publication bias. *CD44* and *ALDH*, in contrast to *CD24*, revealed considerable publication bias in the case of lymph node metastasis (Table S2). Graphically, the funnel plots in the meta-analysis results of 3-year overall survival rates of the studies expressing *CD24*, *CD44*, and *CD133* did not exhibit any asymmetry (Figure S2), and Egger's regression test didn't reveal any significant

		0.5595	1.26[0.58, 2.77]							
7	ND	0.5076	1.33[0.57,3.12]	SO	66	IHC	ALDH, CD44	TSCC	2003-2006	Huang et al. [20]
		0.3321	1.64[0.60, 4.48]							
6	Tumour size, Tumour Differentiation, Lymphnode Metastasis	0.523	0.64[0.17,2.49]	SO	50	FACS, IHC	CD24, CD44	OSCC	2007-2008	Todoroki et al. [19]
6	ND	0.4528	0.67[0.23,1.92]	SO	20	FACS, IHC	CD24	OSCC	ND	Ghuwalewala et al. [27]
7	Tumour Differentiation	0.2452	0.19[0.01,3.11]	SO	53	FACS	CD44	OSCC	2010-2013	Mohanta et al. [17]
7	ND	0.0929	0.08[0.00,1.54]	SO	144	IHC	CD133	OSCC	2000-2010	Wang et al. [29]
7	Tumour Differentiation, Lymphnode Metastasis, Clinical TNM staging	0.1922	2.29[0.66,7.92]	SO	57	IHC	CD44	OSCC	2004-2014	Lee et al. [18]
		0.0214	1.87[1.10,3.17]							
8	ND	0.0852	1.75[0.93, 3.31]	SO	150	IHC	CD44, CD133	OSCC	1990-2009	Oliveria et al. [16]
6	ND	0.4308	0.90[0.70, 1.16]	SO	78	IHC	ALDH	OSCC	2008-2010	Wu et al. [24]
6	Tumour size, Tumour Differentiation, Lymphnode Metastasis	0.4718	1.29[0.65,2.55]	SO	63	IHC	ALDH	OSCC	1970-2013	Filho et al. [25]
7	Clinical TNM staging, Lymphnode Metastasis	0.3414	1.26[0.79,2.00]	SO	140	IHC	ALDH	OSCC	2009-2012	Jayasooriya et al. [21]
7	Tumour Differentiation, Lymphnode Metastasis, Clinical TNM staging	NA	NA	DSS	52	IHC	CD44	OSCC	2000-2005	deMoraes et al. [15]
6	Tumour Differentiation, Lymphnode Metastasis	0.0077	0.25[0.09,0.07]	SO	120	IHC	CD44	TSCC	ND	Sato et al. [14]
6	Tumour Differentiation, Clinical TNM staging	0.0365	2.67[1.06,6.69]	SO	45	IHC	CD44	OSCC	ND	Saghravanian et al. [13]
6	Tumour Differentiation, Lymphnode Metastasis, Clinical TNM staging	0.0252	1.56[1.06,2.29]	SO	108	IHC	CD44	OSCC	2007-2012	Boxberg et al. [12]
6	Tumour size, Tumour Differentiation, Lymphnode Metastasis	NA	NA	DFS	70	IHC	ALDH, CD24	OSCC	ND	Tamtani et al. [23]
6	ND	0.1217	2.17[0.81,5.80]	SO	50	IHC	CD24	OSCC	ND	Mishra et al.[28]
6	ND	0.0364	0.52[0.28, 0.96]	SO	52	ND	CD24	OSCC	ND	Fugle et al. [26]
6	Tumour size, Tumour Differentiation, Lymphnode Metastasis	0.0662	1.98[0.96, 4.10]	SO	52	FACS, IHC	CD133	OSCC	ND	Chiou et al. [30]
8	Lymphnode Metastasis	0.0664	6.00[0.89,40.63]	SO	40	IHC	ALDH	OSCC	ND	Rao et al [22]
Quality Score	Clinicopathological features included	P-Value	RR/OR (95%CI)	Survival measure	Total sample	Technique	Cancer Stem cell marker	Disease	Year of data collection	Reported studies

DOI:10.31557/APJCP.2024.25.10.3597 Prognostic Impact of CSC Markers in OSCC

(a)

