Recent Advances in Cancer Immunotherapy Aided by Regulatory Non-Coding RNA: A Review

Debabrat Baishya^{1*}, Arpita Barman¹, Bijuli Rabha¹, Kaushik Kumar Bharadwaj¹, Joyeeta Talukdar²

Abstract

Cancer immunotherapies have remodeled the way many different forms of cancer are treated by, leveraging the immune system to recognize and attack malignant cells. These therapies have demonstrated durable responses in cancers affecting the blood, bone marrow, and lymph nodes, solidifying their role as a cornerstone of modern oncology. However, despite their success, current immunotherapy tools and techniques require further refinement.Regulatory non-coding RNAs (ncRNAs), actively transcribed by the mammalian genome, play crucial roles in regulating a wide range of cellular processes. They are key players in epigenetic mechanisms that govern differentiation, development, and the progression of invasive carcinogenesis. Emerging evidence suggests that regulatory ncRNAs are also pivotal in cancer immunity, acting as drivers of immune responses in the development of metastatic tumors. These ncRNAs influence the tumor microenvironment (TME), modulate the immune system, and affect the efficacy of immune checkpoint inhibitors and T cell therapies by regulating various signaling pathways. This review article explores the diagnostic and therapeutic potential of ncRNAs in cancer immunotherapy, highlighting their role in modulating carcinogenesis and antitumor immunity. By understanding the influence of regulatory ncRNAs, we can pave the way for the development of prognostic biomarkers and more effective tumor immunotherapies.

Keywords: Regulatory non-coding RNAs- tumor microenvironment- epigenetic regulations

Asian Pac J Cancer Prev, 26 (6), 1915-1930

Introduction

Emerging evidence has revealed non-coding RNAs (ncRNAs) as central architects of gene regulation, revolutionizing our understanding of genome functionality. RNA plays a crucial function in gene expression and regulation. The central dogma of molecular biology reported in late 1950 posited that the messenger RNA is the pillar in this RNA model. Many discoveries such as housekeeping RNAs (rRNA, tRNA)supported this model [1, 2, 3]. However, the discovery of ncRNAs have dramatically challenged the functional understanding of genome. ncRNAs are the junk RNA of the transcriptome that lack the ability of protein coding potential which are found in non-coding regions of eukaryotic genomes [4]. Only 2% of the human genome represents protein coding genes, while the vast portion of the genome is made up of non-coding RNA, which accounts for 98% of the transcriptome [5]. ncRNAs exhibit diverse functional capacities, categorized into housekeeping and regulatory types. Housekeeping ncRNAs include small nucleolar RNAs (snoRNAs), which are involved in the chemical modification of ribosomal and small nuclear RNAs, small nuclear RNAs (snRNAs) that participate in pre-mRNA splicing, guide RNAs (gRNAs) directing site-specific RNA modifications, RNase PRNAs essential for processing precursor tRNAs, telomere-related RNAs crucial for maintaining telomere integrity, and signal recognition particle RNAs (SRP RNAs) which assist in ribosome targeting to the endoplasmic reticulum. Each of these plays a vital role in maintaining fundamental cellular functions and stability. In contrast, regulatory ncRNAs, such as long non-coding RNAs (lncRNAs), microRNAs (miRNAs), piwi-interacting RNAs (piRNAs), and circular RNAs (circRNAs), are involved in modulating gene expression and various cellular events. [6, 7]. Regulatory ncRNAs act as pivotal riboregulators, playing a crucial role in the post-transcriptional regulation of both coding and non-coding gene regions. These ncRNAs impact a broad spectrum of cellular processes through diverse gene regulatory mechanisms, which can influence disease development. Recent studies indicate that ncRNAs also mediate gene silencing, a key form of epigenetic modification, and this mechanism is associated with several diseases, including cancer. The involvement of regulatory ncRNAs in these complex

¹Department of Bioengineering and Technology, Gauhati University, Gwahati-781014, Assam, India. ²Department of Biochemistry, All India Institute of Medica Sciences, New Delhi, India. *For Correspondence: drdbaishya@gmail.com

regulatory networks underscores their significance in both disease pathology and potential therapeutic applications [8 -11]. The integration of ncRNAs into therapeutic strategies represents a promising frontier in medicine. Advances in RNA-based therapies, including antisense oligonucleotides and RNA interference, pull the specific interactions of ncRNAs to target disease-associated genes. Additionally, the potential for ncRNAs to serve as biomarkers for early diagnosis and prognosis of diseases highlights their utility in clinical settings.Continued research into the functional diversity and regulatory mechanisms of ncRNAs is essential for understanding their roles in cellular homeostasis and disease. As our knowledge of ncRNA biology expands, it will lay the groundwork for innovative diagnostic and therapeutic approaches, offering fresh opportunities for personalized medicine and targeted treatments.

Cancer is a multifaceted genetic disorder characterized by a series of progressive mutations in cellular DNA, which ultimately lead to the malignant transformation of normal cells. The role of the immune system in cancer prevention has garnered substantial empirical support from contemporary research. Studies consistently show that individuals with compromised immune function exhibit a markedly increased susceptibility to cancer compared to those with robust immune systems [12-14]. Understanding the genetic and molecular mechanisms underlying cancer has led to the exploration of novel therapeutic targets. Recent research has illuminated how ncRNAs are deeply embedded in regulating various aspects of cancer biology. Earlier studies confirmed the role of lncRNAs in the regulation of cancer development and progression, including growth, metastasis, and recurrence [15, 16]. A major challenge in treating metastasis is the biological heterogeneity of cancer cells, which often leads to diagnosis at more advanced stages. In response, researchers are investigating innovative approaches to address this issue. One promising strategy is immunotherapy, which aims to activate the immune system more effectively to combat cancer. As weak immune system functionality within the cancer microenvironment is now recognized as a key hallmark of cancer, enhancing immune responses through such therapies could significantly improve treatment outcomes. [17]. Immunotherapy has demonstrated remarkable and durable efficacy, establishing itself as a crucial fourth pillar in cancer treatment. Emerging evidence highlights that regulatory non-coding RNAs play significant roles in various stages of tumor immunity [18, 19]. Gaining a deeper understanding of how ncRNAs regulate cancer immunity could unveil novel therapeutic targets and strategies. Therefore, this review aims to elucidate the functions of regulatory non-coding RNAs, offering new insights into cancer diagnosis and the advancement of immunotherapeutic approaches.

Unravelling the biogenesis of regulatory non-coding RNA

Regulatory non-coding RNAs (ncRNAs) represent a diverse class of RNA molecules that, despite their lack of long open reading frames, play crucial roles in the regulation of gene expression. These ncRNAs can range