	Experime	ental	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% C	M-H, Random, 95% Cl
3.1.1 CD24							
de Moraes et al 2016	16	42	5	6	3.9%	0.12 [0.01, 1.15]	
Todoroki et al 2016	6	11	10	39	6.5%	3.48 [0.87, 13.94]	
Tamatani et al 2018 Subtotal (95% CI)	16	40 93	17	28 73	8.3% 18.7%	0.43 [0.16, 1.16] 0.65 [0.11, 3.65]	
Total events	38		32				
Heterogeneity: Tau² = 1.73 Test for overall effect: Z =			2 (P = 0.0	2); I² =	76%		
3.1.2 CD44							
Mohanta et al 2007	3	8	9	18	5.4%	0.60 [0.11, 3.30]	
Saghravanian et al 2017	26	31	3	14	5.8%	19.07 [3.87, 94.02]	
Boxberg et al 2018	28	33	51	56	6.8%	0.55 [0.15, 2.06]	
Fodoroki et al 2016	6	23	10	27	7.3%	0.60 [0.18, 2.02]	
de Moraes et al 2016	9	30	12	20	7.4%	0.29 [0.09, 0.94]	
Lee at al 2015	19	28	16	29	7.9%	1.72 [0.58, 5.05]	
Sato et al 2000	26	53	42	67	9.5%	0.57 [0.28, 1.19]	
Subtotal (95% CI)		206		231	50.0%	0.94 [0.40, 2.22]	
Fotal events	117		143				
Heterogeneity: Tau² = 0.93 Test for overall effect: Z =		·	0 (1 - 0.	001), 1	- 1270		
4 9 00499							
de Moraes et al 2016	2	3	19	47	3.4%	2.95 [0.25, 34.85]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI)	55	3 105 108	27	47 39 86	3.4% 9.3% 12.7%	2.95 [0.25, 34.85] 0.49 [0.22, 1.07] 0.81 [0.17, 3.91]	
de Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Fotal events	55 57	105 108	27 46	39 86	9.3% 12.7%	0.49 [0.22, 1.07]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Fotal events Heterogeneity: Tau ² = 0.74	55 57 4; Chi² = 1.8	105 108 5, df =	27 46	39 86	9.3% 12.7%	0.49 [0.22, 1.07]	
de Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.7 Fest for overall effect: Z =	55 57 4; Chi² = 1.8	105 108 5, df =	27 46	39 86	9.3% 12.7%	0.49 [0.22, 1.07]	
le Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Fotal events Heterogeneity: Tau ² = 0.74 Fest for overall effect: Z = 8.1.4 ALDH	55 57 4; Chi² = 1.8	105 108 5, df =	27 46	39 86	9.3% 12.7%	0.49 [0.22, 1.07]	
le Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.74 Fest for overall effect: Z = 8.1.4 ALDH Filho et al 2020	55 57 4; Chi² = 1.8 0.27 (P = 0.	105 108 5, df = 79)	27 46 1 (P = 0.1	39 86 7); I² =	9.3% 12.7% 46%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91]	
te Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Fotal events Heterogeneity: Tau ² = 0.74 Fest for overall effect: Z = 8.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Famatani et al 2018	55 57 4; Chi² = 1.8 0.27 (P = 0. 5	105 108 5, df = 79) 7	27 46 1 (P = 0.1 12	39 86 7); I ² = 30	9.3% 12.7% 46% 5.1%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.7. Test for overall effect: Z = 3.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Tamatani et al 2018 Subtotal (95% CI)	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3	105 108 35, df = 79) 7 7 18	27 46 1 (P = 0.1 12 18	39 86 7); l ² = 30 43 50	9.3% 12.7% 46% 5.1% 5.7% 7.8%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01]	
te Moraes et al 2016 Nang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.74 Fest for overall effect: Z = 8.1.4 ALDH Filho et al 2020 te Moraes et al 2016 Famatani et al 2018 Subtotal (95% CI) Fotal events Heterogeneity: Tau ² = 0.00	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3 11 19 0; Chi ² = 1.0	105 108 35, df = 7 79) 7 18 32 19, df = 2	27 46 1 (P = 0.1 12 18 22 52	39 86 7); l ² = 30 43 50 123	9.3% 12.7% 46% 5.1% 5.7% 7.8% 18.6%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.7. Test for overall effect: Z = 3.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Tamatani et al 2018 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.00 Test for overall effect: Z =	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3 11 19 0; Chi ² = 1.0	105 108 35, df = 7 79) 7 18 32 19, df = 2	27 46 1 (P = 0.1 12 18 22 52	39 86 7); l ² = 30 43 50 123 8); l ² =	9.3% 12.7% 46% 5.1% 5.7% 7.8% 18.6%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.74 Test for overall effect: Z = 3.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Tamatani et al 2018 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.00 Test for overall effect: Z = Total (95% CI)	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3 11 19 0; Chi ² = 1.0	105 108 55, df = 5 79) 7 7 18 32 99, df = 5 11)	27 46 1 (P = 0.1 12 18 22 52	39 86 7); l ² = 30 43 50 123 8); l ² =	9.3% 12.7% 46% 5.1% 5.7% 7.8% 18.6%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01] 1.93 [0.86, 4.34]	
de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.74 Test for overall effect: Z = 3.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Tamatani et al 2018 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.00 Test for overall effect: Z = Total (95% CI) Total events	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3 11 19 0; Chi ² = 1.0 1.59 (P = 0. 231	105 108 35, df = 7 79) 7 18 32 99, df = 2 11) 439	27 46 1 (P = 0.1 12 18 22 52 2 (P = 0.5 273	39 86 7); l ² = 30 43 50 123 8); l ² = 513	9.3% 12.7% 46% 5.1% 5.7% 7.8% 18.6% 0%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01] 1.93 [0.86, 4.34]	
3.1.3 CD133 de Moraes et al 2016 Wang et al 2017 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.74 Test for overall effect: Z = 3.1.4 ALDH Filho et al 2020 de Moraes et al 2016 Tamatani et al 2018 Subtotal (95% CI) Total events Heterogeneity: Tau ² = 0.01 Test for overall effect: Z = Total (95% CI) Total events Heterogeneity: Tau ² = 0.61 Total events Heterogeneity: Tau ² = 0.61 Total events Heterogeneity: Tau ² = 0.61 Total events Heterogeneity: Tau ² = 0.61 Test for overall effect: Z =	55 57 4; Chi ² = 1.8 0.27 (P = 0. 5 3 11 19 0; Chi ² = 1.0 1.59 (P = 0. 231 6; Chi ² = 38.	105 108 5, df = 7 7 7 18 32 99, df = 2 11) 439 07, df =	27 46 1 (P = 0.1 12 18 22 52 2 (P = 0.5 273	39 86 7); l ² = 30 43 50 123 8); l ² = 513	9.3% 12.7% 46% 5.1% 5.7% 7.8% 18.6% 0%	0.49 [0.22, 1.07] 0.81 [0.17, 3.91] 3.75 [0.62, 22.58] 1.04 [0.21, 5.24] 2.00 [0.67, 6.01] 1.93 [0.86, 4.34]	0.01 0.1 10 Favours [G1] Favours [G2+G3]