from a mere handful of nucleotides to several thousand, reflecting a remarkable variability in their structure and function. The biogenesis of these regulatory ncRNAs, primarily controlled by RNA polymerase II, often involves complex processes of polyadenylation and splicing [5]. The biogenesis of small non-coding RNAs (sncRNAs) has complex linkage with RNA interference (RNAi). SncRNAs form complexes with Argonaute proteins, leading to mature sncRNAs that bind to complementary mRNA targets, silencing them effectively. Among sncRNAs, microRNAs (miRNAs) are processed in both the nucleus and cytoplasm. Primary miRNAs (primiRNAs), characterized by their hairpin structure, are cleaved by the endonuclease DROSHA in the nucleus to produce precursor miRNAs (pre-miRNAs). These are then exported to the cytoplasm, where they are further processed by DICER and incorporated into RNA-induced silencing complexes (RISCs) with Argonaute proteins, forming mature miRNAs that direct the silencing of target mRNAs. Long non-coding RNAs (lncRNAs), typically over 200 nucleotides in length, encompass various types such as long intergenic non-coding RNAs (lincRNAs), enhancer ncRNAs, and transcribed ultraconserved regions (T-UCRs). lncRNAs participate in numerous cellular functions, including chromatin remodeling, transcriptional regulation, and post-transcriptional processing, demonstrating their versatile roles in gene regulation [10, 20-24]. Long non-coding RNAs (lncRNAs) are primarily processed within the nucleus and exhibit features akin to protein-coding genes. These features encompass conserved promoter regions, distinct chromatin structures at their promoters, and regulation mediated by transcription factors and morphogens. Furthermore, IncRNAs demonstrate diverse tissue-specific expression patterns and undergo complex splicing events, including the generation of alternative splice variants [25, 26]. Circular RNAs (circRNAs), a distinct subset of long non-coding RNAs (lncRNAs), are characterized by their single-stranded, closed-loop structure. These circRNAs are derived from previously coding transcripts, and their biogenesis is precisely controlled by specific cis-acting elements and trans-acting factors. circRNAs are primarily generated through head-to-tail back-splicing, where the 3' end donor site is joined to an upstream 5' end acceptor site, or through intron hybridization. Additionally, some circRNAs originate from excised lariat introns [27 -31]. Figure 1 illustrates the biogenesis pathways of various regulatory ncRNAs, including lncRNAs, miRNAs, and circRNAs, and schematically depicts their regulatory mechanisms in tumor immunity. This visual representation underscores the complexity and significance of ncRNA biogenesis and their roles in cellular regulation and disease contexts.

Regulatory non-coding RNA in cancer immunity: Mechanism and functions

Regulatory ncRNAs govern a remarkable variety of biological functions including transcriptional interference, telomere maintenance, epigenetic changes, X chromosome inactivation, genome imprinting, post-transcriptional and translational control, structural organization, cell



Figure 1. Biogenesis of Regulatory ncRNAs(lncRNA,miRNA,circRNA) and Schematic Depicts of Their Regulatory Mechanism in Tumor Immunity. a) circRNAs biogenesis occur in nucleus; it downregulatetumour inflammation inhibition (\downarrow IL6 and \downarrow TNF- α)leading to inhibition of cancer growth also upregulate c-mycand EMT signaling to cause cancer. b)lncRNA biogenesis also processed in nucleus; it upregulate different sigalling pathways by interact with them resulting evolution of carcinogenesis and it may regulate the transcription of various immune related genes also involve in their development and differentiation leading to inhibition of tumor evolution. c)miRNAs biogenesis processed in both nucleus and cytoplasm; it also interact with signaling pathways and regulate the development of immune componenets resulting tumor evolution and tumor growth inhibition respectively.

differentiation and development [32]. ncRNAs are integral to a range of biological processes, including transcriptional regulation, telomere maintenance, and epigenetic modifications. Within the nucleus, lncRNAs and miRNAs modulate gene expression by influencing transcriptional machinery and chromatin remodeling. In the cytoplasm, these ncRNAs regulate mRNA processing, protein modifications, and cell signaling pathways [33]. In the context of cancer immunity, regulatory ncRNAs are of particular interest due to their significant roles in modulating immune responses. They can influence the expression of genes associated with immune cell function, tumor progression, and the response to immune checkpoint inhibitors. By affecting these pathways, ncRNAs can either promote or inhibit the development and progression of cancer, making them valuable targets for therapeutic strategies aimed at enhancing cancer immunity [34]. Their ability to modulate these processes underscores their potential as biomarkers and therapeutic targets for enhancing cancer treatment. The immune system is actively engaged in cancer progression and dissemination, which involves several critical stages: the release of tumor antigens, their presentation to immune cells, activation of immune responses, migration and

infiltration of immune cells into tumor tissues, and the elimination of cancer cells. This continuum of events, referred to as the cancer-immunity cycle, was extensively described by Mellman and Chen in 2013 [35]. lncRNAs and miRNAs are critical in regulating the self-renewal and differentiation of hematopoietic stem cells (HSCs). They modulate the process by which HSCs continuously renew themselves and differentiate into various specialized immune cells. lncRNAs are notably present in diverse immune cell types including monocytes, macrophages, dendritic cells, neutrophils, T cells, and B cells. They are involved in guiding their development and differentiation [36 -38]. Some lncRNA like NeST induce synthesis of IFN- γ in CD8 + T cells [39], lincRNA-Cox2, which regulate the transcription of immune genes [40], and GATA3-AS1 that participate in differentiation of Th2 cells [16]. Recent studies have highlighted that various lncRNAs play a critical role in regulating dendritic cell (DC) functions, which are essential for balancing pathogen elimination and immune tolerance. Key lncRNAs involved in this regulation include lnc-DC, lincRNA-Cox2, lincRNA-EPS, and AS-IL-1a. Among these, Inc-DC (long non-coding RNA in dendritic cells) exhibits a dual function by enhancing both antigen presentation and the

differentiation of CD4+ T cells [41, 42]. lncRNAs are involved in regulating the maturation of antigenpresenting cells (APCs) and the expression of proinflammatory cytokines. IncRNATCONS 00019715 has been reported to facilitate macrophage polarization towards the M(IFN- γ + LPS) phenotype, which is associated with increased inflammatory activity. In contrast, lncRNA Cox-2 can impede tumor growth by inhibiting the polarization of M2 macrophages, thus affecting the immune response within the tumor microenvironment [43, 44]. Long non-coding RNAs (lncRNAs) have been identified as potential tumor suppressors, with some shown to enhance MHC I expression and promote antigen processing in cancer cells. Elevated levels of miRNA (miR-135) have been observed in colon tumors, correlating with reduced antigenpresenting cell (APC) levels. Within the tumor microenvironment (TME), lncRNAs influence the expression of anti-apoptotic factors like tumor necrosis factor-alpha (*TNF*- α) and interact with the NF κ B pathway, often through Toll-like receptor (TLR)-mediated mechanisms. TLRs, in turn, stimulate the expression of various lncRNAs, including lincRNA-Cox2 [45 - 47]. Tumor cells develop resistance to immune responses by skewing the balance between pro-apoptotic and antiapoptotic signals, resulting in tumor immune escape (TIE). Regulatory non-coding RNAs (ncRNAs) are crucial in this process, as they modulate the expression of pro- and anti-apoptotic factors. For example, in breast cancer, miR-195, miR-24-2, and miR-365 have been found to reduce Bcl-2 levels, thus augmenting tumor cell death. In contrast, miR-125b and miR-106a contribute to high Bcl-2 expression, inhibiting apoptosis in leukemia, prostate cancer, lymphomas, and breast cancer cells, which enhances their proliferation and invasion. Moreover, the hyperactivation of the Wnt/ β -catenin signaling pathway is often linked to cancer development and progression. This pathway is regulated by various lncRNAs and miRNAs. miR-93 and miR-145 target β -catenin to modulate Wnt/ β -catenin signaling, while Smad7 acts to downregulate this pathway. These regulatory mechanisms demonstrate complex interactions between ncRNAs and signaling pathways in cancer biology [48 - 54]. Several regulatory non-coding RNAs (ncRNAs) have been identified as activators of the p53 pathway, with their roles examined in both p53-proficient and p53-deficient cells. Comparative analyses of gene expression profiles and *p53*-binding patterns under genotoxic or oncogenic stress have revealed multiple direct lncRNA targets of p53. These findings enhance our understanding of how ncRNAs interact with p53, shedding light on their potential impact on cellular responses to stress and their contributions to tumorigenesis [55, 56]. Again, the epithelial-mesenchymal transition (EMT) signaling pathway plays a significant role in the progression of epithelial tumors. Key transcription factors involved in this pathway drive molecular changes by reducing the expression of epithelial cadherin (*E-cadherin*) while increasing the levels of neural cadherin (N-cadherin) and vimentin, which facilitates carcinogenesis. These transcription factors not only contribute to tumor

development but also promote immunosuppression and aid in achieving tumor immune escape (TIE). Recent studies have shown that miRNAs such as miR-21, miR-137, miR-34a, and miR-106a/b regulate EMT by targeting specific transcription factors like ZEB1, Snail, and Twist1. For instance, miR-21 has been reported to inhibit EMT in colorectal cancer by suppressing both Snail and E-cadherin expression. Similarly, miR-137 and miR-34a impede EMT in ovarian cancer by downregulating Snail expression. These insights signal the crucial role of miRNAs in modulating EMT and their potential impact on tumor progression and immune evasion [57 - 63]. However, the key pathways underlying EMT and tumorigenesis remain incompletely understood. Recent studies have identified lncRNAs like LINC01186 and lncRNA-HIT as key regulators in the TGF- β signaling pathway, significantly impacting cell migration and invasion in lung and breast cancer cells, respectively. However, the depth of their mechanistic roles remains underexplored [64, 65]. Although promising, these findings warrant deeperanalysis on how these lncRNAs interact with other pathways and their potential relevance across multiple cancer types. The current evidence supporting these lncRNAs is valuable. But however, further validation is required to address potential limitations and inconsistencies. Moreover, translating these insights into clinical therapies poses challenges such as specificity and delivery. Future research should focus on overcoming these obstacles and expanding the investigation to other lncRNAs and cancers to fully realize their therapeutic potential. Certain lncRNAs have been identified as interactors with the polycomb repressive complex 2 (PRC2), often through the recruitment of enhancer of zeste homolog 2 (EZH2) or other PRC2 components, suggesting a mechanism by which lncRNAs regulate EMT [66, 67]. Although these findings shed light on the potential regulatory roles of lncRNAs in EMT, a comprehensive evaluation is necessary to fully understand their functional implications. The mechanism through which lncRNAs influence EMT via PRC2 could involve several plausible pathways: lncRNAs may guide PRC2 to specific genomic loci to modulate histone modifications, thereby repressing genes that maintain epithelial characteristics or activating those that promote mesenchymal traits. Alternatively, lncRNAs might alter the recruitment or activity of PRC2, affecting its ability to regulate target genes. Despite these possibilities, the evidence remains preliminary, and further research is needed to elucidate the exact molecular mechanisms and determine the broader applicability of these interactions across different cancer types. Addressing these questions will be crucial for advancing targeted therapies that leverage lncRNA-PRC2 interactions to modulate EMT and cancer progression. Recent research has highlighted the role of circular RNAs (circRNAs) in modulating immune responses and tumor progression. For instance, circAmotl1 has been shown to enhance STAT3 expression via the STAT3 signaling pathway and promote nuclear c-myc expression in tumor cells, potentially driving carcinogenesis [36]. However, a deeper understanding of how circAmotl1 influences c-myc and its broader impact

on tumor dynamics is needed. Similarly, hsa_ circRNA_002178 affects breast cancer development by downregulating miR-328-3p and upregulating COL1A1, and its silencing can reduce tumor inflammation and growth by lowering *IL-6* and *TNF-a* levels [68, 69]. To advance the field, future research should focus on elucidating the precise molecular mechanisms by which circRNAs like circAmot11 and hsa_circRNA_002178 modulate cancer progression and immune responses. Additionally, studies should explore the consistency of these findings across various cancer types and investigate the potential of targeting circRNAs for therapeutic applications. Addressing these areas will be crucial for utilizing circRNAs in cancer treatment and improving our understanding of their role in tumor biology.

Application of regulatory non-coding RNA in cancer immunotherapeutics

RNA-based therapeutics have transitioned from conceptual frameworks to tangible treatments, with noncoding RNAs (ncRNAs) emerging as a notable class of therapeutic agents. Unlike protein-coding RNAs, ncRNAs exert their effects by modulating gene expression without encoding proteins. Their therapeutic potential lies in their ability to regulate gene expression at the posttranscriptional level. ncRNAs achieve this by binding to specific messenger RNAs (mRNAs), thereby inhibiting their translation or promoting their degradation, which can disrupt tumorigenic pathways and influence cancer progression [70]. Despite promising preclinical and early clinical data, translating ncRNA-based therapies into routine clinical practice presents several challenges. To overcome these hurdles, future research should focus on improving the delivery mechanisms and stability of ncRNA therapeutics, understanding their interactions with target mRNAs, and evaluating their effectiveness across various cancer types. Addressing these challenges is crucial for advancing ncRNA-based therapies from experimental stages to widespread clinical use. miRNAs are short, approximately 20-25 nucleotides in length, and play a crucial role in the post-transcriptional regulation of gene expression. Since their initial discovery in Caenorhabditis elegans in 1993, miRNAs have emerged as significant non-coding RNAs (ncRNAs) involved in gene silencing [71]. They function by binding to complementary sequences in target messenger RNAs (mRNAs), resulting in either mRNA degradation or translational repression. This regulatory mechanism constitutes a major form of epigenetic alteration, influencing various cellular processes. Owing to their ability to modulate gene expression, miRNAs are implicated in numerous diseases, including human carcinogenesis [10]. Their significance in cancer is particularly noteworthy since they influence tumor proliferation, metastasis, and resistance to therapies, rendering them key targets for research study and prospective treatment strategies.

The regulatory roles of key ncRNAs are analyzed, highlighting their potential as standalone or combinatorial therapies. Their value as predictive biomarkers for anticancer immunotherapy is also examined. A comprehensive summary of the roles and applications of miRNA is presented in Supplementary Table 1, that details the functions and applications of lncRNA. Additionally, Table 2 provides an overview of the functions of siRNA and circRNA.

Inflammation and ncRNA in Cancer Immunotherapy

Inflammation is recognized as a significant indicator of malignant growth, with over 20% of tumors attributed to chronic inflammation. Immune regulatory molecules such as cytokines and chemokines play pivotal roles in managing immune responses and are integral to immunotherapy. The discovery of ncRNAs has added a new layer of control over immunity and inflammatory processes, spurring interest in inflammation-related research. In a study by Cao and colleagues, 82 lncRNAs were screened using Spearman correlation analysis with the immune score, identifying five key lncRNAs-AC005014.2, AC010503.4, AL450384.2, LINC00930, and SH3BP5-AS1 that were downregulated in bladder cancer tissues compared to normal bladder epithelial cells. Additionally, an in silico approach introduced the ncRI dataset, which compiles experimentally validated ncRNAs in inflammatory diseases from over 2,000 publications, totaling 11,166 curated entries, including 1,377 lncRNAs, 1,976 miRNAs, and 107 other ncRNAs from humans, rodents, and mice[131, 132]. The analysis of studies, such as those conducted by Cao and colleagues, reveald that while promising, ncRNA-based therapies face significant hurdles that must be overcome before they can be integrated into routine clinical practice. Challenges such as enhancing the delivery mechanisms and stability of ncRNA therapeutics, deciphering their complex interactions with target mRNAs, and assessing their efficacy across diverse cancer types are pivotal areas that require further exploration. By addressing these issues, the potential of ncRNA-based therapies to transform cancer immunotherapy can be fully realized, offering a new frontier in the fight against cancer.