(b)

	Experim	ental	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% C	I M-H, Fixed, 95% CI
5.1.1 CD 24							
Todoroki et al 2016	3	11	8	39	13.6%	1.45 [0.31, 6.76]	
Tamatani et al 2018	29	41	24	29	43.7%	0.50 [0.16, 1.63]	
Subtotal (95% CI)		52		68	57.3%	0.73 [0.29, 1.84]	
Total events	32		32				
Heterogeneity: Chi ² = ¹	1.15, df = 1	(P = 0.2	28); l² = 1	3%			
Test for overall effect:	Z = 0.67 (F	P = 0.50)					
5.1.2 ALDH							
Filho et al 2020	12	16	25	40	19.0%	1.80 [0.49, 6.61]	
Tamatani et al 2018	14	18	39	52	23.7%	1.17 [0.33, 4.18]	
Subtotal (95% CI)		34		92	42.7%	1.45 [0.58, 3.60]	
Total events	26		64				
Heterogeneity: Chi ² = (0.22, df = 1	(P = 0.0	64); I² = 0	%			
Test for overall effect:	Z = 0.80 (F	P = 0.43)					
Total (95% CI)		86		160	100.0%	1.04 [0.55, 1.95]	+
Total events	58		96				
Heterogeneity: Chi ² = 2	2.36, df = 3	(P = 0.	50); I² = 0	%			
Test for overall effect:	Z = 0.11 (F	e = 0.91)					0.01 0.1 1 10 100 Favours [Small] Favours [Large]
Test for subaroup diffe	erences: Ch	i ² = 1.07	7. df = 1 (P = 0.3	0), l ² = 6.9	9%	Tavou's [Omaii] Favou's [Laige]

Figure 3. The Forest Plot of OR was Assessed for the Association between Cancer Stem Cell Marker and Tumour Characteristics, Analysis of *CD 24, CD44, CD 133* and *ALDH* Expression and Tumor Differentiation in OSCC Patients (a); Analysis of CD 24, and ALDH expression and tumor size in OSCC patients (b).

publication bias. In contrast, *ALDH* showed funnel plots with asymmetry and significant publication bias in Egger's

regression test (Table S3).

(a)							
	Case		Contro	ol		Risk Ratio	Risk Ratio
Study or Subgroup	Events T	otal	Events	Total	Weight	M-H, Fixed, 95% C	I M-H, Fixed, 95% Cl
Rao et al 2020	14	28	1	12	2.1%	6.00 [0.89, 40.63]	· · · · · · · · · · · · · · · · · · ·
Huang et al 2014	7	18	7	24	8.9%	1.33 [0.57, 3.12]	
Filho et al 2020	7	16	16	47	12.0%	1.29 [0.65, 2.55]	- +-
Jayasooriya et al 2016	30	78	19	62	31.3%	1.26 [0.79, 2.00]	- -
Wu et al 2017	28	39	31	39	45.8%	0.90 [0.70, 1.16]	†
Total (95% CI)		179		184	100.0%	1.20 [0.95, 1.52]	•
Total events	86		74				
Heterogeneity: Chi ² = 7.7	75, df = 4 (P =	= 0.10); I² = 48	3%			
Test for overall effect: Z	= 1.56 (P = 0	0.12)					0.01 0.1 1 10 100 High Survival Low Survival

(b)	Case	9	Contr	ol		Risk Ratio	Risk Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% CI	M-H, Fixed, 95% CI
Wang et al 2017	0	105	2	39	15.5%	0.08 [0.00, 1.54]	
Chiou et al 2008	19	32	6	20	31.6%	1.98 [0.96, 4.10]	⊢ ∎−-
Oliveria et al 2013	22	39	13	43	52.9%	1.87 [1.10, 3.17]	
Total (95% CI)		176		102	100.0%	1.62 [1.08, 2.44]	•
Total events	41		21				
Heterogeneity: Chi ² = 4	4.53, df =	2 (P = (0.10); l² =	56%			
Test for overall effect:	Z = 2.34 (P = 0.0	2)			0.01	0.1 1 10 100 High Survival Low Survival

(c)	Case	9	Contr	ol		Risk Ratio	Risk Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% CI	M-H, Random, 95%	CI
Mohanta et al 2017	0	8	5	18	2.7%	0.19 [0.01, 3.11]		
Lee at al 2015	16	28	2	8	9.2%	2.29 [0.66, 7.92]		
Sato et al 2000	4	53	20	67	11.6%	0.25 [0.09, 0.70]	_	
Todoroki et al 2016	7	23	5	27	11.7%	1.64 [0.60, 4.48]		
Saghravanian et al 2017	16	27	4	18	12.8%	2.67 [1.06, 6.69]		_
Huang et al 2014	10	26	7	23	14.6%	1.26 [0.58, 2.77]	_	
Oliveria et 2013	9	17	13	43	16.8%	1.75 [0.93, 3.31]	⊢ ∎−-	
Boxberg et al 2018	22	33	24	56	20.7%	1.56 [1.06, 2.29]		
Total (95% CI)		215		260	100.0%	1.32 [0.81, 2.13]	•	
Total events	84		80					
Heterogeneity: Tau ² = 0.25	5; Chi² = 1 [·]	7.50, df	= 7 (P =	0.01); I	² = 60%			10 100
Test for overall effect: Z =	1.12 (P =)	0.26)	,				0.01 0.1 1 High Survival Low Sur	10 100 vival

(d)	Case	Э	Contr	ol		Risk Ratio		Risk Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% C	I	M-H, Fixed, 95% CI	
Ghuwalewala et al 2016	2	4	6	8	13.8%	0.67 [0.23, 1.92]			
Mishra et al 2020	12	29	4	21	16.0%	2.17 [0.81, 5.80]		+	
Todoroki et al 2016	2	11	11	39	16.7%	0.64 [0.17, 2.49]			
Fugle et al 2016	10	31	13	21	53.5%	0.52 [0.28, 0.96]			
Total (95% CI)		75		89	100.0%	0.83 [0.53, 1.28]		•	
Total events	26		34						
Heterogeneity: Chi ² = 6.20), df = 3 (P	9 = 0.10); l ² = 52%	6					100
Test for overall effect: Z =	0.86 (P =	0.39)					0.01	0.1 1 10 High Survival Low Survival	100

Figure 4. Analysis of *ALDH* (a), *CD 133* (b), *CD 44* (c), and *CD24*(d) Expression and Survival of OSCC Patients, Forest Plot of RR was assessed for the association between Cancer Stem Cell Markers and Survival.

Discussion

Recently, several treatment options have been developed for oral cancer, but still, it has poor outcomes.

One of the reasons for metastasis and recurrence of cancer is believed to be CSCs within tumors. Cancer stem cells can be identified using CSC markers to study their influence on tumor formation and growth [9, 31].

Abhilasha Tripathi et al

This meta-analysis investigated the correlation among CSC marker expression and tumour characteristics and their function in predicting the survival of subjects with oral carcinoma. Our study provides the first comprehensive assessment by comparing prevalent CSC markers (*ALDH*, *CD24*, *CD133* and CD 44) in oral cancer. We found an association between these markers and tumor characteristics, suggesting that they have an adverse prognostic impact.

The presence of CD133 is highly correlated with a worse outcome in individuals with oral cancer, consistent with prior well-documented studies [32]. Research indicates that cancer stem cells possess the ability to undergo self-renewal, which is believed to be a contributing element to their ability to form tumors [33]. Some other studies done in colon and lung cancer also showed similar result [34, 35]. Due to their chemotherapeutic resistance, CD133-positive cancer stem-like cells have been hypothesized to be accountable for tumor development and reappearance [36], leading to a poor prognosis. Conversely, one of the included studies, Wang et al. established the CD133 expression was connected with a higher survival rate [29]. These results could be attributed to selection bias because they only included patient population with early-stage oral cancer.

Our investigation found no significant correlation between the expression of CD44, ALDH, and CD24 with survival. CD44 expression has been associated with an unfavorable outcome in many types of cancer [37, 38]. The impact of CD44 expression on the prognosis of oral squamous cell carcinoma (OSCC) is still uncertain, as conflicting findings have been reported [39]. ALDH expression has been linked to worse outcomes in different types of cancers, such as breast [40] and colorectal cancer [41]. However, one study showed similar results to our study [42]. Among the included studies, only one study by Rao et al. showed low survival with the expression of ALDH [22]. However, our findings on CD24 expression are consistent with those of another study on OSCC [16,43]. CD24 expression has been linked to survival in breast cancer [44] and hepatocellular carcinoma [45], whereas our findings contradict these studies. Previous studies have indicated that CD24 has a detrimental impact on cell migration and epithelial-mesenchymal transition. This implies that cells with a CD44 low and CD24 high phenotype exhibit comparable behavior to non-cancer stem cell features [27, 46]. Several laboratory investigations conducted in test tubes and living organisms have demonstrated that low levels of CD24 expression in oral cancer are linked to a negative prognosis [47].

According to previously reported findings, we have confirmed that the expression of *ALDH* is associated with the dissemination of cancer cells to the lymph nodes and a more advanced stage of the disease. Several studies have established a correlation between *ALDH* expression and aggressive tumors, lymph node metastases, and advanced tumor stage [42, 48, 49]. Nevertheless, there was no discernible association observed between the expression of *ALDH* and the size or differentiation of the tumor. Furthermore, Szafarowski et al. found that *CD24* was linked to the spread of cancer to the lymph nodes in head and neck cancer, but it did not have an impact on survival [50]. Owing to inadequate information, *CD24* has not been evaluated for clinical tumor staging. Even though CD 24 has a distinct function in the preservation of cancer stem cells, not many studies evaluate this marker, and publication bias still exists; we believe that *CD24* is not a specific prognostic marker for Oral Cancer.