Regulatory non coding RNA as immune checkpoint blocker

Immune-checkpoint proteins on T cells regulate immune responses, acting to prevent the immune system from attacking tumor cells when engaged by specific tumor-expressed proteins. Key proteins such as CTLA-4, TIM-3, LAG-3, BTLA, PD-1, and its ligand PD-L1 are central to this regulatory process, with elevated levels of PD-1 and *PD-L1* often indicating a poor prognosis due to their suppression of the anticancer immune response. Immune checkpoint blockade therapies have significantly advanced cancer treatment by reactivating T cell activity and enhancing their survival. However, traditional therapies that rely on monoclonal antibodies tend to target single molecules, which limits their ability to address entire physiological pathways or multiple targets simultaneously. Regulatory non-coding RNAs (ncRNAs) present a promising alternative, as they can target multiple genes concurrently, offering a more comprehensive approach to modulating the immune system. Incorporating ncRNAs into cancer immunotherapy could potentially overcome the limitations of traditional treatments, paving the way

Debabrat Baishya et al

Table 1. V	Various IncRN	A and Their	Role in C	Cancer Imm	unotherapy
------------	---------------	-------------	-----------	------------	------------

lncRNA	Cancer type	In vitro/ in vivo	Model	Function	references
ANRIL	Burkitt lymphoma	in vitro	Burkitt lymphoma cells	ANRIL silencing inhibited proliferation; promoted the apoptosis ↓cyclin D1, ↓E2F transcription factor 1 ↓Bcl-2	[104]
MIR22HG	colorectal cancer	in vitro	LoVo and HCT116	acts as tumor suppressor, silencing ofMIR22HG enhances tumor cell proliferation and survival overexpression of MIR22HG triggers T cell infiltration, enhancing Immunotherapy	[105]
AK036396	Lung , Colon cancer	in vitro	LLC, CT-26 cell	AK036396 knockdown downregulate PMN-MDSC immunosuppression Administration of siAK036396 delays tumor progression, stability of Fcnb	[106]
F630028O10Rik	Lung cancer	in vitro	LLC, MS1	↓VEGFA, ↓VEGFR2 inhibitendothelialcells clone formation, migration, invasion Injection of cholesterol-modified siRNA-F63into mice tumour tissues	[107]
		in vivo	C57BL/6 mice	inhibit tumour growth and progression by modulating tumour angiogenesis	
LINC00449	Leukemia	in vitro	AML cells	overexpression of LINC00449 inhibit the cell proliferation and invasion	[108]
lncMX1-215	HNSCC Ovarian cancer	in vitro	HN4, HN6 and HN30 Cal27, SCC4, SCC25 Detroit 562 ,293 T, SCC7	inhibits tumor proliferation capacity suppress cancer metastasis	[109]
		in vivo	BALB/c nude mice		
HOTTIP		in vitro	Neutrophils	enhance IL-6 expression,↑PD-L1	[110]
LINC-PIN	lung ,colon cancer	in vitro	HCT116, A549, &DLD1	Inhibits migration as well as invasion of cancer cells both in vitro/ in vivo	[111]
		in vivo	Mice		

for a more effective and multifaceted therapeutic strategy [133 - 135]. miR-142-5p has been shown to directly suppress PD-L1 expression in tumor cells by binding to its 3' untranslated region (3'UTR). This downregulation of PD-L1 not only enhances antitumor immunity but also significantly inhibits pancreatic cancer growth in vivo [92]. The findings suggest that miR-142-5p could serve as a promising therapeutic agent in cancer immunotherapy by targeting the PD-1/PD-L1 axis. This highlights the potential of microRNA-based interventions to modulate immune checkpoints, offering a novel strategy to enhance the effectiveness of existing treatments and overcome resistance in pancreatic and possibly other cancers. Another study revealed that miR-149-3p significantly impacts cancer immunotherapy in triple-negative breast cancer by downregulating mRNAs responsible for encoding immune checkpoint proteins such as PD-1, TIM-3, BTLA, and Foxp1. This downregulation effectively reverses CD8+T cell exhaustion, a key challenge in cancer treatment, and restores their cytokine secretion capabilities [88]. These findings suggest that targeting miR-149-3p

could be a promising approach to overcoming immune resistance in this aggressive form of breast cancer, offering a new direction for therapeutic. In addition, studies highlighted the crucial role of microRNAs in modulating immune checkpoint pathways with miRNA like miRNA-424-5p [73], miRNA-5119[76], miRNA-200a [77], miRNA-148a-3p [83], Let-7 family protein [86], microRNA-192-5p, microRNA-149-3p [88], miRNA-142-5p [92], miRNA-424(322) [93], miRNA-621[95] and miRNA-374b [98] demonstrating a significant association with the inhibition of immune checkpoint proteins such as *PD-L1*. This inhibition is particularly important because *PD-L1* is a key player in immune evasion by cancer cells, allowing tumors to suppress the immune response. By downregulating PD-L1 and other immune checkpoint molecules, these miRNAs can effectively disrupt this suppression, reactivating the immune system's ability to recognize and attack tumor cells. Regulatory noncoding RNAs, such as siRNA, lncRNA, and circRNA, have emerged as significant players in the modulation of PD-L1 activity. One such study revealed that the long

Table 2. Various siRNA and circRNA and Their Role in Cancer Immunotherapy

siRNA	Cancer type	In vitro/ in vivo	Model	Function	references
ODC-targeting siRNA	ESCC	in vitro	M2 macrophage	↓IL-10 and TGF-β, ↑IL-12p35	[112]
STAT3+PD-L1 siRNA	Skin melanoma Breast cancer	in vitro	B16-F10 & 4T1 cell lines	↓PD-L1 and STAT3 genes Suppresses	[113]
		in vivo	BALB/C and C57BL/6J mice	proliferation, migration, and angiogenesis suppresses cancer progression	
siRNA-loaded nanocarrier	-	in vitro	CD8+ T-cells from human blood	enhanced T-cell survival ↓PD-L1	[114]
siRNA-loaded PCAnp	Colon cancer	in vitro	T cells of colon cancer bearing mice	enhanced T cell responses blockade of A2AR & CTLA-4	[115]
siRNA-PD-L1 imatinib in a liposomal NP	Melanoma	in vitro	B16F10 cells	↑cancer cell apoptosis delayed tumor growth	[116]
NP-siCD47 /CCL25	Breast Cancer	in vitro	4T1 cells	tumor infiltration of CCR9+CD8+ T cells ↓CD47,PD-1/PD-L1 blockade	[117]
IL-6- & STAT3 SiRNA+TMC Np	Breast Cancer Colorectal Cancer Melanoma	in vitro	4T1, CT26, B16-F10	↓IL-6/STAT3 expression	[118]
PD-1 siRNA/ PD-L1 siRNA loaded PLGA np	Colon cancer	in vitro	CTLs MC38 tumor-bearing mice	↓mTOR signaling cosilencing of PD-1 and PD-L1	[119]
		in vivo		showed more significant tumor growth suppression and long-term tumor inhibition in colon cancer	
siRNA-PD-L1	-	in vitro	-	co-delivery of siRNA-PD-L1 and doxorubicin can block the inhibitory signal responding to T cells and enhancecytotoxicity of Dox against tumor cells	[120]
siLuc	Hepatocellular carcinoma	in vivo	Mice model	knockdown of Luc (siLuc)significant reductionin Luc activity for mice receiving NP-delivered siLuc treatments were confirmed by Luminescence imaging of the tumors confirming successfulsiRNA delivery and gene silencing	[121]
siPDL1	Pancreatic cancer	in vitro	PAN 02 cell line	Combination therapy consisting of gemcitabine and MN-siPDL1	[122]
		in vivo	and mice model	in a syngeneic murine pancreatic cancermodel resulted in a significant reduction in tumor growth and an increase in survival	
PD-1 siRNA	skin cancer	in vitro	B16 cell line and	PD-1 siRNA delivered by attenuated	[123]
		in vivo	C57BL/6 mice	Salmonella to melanoma-bearing mice in combination with pimozide showed optimal therapeutic effect on melanoma	
Si-ChLa NPs +Dendritic cell	breast cancer	in vivo	Mice	decreased tumor growth and metastasis ↓CD73 ↓Regulatory T ↓mycloid derived suppressor cells (MDSCs) ↓ and tumor associated macrophages ↑T cells, ↑inflammatory cytokines ↑interferon (IFN)-γ, ↑(IL)-17, ↓IL-10	[124]
siRNA	Melanoma	in vitro	B16F1	↓cell proliferation	[125]
Targeting Notch1		in vivo	mice	↓ Notch1, ↓hey1, ↓tumor growth ↑tumor-infiltrating CD8+ T lymphocytes and IFN-γ	
siIL-10RA+siTGF- βR	cervical cancer	in vitro	dendritic cell TC-1 P0 tumour model	initiated the strongest antigen-specific CD8(+) T cell immune responses	[126]
HPV Targetting	-	in vivo	C57BL/6 mice	reduced tumor growth	[127]
siRNA polyplex encapsulated in PLGA np	Melanoma	in vitro	DC	STAT3 silencing by nps restored DC maturation ↑CD86,↑TNF-α ↑T cell proliferation	[128]