In our study, CD133 was not connected with tumour differentiation; however, CD133 expression was correlated with poorly differentiated tumors in some studies [51]. Insufficient data was available to study CD133 expression for all tumour characteristics, except for tumour differentiation. Our findings revealed that CD44 expression was not to be related with lymph node metastasis, TNM staging, and tumour differentiation; although some studies were incongruent, most supported these findings. CD44 is associated with tumors differentiation. This discrepancy could be explained as CD44 has two isoforms, CD44v and CD44s, which are involved in cancer's cellular function and pathogenesis [52-54]. In head and neck cancers, CD44v3 levels are elevated, leading to increased cell migration. Low expression of the CD44v9 variant is related to higher survival, according to Sato et al. [14]. In our study, we did not characterize CD44 based on its isoforms. Various CD44 isoforms exhibit different cellular characteristics when co-expressed with other CSC markers [55]. The findings of our investigation indicate that CD44 expression in oral tumour should be studied based on its isoforms and its co-expression with other CSC markers. We were unable to analyze CD44 expression in relation to tumor size because of the lack of relevant studies.

Our results are in line with previous studies with some discrepancies. It is possible that these differences are due to the fact that previous studies included earlystage oral cancer, different subtypes of oral cancer, different antibodies for analysis, and had different patient demographics. According to Forest Plot, CD 24 is the only marker that supports a good prognosis, while CD 133 showed a statistically significant poor prognosis out of the four markers. Expression of all four cancer stem cell markers supports lymph node metastases (N+), higher clinical TNM stages (stage III + IV), large tumor size, and tumor differentiation grade (G2+G3); however, only ALDH showed statistical significance with lymph node metastasis and clinical stage. The sample size was comparatively small, perhaps diminishing the data's ability to identify any pattern. Due to the scarcity of studies that fit our specific criteria, only a small number of papers were included in the current investigation. However, we believe that strict inclusion and exclusion criteria led to high-quality and more relevant results. Finally, the events were calculated using data digitalization of Kaplan-Meier curves, which may have resulted in minor differences with the actual data due to manual errors. Despite this limitation, one of the key strengths of this meta-analysis is that we only included studies which examined cancer stem cell markers in patient derived primary tumour, which made it more clinically relevant. Patient-derived samples can retain heterogeneity and tumour microenvironment, so they have more translational value than the secondary

cell line.

In conclusion, overall, increased research and understanding of CSC markers may allow clinicians to better prognosticate and choose more personalized treatment options for patients with oral cancer. Coexpression of these markers should also be considered so we can get a specific phenotype or panel of markers to understand the prognostic role of CSC markers in oral cancer. Our study contributes to the literatures on survival outcomes and tumor characteristics of patients with OSCC. Further extensive, well-designed cohort studies using primary human tumor cells must be required to explore their role in clinical factors & treatment-related outcomes in oral cancer.

Author Contribution Statement

Abhilasha Tripathi (M. Pharm.) and Dr. Vijay Kumar (MCh)conceptualization; Dr. Prabhaker Mishra (PhD) methodology, software analysis, data interpretation, and statistical analysis; Abhilasha Tripathi, Mohit Singh (M.Sc.) and Dr. Prabhaker Mishra data curation, writingoriginal draft preparation, prepared the figure, charts, and tables; Abhilasha Tripathi and Dr. Naseem Fatima (PhD) writing, reviewing, and editing. Each author contributed to the manuscript and reviewed the final draft before submission.

Acknowledgements

Ethical Consideration/Approval

This meta-analysis, approved by the Institutional Ethics Committee, KGMU, Lucknow (Ref no 1460/ Ethics/R.Cell-16) as part of an approved student thesis, was conducted in accordance with institutional ethical standards and the Declaration of Helsinki. The study involved no patients, public, or animals, and was designed independently by the authors, waiving the need for informed consent.

Data Availability

All data generated or analyzed during this study are included in this article.

Conflict of Interest

The authors declare no potential conflicts of interest.

Abbreviations

References

- Chen YJ, Lin SC, Kao T, Chang CS, Hong PS, Shieh TM, et al. Genome-wide profiling of oral squamous cell carcinoma. J Pathol. 2004;204(3):326–32. https://doi.org/10.1002/ path.1640.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71:209-49. https://doi.org/10.3322/caac.21660.
- 3. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A, et al . Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424. https://doi.org/10.3322/caac.21492.
- Ren ZH, Hu CY, He HR, Li YJ, Lyu J. Global and regional burdens of oral cancer from 1990 to 2017: Results from the global burden of disease study. Cancer Commun (Lond). 2020;40(2-3):81-92. https://doi.org/10.1002/cac2.12009.
- Woolgar JA, Triantafyllou A, Lewis JS Jr, Hunt J, Williams MD, Takes RP, et al. Prognostic biolog- ical features in neck dissection specimens. Eur Arch Otorhinolaryngol. 2013;270(5):1581-92. https://doi.org/10.1007/s00405-012-2170-9.
- Boonpoapichart S, Punyavong P, Jenwitheesuk K, Surakunprapha P, Winaikosol K. Significant prognostic factors influencing the survival difference of oral tongue squamous cell carcinoma. Plast Reconstr Surg Glob Open. 2021;9(10). https://doi.org/10.1097/GOX.00000000003889.
- Montoro JRM, Hicz HA, Souza L, Livingstone D, Melo DH, Tiveron RC, et al. Prognostic factors in squamous cell carcinoma of the oral cavity. Braz J Otorhinolaryngol. 2008;74(6):861-6. https://doi.org/10.1016/S1808-8694(15)30146-4.
- Chaturvedi AK, Anderson WF, Lortet-Tieulent J, Curado MP, Ferlay J, Franceschi S, et al. Worldwide trends in incidence rates for oral cavity and oropharyngeal cancers. J Clin Oncol. 2013;31(36):4550-9. https://doi.org/10.1200/ JCO.2013.50.3870.
- Clevers H. The cancer stem cell: premises, promises and challenges. Nat Med. 2011;17(3) :313-9. https://doi. org/10.1038/nm.2304.
- Agliano A, Calvo A, Box C. The challenge of targeting cancer stem cells to halt metastasis. Semin Cancer Biol. 2017;44:25-42. https://doi.org/10.1016/j.semcancer.2017.03.001.
- Chen D, Wang CY. Targeting cancer stem cells in squamous cell carcinoma. Precis Clin Med. 2019;2(3):152–65. https:// doi.org/10.1093/pcmedi/pbz016.
- 12. Boxberg M, Götz C, Haidari S, Dorfner C, Jesinghaus M, Drecoll E, et al. Immunohistochemical expression of *CD44* in oral squamous cell carcinoma in relation to histomorphological param-eters and clinicopathological factors. Histopathology. 2018;73(4):559-72. https://doi.org/10.1111/ his .13496.
- Saghravanian N, Anvari K, Ghazi N, Memar B, Shahsavari M, Afzal Aghaee M. Expression of p63 and *CD44* in oral squamous cell carcinoma and correlation with clinicopathological parameters. Arch Oral Biol. 2017;82:160-5. https://doi.org/10.1016/j.archoralbio.2017.06.011.
- 14. Sato S, Miyauchi M, Takekoshi T, Zhao M, Kudo Y, Ogawa I, et al. Reduced expression of *CD44* variant 9 is related to lymph node metastasis and poor survival in squamous cell carcinoma of tongue. Oral Oncol. 2000;36(6):545-9. https://doi.org/10.1016/S1368-8375(00)00038-5.
- 15. Prudente de Moraes F, Lourenço SV, Fraga Ianez RC, de Sousa EA, Messias da Conceição Silva M, Santos

Damascena A, et al. Expression of stem cell markers in oral cavity and oropharynx squamous cell carcinoma. Oral Surg Oral Med Oral Pathol Oral Radiol. 2017;123(1):113-22. https://doi.org/10.1016/j.0000.2016.09.009.

- 16. Oliveira LR, Oliveira-Costa JP, Araujo IM, Soave DF, Zanetti JS, Soares FA, et al. Cancer stem cell immunophenotypes in oral squamous cell carcinoma. J Oral Pathol Med. 2011;40(2):135-42. https://doi.org/10.1111/j.1600-0714.2010.00967.x.
- 17. Mohanta S, Siddappa G, Valiyaveedan SG, Ramanjanappa RD, Das D, Pandian R, et al. Cancer stem cell markers in patterning differentiation and in prognosis of oral squamous cell carcinoma. Tumour Biol. 2017;39(6):1010428317703656. https://doi.org/10.1177/1010428317703656.
- Lee HJ, Kang YH, Lee JS, Byun JH, Kim UK, Jang SJ, et al. Positive expression of NANOG, mutant p53, and *CD44* is directly associated with clinicopathological features and poor prognosis of oral squamous cell carcinoma. BMC Oral Health. 2015;15:153. https://doi.org/10.1186/s12903-015-0188-0.
- Todoroki K, Ogasawara S, Akiba J, Nakayama M, Naito Y, Seki N, et al. *CD44v3+/CD24-* cells possess cancer stem cell-like properties in human oral squamous cell carcinoma. Int J Oncol. 2016;48(1):99-109. https://doi.org/10.3892/ ijo.2015.3261.
- Huang CF, Xu XR, Wu TF, Sun ZJ, Zhang WF. Correlation of *ALDH1*, *CD44*, OCT4 and SOX2 in tongue squamous cell carcinoma and their association with disease progression and prognosis. J Oral Pathol Med. 2014;43(7):492-8. https://doi. org/10.1111/jop.12159.
- Jayasooriya P, Fernando C, Suraweera A, Dissanayake U. Stem cell markers as a resource to predict prognosis of betel quid induced oral squamous cell carcinoma: an immunohistochemical investigation. Stomatological Dis Sci. 2017;1:29-34. https://doi.org/10.20517/2573-0002.2016.11.
- Rao RS, K LR, Augustine D, Patil SG. Prognostic significance of ALDH1, Bmi1, and OCT4 expression in oral epithelial dysplasia and oral squamous cell carcinoma. Cancer Control. 2020;27(1):1073274820904959. https://doi. org/10.1177/1073274820904959.
- 23. Tamatani T, Takamaru N, Ohe G, Akita K, Nakagawa T, Miyamoto Y. Expression of *CD44*, *CD44* v9, *ABCG2*, *CD24*, Bmi-1 and *ALDH1* in stage I and II oral squamous cell carcinoma and their association with clinicopathological factors. Oncol Lett. 2018;16(1):1133-40. https://doi. org/10.38 92/ ol.2018.8703.
- 24. Wu TF, Li YC, Ma SR, Liu B, Zhang WF, Sun ZJ. Expression and associations of *TRAF1*, *BMI-1*, *ALDH1*, and *Lin28B* in oral squamous cell carcinoma. Tumour Biol. 2017; 39(4): 10104 283 17695930. https://doi. org/10.1177/1010428317695930.
- 25. de Freitas Filho SAJ, Coutinho-Camillo C, Klug Oliveira K, Bettim BB, Lopes Pinto CA, Kowalski LP, et al. Prognostic implications of *ALDH1* and *Notch1* in different subtypes of oral cancer. J Oncol. 2021;2021:6663720. https://doi. org/10.1155/2021/6663720.
- 26. Fugle CW, Zhang Y, Hong F, Sun S, Westwater C, Rachidi S, et al. *CD24* blunts oral squamous cancer development and dampens the functional expansion of myeloid-derived suppressor cells. Oncoimmunology. 2016;5(10). https://doi.org/10.1080/2162402X.2016.1226719.
- 27. Ghuwalewala S, Ghatak D, Das P, Dey S, Sarkar S, Alam N, et al. *CD44*(high)*CD24*(low) molecular signature determines the cancer stem cell and EMT phenotype in oral squamous cell carcinoma. Stem Cell Res. 2016;16(2):405-17. https:// doi.org/10.1016/j.scr.2016.02.028.
- 28. Mishra S, Tiwari V, Arora A, Gupta S, Anand N, Husain N.