Table 2. Continued

siRNA	Cancer type	In vitro/ in vivo	Model	Function	references
Hsacirc0009361	colorectal cancer	in vitro in vivo	CRC tissues and cells Mice	↑expression of APC2 and inhibited the activity of the Wnt/β-catenin pathway by competitively combining with miR-582 inhibited the growth & metastasis of CRC	[129]
circCACTIN	Gastric cancer	In vitro	BGC-823,MGC-803	↑CircCACTIN expression Knockdown of circCACTIN inhibited GC cells proliferation migration, invasion	[130]
		In vivo	BAL/BC Mice model	circCACTIN up-regulation promoted GC tumor growth and EMT, and circCACTIN down-regulation inhibited GC tumor growth and EMT SGC-7901 cell line	

non-coding RNA HOTTIP enhances immune evasion in ovarian cancer by upregulating *IL-6*, which in turn drives *PD-L1* expression in neutrophils. This insight suggests that HOTTIP could be a valuable therapeutic target in ovarian cancer, offering a novel approach to counteract the tumor's immune escape mechanisms and potentially improve treatment efficacy [110]. In yet another study, the circular RNA hsa_circ_0020397 was identified as a miR-138 sponge, leading to increased expression of TERT and *PD-L1*. This modulation impacts key cellular processes, including viability, apoptosis, and invasion of colorectal cancer cells [136]. The generalized mechanisms of non-coding regulatory RNAs, with a focus on miRNAs, are illustrated in Figure 2, while Figure 3 provides an overview of checkpoint-inhibiting receptors and their interactions. These figures collectively enhance our understanding of how regulatory RNAs function as immune checkpoint modulators, offering insights into their potential as therapeutic targets in cancer treatment.

Regulatory non coding RNA in T cell therapy

Immune cells called T cells have their own proteins



Figure 2. Generalized Mechanism of PD 1 and PDL1 Degradation by microRNA. Likewise, other types of regulatory ncRNA, such as circRNA, lncRNA, siRNA also follow this pathway for regulation of PD1-PDL1



Figure 3. Schematic Diagram Showing PD1- PD L1 Interaction and Their Inhibitors

called receptors that attach to foreign antigens and help trigger other parts of the immune system to destroy the foreign substance. There are many evidences that reveals the association between T cell exhaustion and dysfunction with cancer. During the process of exhaustion, T cells chronically exposed to tumor antigen and viral antigen gradually loses the ability to kill cancer cell [137]. CAR T-cell therapy is a type of adaptive cell transfer therapy in which a patient's T cells (a type of immune cell) are genetically engineered in the laboratory and reinfused to the patient's body where they specifically recognized and eliminate cancer cells [138]. A recent study on T cell therapy has been reported that treatment of CD8+ T cells with an miR-149-3p mimic reduced apoptosis, attenuated changes in mRNA markers of T-cell exhaustion and downregulated mRNAs encoding PD-1, TIM-3, BTLA and Foxp1. Also the treatment with miR-149-3p promoted the capacity of CD8+ T cells to kill targeted 4T1 mouse breast tumor cells [88]. In addition, there are more studies conducted on T cell therapy, which used siRNA mediated therapy in T cell therapy. Thiramanas [114] showed the effect of siRNA-loaded Nano carrier used in T cell therapy. They transfected CD8+ T cell with siRNA loaded Nano carrier which resulted in enhancement of survival of T cell. In another study, siRNA loaded PCA nanoparticle used in T cell therapy were used which also resulted in T cell enhancement and ability to kill tumor cell [115].

Regulatory non coding RNA in Dendritic cell (DC) therapy The advancement of dendritic cell (DC)-based immunotherapy represents a significant leap in cancer treatment, leveraging patient-derived DCs to mount potent anti-tumor immune responses. Recent innovations reveal the promise of combining DC vaccines with complementary therapies to enhance their therapeutic impact. For example, research has shown that siRNA polyplexes encapsulated in PLGA nanoparticles can effectively silence STAT3, leading to enhanced DC maturation and increased expression of activation markers such as CD86 and *TNF-a*, thereby boosting T cell proliferation. Additionally, studies using miR-5119 mimics in DCs have demonstrated improved T cell functionality and increased cytokine production in breast cancer models. Future progress in this field should focus on optimizing these therapies through strategic combinations with other immunotherapeutic agents, refining adjuvant selection, and perfecting administration routes and timing. Ensuring that DCs are thoroughly matured and fully capable of activating effector T cells will be essential for advancing DC-based immunotherapy and achieving more effective, personalized cancer treatments [139, 128, 76].

Non-coding RNAs (ncRNAs) have been shown to play a significant role in modulating tumor immunity by affecting various biological processes. These include influencing macrophage polarization, enhancing or inhibiting NK cell cytotoxicity, and regulating T cell responses [115, 98, 86, 78, 79, 101]. By targeting these key immune functions, ncRNAs can impact the overall efficacy of the immune response against tumors. Recent advancements in the study of non-coding

RNAs (ncRNAs) have illuminated their pivotal roles in modulating tumor immunity. Comprehensive lncRNA microarray analyses have identified 9,343 deregulated lncRNAs in proinflammatory macrophages, with lncRNA TCONS 00019715 emerging as a key regulator in driving macrophage polarization towards the M1 phenotype, thereby amplifying tumoricidal responses. Additionally, IncRNAs such as Flicr have been linked to prognosis in colorectal cancer, while lncRNA NKILA facilitates tumor immune evasion and lncRNA Tim3 impedes antitumor immunity. lncRNAs also significantly influence natural killer (NK) cell cytotoxicity and the PD-1/PD-L1 signaling pathway. For instance, microRNA-130a has been shown to enhance NK cell-mediated killing of non-small cell lung cancer (NSCLC) cells by targeting STAT3. However, STAT3 overexpression can negate the beneficial effects of miR-130a on NK cell activity [98, 140, 79]. These findings suggest that ncRNAs are not only integral to understanding tumor immune dynamics but also represent promising targets for novel therapeutic strategies aimed at improving cancer immunotherapy.

Regulatory non coding RNA in Nanomedicine

Nanotechnology, since its emergence in the 1990s, has rapidly advanced, distinguished by its nanometerscale dimensions, biodegradability, and the ability to covalently interact with a diverse array of small molecule drugs. These properties enable precise targeting of subnuclear sites, thus revolutionizing the potential for treating cancer and other diseases. Among the various strategies developed for the effective delivery of nanomedicines into the nucleus, RNA interference (RNAi) has become a particularly influential technique. Recent studies have demonstrated the efficacy of combining siRNA-loaded liposomal nanoparticles with therapeutic agents such as imatinib to target *PD-L1* in skin melanoma. This approach has been shown to induce significant apoptosis in cancer cells in vitro and inhibit tumor growth in vivo. [116].