Increased expression of Oct4, Nanog and *CD24* predicts poor response to chemo-radiotherapy and unfavorable prognosis in locally advanced oral squamous cell carcinoma. Asian Pac J Cancer Prev. 2020;21(9):2539-47. https://doi. org/10.31557/APJCP.2020.21.9.2539.

- Wang S, Fan H, Xu J, Zhao E. Prognostic implication of NOTCH1 in early stage oral squamous cell cancer with occult metastases. Clin Oral Investig. 2018;22(9):1131-8. https://doi.org/10.1007/s00784-017-2301-7.
- 30. Chiou SH, Yu CC, Huang CY, Lin SC, Liu CJ, Tsai TH, et al. Positive correlations of Oct-4 and Nanog in oral cancer stem-like cells and high-grade oral squamous cell carcinoma. Clin Cancer Res. 2008;14(13):4085-95. https:// doi.org/10.1158/1078-0432.CCR-07-4404.
- Yu Z, Pestell TG, Lisanti MP, Pestell RG. Cancer stem cells. Int J Biochem Cell Biol. 2012;44(12):2144-51. https://doi. org/10.1016/j.biocel.2012.08.022.
- 32. Gokulan RC, Devaraj H. Stem cell markers CXCR-4 and CD133 predict aggressive phenotype and their double positivity indicates poor prognosis of oral squamous cell carcinoma. Cancers (Basel). 2021;13(23):5895. https://doi. org/10.3390/cancers13235895.
- 33. Shipitsin M, Polyak K. The cancer stem cell hypothesis: in search of definitions, markers, and relevance. Lab Invest. 2008;88(5):459-63. https://doi.org/10.1038/ labinvest.2008.14.
- 34. Le H, Zeng F, Xu L, Liu X, Huang Y. The role of *CD133* expression in the carcinogenesis and prog- nosis of patients with lung cancer. Mol Med Rep. 2013;8(5):1511-8. https:// doi.org/10.3892/mmr.20 13.1667.
- Horst D, Kriegl L, Engel J, Kirchner T, Jung A. *CD133* expression is an independent prognostic m- arker for low survival in colorectal cancer. Br J Cancer. 2008;99(8):1285-9. https://doi.org/10.1038/sj.bjc.6604664.
- 36. Zhang Q, Shi S, Yen Y, Brown J, Ta JQ, Le AD. A subpopulation of *CD133*(+) cancer stem-like cells characterized in human oral squamous cell carcinoma confer resistance to chemotherapy. Cancer Lett. 2010;289(2):151-60. https://doi.org/10.1016/j.canlet.2009.08.010.
- 37. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. Cell. 2008;133(4):704-15. https://doi.org/10.1016/j.cell.2008.03.027.
- [38. Naor D, Wallach-Dayan SB, Zahalka MA, Vogt Sionov R. Involvement of *CD44*, a molecule with a thousand faces, in cancer dissemination. Semin Cancer Biol. 2008;18(4):260-7. https://doi.org/10.1016/j.semcancer.2008.03.015.
- Zhao P, Damerow MS, Stern P, Liu AH, Sweet-Cordero A, Siziopikou K, et al. *CD44* promotes Kras-dependent lung adenocarcinoma. Oncogene. 2013;32(43):5186-90. https:// doi.org/10.1038/onc.2012.542.
- 40. Ohi Y, Umekita Y, Yoshioka T, Souda M, Rai Y, Sagara Y, et al. Aldehyde dehydrogenase 1 expression predicts poor prognosis in triple-negative breast cancer. Histopathology. 2011;59(4):776-80. https://doi.org/10.1111/j.1365-2559.2011.03884.x.
- Hessman CJ, Bubbers EJ, Billingsley KG, Herzig DO, Wong MH. Loss of expression of the cancer stem cell marker aldehyde dehydrogenase 1 correlates with advanced-stage colorectal cancer. Am J Surg. 2012;203(5):649-53. https:// doi.org/10.1016/j.amjsurg.2012.01.003.
- 42. Götz C, Bissinger O, Nobis C, Wolff KD, Drecoll E, Kolk A. *ALDH1* as a prognostic marker for lymph node metastasis in OSCC. Biomed Rep. 2018;9(4):284-90. https://doi. org/10.3892/br.2018.1131.
- 43. Oliveira LR, Ribeiro-Silva A. Prognostic significance of immunohistochemical biomarkers in oral squamous cell

carcinoma. Int J Oral Maxillofac Surg. 2011;40(3):298-307. https://doi.org/10.101 6/j.ij om.2010.12.003.

- 44. Jing X, Cui X, Liang H, Hao C, Yang Z, Li X,et al . CD24 is a potential biomarker for prognosis in h- uman breast carcinoma. Cell Physiol Biochem. 2018;48(1):111-9. https:// doi.org/10.1159/000491667.
- 45. Yang XR, Xu Y, Yu B, Zhou J, Li JC, Qiu SJ, et al. *CD24* is a novel predictor for poor prognosis of hepatocellular carcinoma after surgery. Clin Cancer Res. 2009;15(17):5518-27. https://doi.org/10.1158/1078-0432.CCR-09-0151.
- 46. Su YJ, Chang YW, Lin WH, Liang CL, Lee JL. An aberrant nuclear localization of E-cadherin is a potent inhibitor of Wnt/β-catenin-elicited promotion of the cancer stem cell phenotype. Oncogenesis. 2015;4. https://doi.org/10.1038/ oncsis.2015.17.
- Ghazi N, Saghravanian N, Shakeri M, Jamali M. Evaluation of *CD44* and TGF-B Expression in Oral Carcinogenesis. Dent J (Tehran). 2021;22(1):33-40. https://doi.org/10.30476/ dentjods.2020.84393.1079.
- 48. Curtarelli RB, Gonçalves JM, Pena Dos Santos LG, Savi MG, Nör JE, Mezzomo LAM, et al. Expression of cancer stem cell biomarkers in human head and neck carcinomas: a systematic review. Stem Cell Rev Rep. 2018;14(6):769-84. https://doi.org/10.1007/s12015-018-9839-4.
- 49. Michifuri Y, Hirohashi Y, Torigoe T, Miyazaki A, Kobayashi J, Sasaki T, et al. High expression of *ALDH1* and *SOX2* diffuse staining pattern of oral squamous cell carcinomas correlates to lymph node metastasis. Pathol Int. 2012;62(10):684-9. https://doi.org/10.1111/j.1440-1827.2012.02851.x.
- 50. Szafarowski T, Sierdziński J, Ludwig N, Głuszko A, Filipowska A, Szczepański MJ. Assessment of cancer stem cell marker expression in primary head and neck squamous cell carcinoma shows prognostic value for aldehyde dehydrogenase (*ALDH1A1*). Eur J Pharmacol. 2020;867:172837. https://doi.org/10.1016/j. ejphar.2019.172837.
- 51. Singh A, Srivastava AN, Akhtar S, Siddiqui MH, Singh P, Kumar V. Correlation of *CD133* and Oct-4 expression with clinicopathological and demographic parameters in oral squamous cell carci-noma patients. Natl J Maxillofac Surg. 2018;9(1):8-13. https://doi.org/10.4103/njms.NJMS 60 17.
- 52. Hu Y, Zhang Y, Gao J, Lian X, Wang Y. The clinicopathological and prognostic value of *CD44* expression in bladder cancer: a study based on meta-analysis and TCGA data. Bioengineered. 2020;11(1):572-81. https://doi.org/10.1080 /21655979.2020.1765500.
- 53. Liu Y, Wu T, Lu D, Zhen J, Zhang L. *CD44* overexpression related to lymph node metastasis and poor prognosis of pancreatic cancer. Int J Biol Markers. 2018;33(3):308-13. https://doi.org/10.1177/1724600817746951.
- 54. Reategui EP, Antúnez de Mayolo A, Das PM, Astor FC, Singal R, Hamilton KL, et al. Characterization of *CD44v3*containing isoforms in head and neck cancer. Cancer Biol Ther. 2006;5(9):1163-8. https://doi.org/10.4161/ cbt.5.9.3065.
- 55 Chen C, Zhao S, Karnad A, Freeman JW. The biology and role of *CD44* in cancer progression: therapeutic implications. J Hematol Oncol. 2018;11:64. https://doi.org/10.1186/ s13045-018-0605-5.



This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.