These innovations underscore the potential of RNAimediated nanotechnology to transform cancer therapy through enhanced specificity and targeted delivery. In a significant study, researchers demonstrated that targeting STAT3 and IL-6 with siRNA-loaded chitosan nanoparticles markedly curtails cancer cell progression, illustrating the advanced capabilities of nanotechnology in disrupting critical tumor growth pathways. Similarly, another study employed PLGA nanoparticles to deliver siRNAs targeting both PD-1 and its ligand, achieving notable suppression of colon tumor growth [119]. miRNAs have emerged as a powerful contender in the battle against cancer, owing to their ability to simultaneously target multiple oncogenic pathways. In another significant study, the therapeutic potential of lipid nanoparticles (LNPs) encapsulating miR-634 were explored. When administered intravenously, these LNPs demonstrated a remarkable capacity to reduce tumor growth in a xenograft mouse model implanted with BxPC-3 pancreatic cancer cells [141]. This research underscores the tumor-suppressive power of miR-634 and highlights the promise of LNPs as an advanced delivery system, capable of enhancing the effectiveness of miRNAbased cancer therapies. We have tried to highlight diverse array of application of regulatory ncRNA in nanomedicine through a schematic representation as shown in Figure 4.

Major Challenges faced by ncRNA based therapeutics

The therapeutic targeting of noncoding RNAs (ncRNAs), including microRNAs (miRNAs) and long noncoding RNAs (lncRNAs), has emerged as an innovative approach for treating various cancers and other diseases. Despite the significant strides made towards the clinical application of RNA-based therapeutics—such as the development of antisense oligonucleotides and small interfering RNAs (siRNAs), with several gaining FDA approval trial outcomes have been inconsistent. Some studies report robust therapeutic effects, while others reveal limited efficacy or notable toxicity. As a



Figure 4. Applications of Regulatory ncRNA in Naomedicine

DOI:10.31557/APJCP.2025.26.6.1915 Role of ncRNA in Cancer Immunotherapy

result, alternative therapeutic entities like antimiRNAs are undergoing clinical testing, and lncRNA-based therapeutics are garnering increasing interest.

However, despite their promise, ncRNA-based therapies face considerable challenges. Chief among these are issues of specificity, tolerability, targeted delivery, avoidance of off-target effects, reduction of toxicity, and the ability to overcome mutations and drug resistance. One of the most significant obstacles is the efficient and precise delivery of oligonucleotide-based therapies particularly anti-miRNA, miRNA mimics, siRNA, and shRNA—into specific tissues and cells. The dual role of certain ncRNAs across different tissues complicates this further; for example, miR-26a promotes gliomagenesis but exhibits both tumor-suppressive and oncogenic roles in hepatocellular carcinoma, underscoring the need for context-specific therapeutic strategies.

There is also a critical need for more extensive preclinical and clinical studies to advance our understanding and application of ncRNA therapeutics. Current research predominantly relies on chemically synthesized oligonucleotides with extensive chemical modifications. However, the development of bioengineered or recombinant RNA molecules produced and folded in living cells represents a novel and potentially superior class of agents for both basic research and drug development, warranting rigorous evaluation.

A landmark achievement in RNA-based therapeutics was the FDA approval of Patisiran, which utilizes a lipid nanoparticle (NP) delivery system to accumulate in the liver and inhibit transthyretin production. Despite its success, Patisiran is not universally recognized as a therapeutic drug for broader applications, particularly due to its limited target specificity.

Off-target effects represent another significant concern, demanding meticulous consideration during the design of oligonucleotide-based drugs. Factors such as RNA structure, protein interactions, and mismatches at cleavage sites can lead to unintended consequences, further complicating therapeutic development.

Moreover, the functional impact of ncRNAs can vary dramatically across different cancer types. For instance, lncRNA Xist functions as a tumor suppressor in osteosarcoma, cervical cancer, and breast cancer, yet acts as an oncogene in pancreatic, gastric, thyroid, colon, hepatocellular, non-small-cell lung, bladder cancers, glioblastoma, and nasopharyngeal carcinoma. Such functional variability poses significant challenges in predicting therapeutic outcomes and necessitates a deeper understanding of ncRNA biology in diverse pathological contexts.

Another complicating factor is the inconsistency between short-term chemical inhibition of ncRNAs and long-term genetic deletion. Chronic loss-of-function of ncRNAs may trigger adaptive responses that compensate for their absence, raising concerns about the durability of therapeutic efficacy over time. This highlights the challenge of whether the effectiveness of ncRNA-targeting drugs may diminish as the cellular network adapts to reduced ncRNA activity.

Finally, the fundamental differences between targeting

IncRNAs and miRNAs must be carefully considered and addressed. miRNAs, by modulating a broad spectrum of targets, create a complex landscape where it is difficult to establish a direct correlation between target engagement and therapeutic efficacy. Additionally, the sequestration of oligonucleotides within various cellular compartments further complicates their therapeutic impact.

In conclusion, despite rapid advancements in cancer biology, the role of noncoding RNAs (ncRNAs) in cancer immunotherapy remains only partially understood. Abnormally expressed ncRNAs hold promise as diagnostic and prognostic biomarkers, as well as therapeutic targets for cancer management. However, there is an urgent need for more in-depth studies to unravel the novel mechanisms by which ncRNAs influence cancer initiation and progression, particularly through immunotherapeutic approaches. The challenges of translating ncRNA research into clinical practice must be addressed to embrace the paradigm shift from protein-coding to non-coding RNA therapeutics. As ncRNA research advances, the development of new methodologies based on Artificial Intelligence (AI) and Machine Learning (ML) will be crucial for identifying different types of ncRNAs, their interacting molecules, biological functions, and roles in diseases. In recent years, noncoding RNA-loaded nanoparticles have emerged as significant players in drug delivery, offering enhanced efficacy and optimized material size. The integration of nanotechnology and AI could provide innovative solutions to many formulation and development challenges. To efficiently prioritize and study ncRNAs, a multi-phased approach is required. This should include the implementation of long-read, highdepth sequencing, and the improvement of computational tools like AI and ML. AI-driven signaling amplification systems could be employed to visualize different regulatory ncRNAs. Looking ahead, AI is poised to play a pivotal role in transforming healthcare towards precision medicine. To achieve these goals, integrative efforts in immunotherapy must consider a multi-disciplinary approach, combining nanomaterials, computational systems biology, molecular biology, and clinical medicine. These approaches should be intelligently synergized to overcome the current challenges and fully realize the therapeutic potential of ncRNAs. In conclusion, while ncRNA-based therapeutics hold significant promise, their successful translation into clinical practice will require a concerted effort across multiple domains of science. Ongoing research and innovation will be vital in unlocking the full potential of ncRNAs, paving the way for their use in advanced therapeutic applications.

Author Contribution Statement

We would like to acknowledge the contributions of all authors in the preparation of this manuscript. The conceptualization, writing, and editing of the manuscript were primarily carried out by DB. AB contributed to writing the first draft and data acquisition, while BR was responsible for data acquisition, figure drawing, and writing the initial draft. KKB was involved in writing the first draft and editing. JT assisted with data acquisition and editing

Acknowledgements

None.

Conflict of Interest

The authors of this manuscript declares no conflict of interest.

References

- Chaffey N, Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. Molecular biology of the cell. 4th ed. Ann Bot. 2003;91(3):401. https://doi.org/10.1093/aob/mcg023.
- Jarroux J, Morillon A, Pinskaya M. History, discovery, and classification of lncRNAs. Adv Exp Med Biol. 2017;1008:1-46. https://doi.org/10.1007/978-981-10-5203-3_1.
- Cobb M. 60 years ago, Francis Crick changed the logic of biology. PLoS Biol. 2017;15(9):e2003243. https://doi. org/10.1371/journal.pbio.2003243.
- DeOcesano-Pereira C, Machado RAC, Chudzinski-Tavassi AM, Sogayar MC. Emerging roles and potential applications of non-coding RNAs in glioblastoma. Int J Mol Sci. 2020;21(7):2611. https://doi.org/10.3390/ijms21072611.
- Kung JT, Colognori D, Lee JT. Long noncoding RNAs: past, present, and future. Genetics. 2013;193(3):651-69. https:// doi.org/10.1534/genetics.112.146704.
- Dai X, Kaushik AC, Zhang J. The emerging role of major regulatory RNAs in cancer control. Front Oncol. 2019;9:920. https://doi.org/10.3389/fonc.2019.00920.
- Grammatikakis I, Panda AC, Abdelmohsen K, Gorospe M. Long noncoding RNAs and the molecular hallmarks of aging. Aging (Albany NY). 2014;6(12):992-1009. https:// doi.org/10.18632/aging.100710
- Fang Y, Fullwood MJ. Roles, functions, and mechanisms of long non-coding RNAs in cancer. Genomics Proteomics Bioinformatics. 2016;14(1):42-54. https://doi.org/10.1016/j. gpb.2015.09.006.
- Maston GA, Evans SK, Green MR. Transcriptional regulatory elements in the human genome. Annu Rev Genomics Hum Genet. 2006;7:29-59. https://doi.org/10.1146/annurev. genom.7.080505.115623.
- Spizzo R, Almeida MI, Colombatti A, Calin GA. Long noncoding RNAs and cancer: a new frontier of translational research? Oncogene. 2012;31(43):4577-87. https://doi. org/10.1038/onc.2011.621.
- Malecová B, Morris KV. Transcriptional gene silencing through epigenetic changes mediated by non-coding RNAs. Curr Opin Mol Ther. 2010;12(2):214-22.
- Krynitz B, Edgren G, Lindelöf B, Baecklund E, Brattström C, Wilczek H, Smedby KE. Risk of skin cancer and other malignancies in kidney, liver, heart and lung transplant recipients 1970 to 2008--a Swedish population-based study. Int J Cancer. 2013;132(6):1429-38. https://doi.org/10.1002/ ijc.27765.
- Guiguet M, Boué F, Cadranel J, Lang JM, Rosenthal E, Costagliola D. Effect of immunodeficiency, HIV viral load, and antiretroviral therapy on the risk of individual malignancies. Lancet Oncol. 2009;10(12):1152-9. https:// doi.org/10.1016/S1470-2045(09)70282-7.
- 14. Le Mire L, Hollowood K, Gray D, Bordea C, Wojnarowska F. Melanomas in renal transplant recipients. Br J Dermatol. 2006;154(3):472-7. https://doi.org/10.1111/j.1365-2133.2005.07094.x.
- 15. Liang WC, Fu WM, Wong CW, Wang Y, Wang WM, Hu GX,

et al. The lncRNA H19 promotes epithelial to mesenchymal transition by functioning as miRNA sponges in colorectal cancer. Oncotarget. 2015;6(26):22513-25. https://doi. org/10.18632/oncotarget.4154.

- 16. Zhang H, Nestor CE, Zhao S, Lentini A, Bohle B, Benson M, Wang H. Profiling of human CD4+ T-cell subsets identifies the TH2-specific noncoding RNA GATA3-AS1. J Allergy Clin Immunol. 2013;132(4):1005-8. https://doi.org/10.1016/j.jaci.2013.05.033.
- Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. Cell. 2010;140(6):883-99. https:// doi.org/10.1016/j.cell.2010.01.025.
- McCune JS. Rapid advances in immunotherapy to treat cancer. Clin Pharmacol Ther. 2018;103(4):540-544. https:// doi.org/10.1002/cpt.985.
- Denaro N, Merlano MC, Lo Nigro C. Long noncoding RNAs as regulators of cancer immunity. Mol Oncol. 2019;13(1):61-73. https://doi.org/10.1002/1878-0261.12413.
- Moazed D. Small RNAs in transcriptional gene silencing and genome defence. Nature. 2009;457(7228):413-20. https:// doi.org/10.1038/nature07756.
- O'Brien J, Hayder H, Zayed Y, Peng C. Overview of microRNA biogenesis, mechanisms of actions, and circulation. Front Endocrinol (Lausanne). 2018;9:402. https://doi.org/10.3389/fendo.2018.00402.
- Carthew RW, Sontheimer EJ. Origins and mechanisms of miRNAs and siRNAs. Cell. 2009;136(4):642-55. https://doi. org/10.1016/j.cell.2009.01.035.
- 23. Macfarlane LA, Murphy PR. MicroRNA: biogenesis, function and role in cancer. Curr Genomics. 2010;11(7):537-61. https://doi.org/10.2174/138920210793175895.
- 24. Kapranov P, Cheng J, Dike S, Nix DA, Duttagupta R, Willingham AT, et al. Rna maps reveal new rna classes and a possible function for pervasive transcription. Science. 2007;316(5830):1484-8. https://doi.org/10.1126/ science.1138341.
- 25. Wu Z, Liu X, Liu L, Deng H, Zhang J, Xu Q, et al. Regulation of lncRNA expression. Cell Mol Biol Lett. 2014;19(4):561-75. https://doi.org/10.2478/s11658-014-0212-6.
- 26. Rinn JL, Kertesz M, Wang JK, Squazzo SL, Xu X, Brugmann SA, et al. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. Cell. 2007;129(7):1311-23. https://doi.org/10.1016/j. cell.2007.05.022.
- Chen LL. The expanding regulatory mechanisms and cellular functions of circular RNAs. Nat Rev Mol Cell Biol. 2020;21(8):475-490. https://doi.org/10.1038/s41580-020-0243-y.
- Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, Kjems J. Natural RNA circles function as efficient microRNA sponges. Nature. 2013;495(7441):384-8. https://doi.org/10.1038/nature11993.
- 29. Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. RNA. 2013;19(2):141-57. https://doi.org/10.1261/rna.035667.112.
- Salzman J, Gawad C, Wang PL, Lacayo N, Brown PO. Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types. PLoS One. 2012;7(2):e30733. https://doi.org/10.1371/journal. pone.0030733.
- Zhang Y, Zhang XO, Chen T, Xiang JF, Yin QF, Xing YH, et al. Circular intronic long noncoding RNAs. Mol Cell. 2013;51(6):792-806. https://doi.org/10.1016/j. molcel.2013.08.017.
- 32. Fatica A, Bozzoni I. Long non-coding RNAs: new players in cell differentiation and development. Nat Rev Genet.

2014;15(1):7-21. https://doi.org/10.1038/nrg3606. Epub 2013 Dec 3.

- 33. Choudhari R, Sedano MJ, Harrison AL, Subramani R, Lin KY, Ramos EI, et al. Long noncoding RNAs in cancer: From discovery to therapeutic targets. Adv Clin Chem. 2020;95:105-147. https://doi.org/10.1016/ bs.acc.2019.08.003.
- Zhang L, Xu X, Su X. Noncoding RNAs in cancer immunity: functions, regulatory mechanisms, and clinical application. Mol Cancer. 2020;19:48. https://doi.org/10.1186/s12943-020-01154-0.
- Gonzalez H, Hagerling C, Werb Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. Genes Dev. 2018;32(19-20):1267-1284. https:// doi.org/10.1101/gad.314617.118.
- 36. Chen YG, Kim MV, Chen X, Batista PJ, Aoyama S, Wilusz JE, et al. Sensing Self and Foreign Circular RNAs by Intron Identity. Mol Cell. 2017;67(2):228-238.e5. https://doi.org/10.1016/j.molcel.2017.05.022.
- Raisch J, Darfeuille-Michaud A, Nguyen HT. Role of microRNAs in the immune system, inflammation and cancer. World J Gastroenterol. 2013;19(20):2985-96. https://doi. org/10.3748/wjg.v19.i20.2985.
- 38. Ahmad I, Valverde A, Ahmad F, Naqvi AR. Long Noncoding RNA in Myeloid and Lymphoid Cell Differentiation, Polarization and Function. Cells. 2020;9(2):269. https://doi. org/10.3390/cells9020269.
- 39. Gomez JA, Wapinski OL, Yang YW, Bureau JF, Gopinath S, Monack DM, et al. The NeST long ncRNA controls microbial susceptibility and epigenetic activation of the interferon-γ locus. Cell. 2013;152(4):743-54. https://doi. org/10.1016/j.cell.2013.01.015.
- 40. Carpenter S, Aiello D, Atianand MK, Ricci EP, Gandhi P, Hall LL, et al. A long noncoding RNA mediates both activation and repression of immune response genes. Science. 2013;341(6147):789-92. https://doi.org/10.1126/ science.1240925.
- 41. Atianand MK, Hu W, Satpathy AT, Shen Y, Ricci EP, Alvarez-Dominguez JR, et al. A Long Noncoding RNA lincRNA-EPS Acts as a Transcriptional Brake to Restrain Inflammation. Cell. 2016;165(7):1672-85. https://doi.org/10.1016/j. cell.2016.05.075.
- Kotzin JJ, Spencer SP, McCright SJ, Kumar DBU, Collet MA, Mowel WK, et al. The long non-coding rna morrbid regulates bim and short-lived myeloid cell lifespan. Nature. 2016;537(7619):239-43. https://doi.org/10.1038/ nature19346.
- 43. Huang Z, Luo Q, Yao F, Qing C, Ye J, Deng Y, Li J. Identification of differentially expressed long non-coding RNAs in polarized macrophages. Sci Rep. 2016;6:19705. https://doi.org/10.1038/srep19705.
- 44. Li X, Lei Y, Wu M, Li N. Regulation of macrophage activation and polarization by HCC-derived exosomal lncRNA TUC339. Int J Mol Sci. 2018;19(10):2958. https:// doi.org/10.3390/ijms19102958.
- 45. Yang Q, Xu B, Sun H, Wang X, Zhang J, Yu X, Ma X. A genome-wide association scan of biological processes involved in oral lichen planus and oral squamous cell carcinoma. Medicine (Baltimore). 2017;96(25):e7012. https://doi.org/10.1097/MD.000000000007012.
- 46. Akbari A, Ghahremani MH, Mobini GR, Abastabar M, Akhtari J, Bolhassani M, Heidari M. Down-regulation of miR-135b in colon adenocarcinoma induced by a TGF-β receptor I kinase inhibitor (SD-208). Iran J Basic Med Sci. 2015;18(9):856-61.
- 47. Johannessen M, Askarian F, Sangvik M, Sollid JE. Bacterial interference with canonical NFκB signalling.

Microbiology (Reading). 2013;159(Pt 10):2001-13. https:// doi.org/10.1099/mic.0.069369-0.

- Liu L, Wang Q, Qiu Z, Kang Y, Liu J, Ning S, et al. Noncoding RNAs: the shot callers in tumor immune escape. Signal Transduct Target Ther. 2020;5(1):102. https://doi. org/10.1038/s41392-020-0194-y.
- 49. Singh R, Saini N. Downregulation of BCL2 by miRNAs augments drug-induced apoptosis—a combined computational and experimental approach. J Cell Sci. 2012;125(Pt 6):1568-78. https://doi.org/10.1242/jcs.095976.
- Liu J, Tong CQ. Inhibitory effect of miR-125b downregulation on proliferation of leukemia cell K562. Zhongguo Shi Yan Xue Ye Xue Za Zhi. 2018;26(2):336-40. Chinese. https://doi.org/10.7534/j.issn.1009-2137.2018.02.005.
- 51. Huang X, Zhong R, He X, Deng Q, Peng X, Li J, Luo X. Investigations on the mechanism of progesterone in inhibiting endometrial cancer cell cycle and viability via regulation of long noncoding RNA NEAT1/microRNA-146b-5p mediated Wnt/β-catenin signaling. IUBMB Life. 2019;71(2):223-34. https://doi.org/10.1002/iub.1959.
- Wang G, Zhao Y, Zheng Y. MiR-122/Wnt/β-catenin regulatory circuitry sustains glioma progression. Tumour Biol. 2014;35(9):8565-72. https://doi.org/10.1007/s13277-014-2089-4.
- 53. Yamada N, Noguchi S, Mori T, Naoe T, Maruo K, Akao Y. Tumor-suppressive microRNA-145 targets catenin δ-1 to regulate Wnt/β-catenin signaling in human colon cancer cells. Cancer Lett. 2013;335(2):332-42. https://doi.org/10.1016/j.canlet.2013.02.060.
- 54. Tang Q, Zou Z, Zou C, Zhang Q, Huang R, Guan X, et al. MicroRNA-93 suppresses colorectal cancer development via Wnt/β-catenin pathway downregulation. Tumour Biol. 2015;36(3):1701-10. https://doi.org/10.1007/s13277-014-2771-6.
- 55. Olivero CE, Dimitrova N. Identification and characterization of functional long noncoding RNAs in cancer. FASEB J. 2020;34(12):15630-46. https://doi.org/10.1096/ fj.202001951R.
- Zhang A, Xu M, Mo YY. Role of the lncRNA-p53 regulatory network in cancer. J Mol Cell Biol. 2014;6(3):181-91. https:// doi.org/10.1093/jmcb/mju013.
- Micalizzi DS, Farabaugh SM, Ford HL. Epithelialmesenchymal transition in cancer: parallels between normal development and tumor progression. J Mammary Gland Biol Neoplasia. 2010;15(2):117-34. https://doi.org/10.1007/ s10911-010-9178-9.
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. Nat Rev Mol Cell Biol. 2014;15(3):178-196. https://doi.org/10.1038/nrm3758.
- Kudo-Saito C, Shirako H, Takeuchi T, Kawakami Y. Cancer metastasis is accelerated through immunosuppression during Snail-induced EMT of cancer cells. Cancer Cell. 2009;15(3):195-206. https://doi.org/10.1016/j. ccr.2009.01.023.
- Slaby O, Svoboda M, Michalek J, Vyzula R. MicroRNAs in colorectal cancer: translation of molecular biology into clinical application. Mol Cancer. 2009;8:102. https://doi. org/10.1186/1476-4598-8-102.
- Sánchez-Tilló E, Liu Y, de Barrios O, Siles L, Fanlo L, Cuatrecasas M, et al. EMT-activating transcription factors in cancer: beyond EMT and tumor invasiveness. Cell Mol Life Sci. 2012;69(20):3429-56. https://doi.org/10.1007/ s00018-012-1122-2.
- 62. Zhong G, Cheng X, Long H, He L, Qi W, Xiang T, et al. Dynamically expressed microRNA-15b modulates the activities of CD8+ T lymphocytes in mice with Lewis lung carcinoma. J Transl Med. 2013;11:71. https://doi.

org/10.1186/1479-5876-11-71.

- 63. Dong P, Xiong Y, Watari H, Hanley SJ, Konno Y, Ihira K, et al. MiR-137 and miR-34a directly target Snail and inhibit EMT, invasion and sphere-forming ability of ovarian cancer cells. J Exp Clin Cancer Res. 2016;35(1):132. https://doi. org/10.1186/s13046-016-0415-y.
- 64. Hao Y, Yang X, Zhang D, Luo J, Chen R. Long noncoding RNA LINC01186, regulated by TGF-β/SMAD3, inhibits migration and invasion through Epithelial-Mesenchymal-Transition in lung cancer. Gene. 2017;608:1-12. https://doi. org/10.1016/j.gene.2017.01.023.
- 65. Richards EJ, Zhang G, Li ZP, Permuth-Wey J, Challa S, Li Y, et al. Long non-coding RNAs (LncRNA) regulated by transforming growth factor (TGF) β: LncRNA-hit-mediated TGFβ-induced epithelial to mesenchymal transition in mammary epithelia. J Biol Chem. 2015;290(11):6857-67. https://doi.org/10.1074/jbc.M114.610915.
- 66. Battistelli C, Cicchini C, Santangelo L, Tramontano A, Grassi L, Gonzalez FJ, et al. The Snail repressor recruits EZH2 to specific genomic sites through the enrollment of the lncRNA HOTAIR in epithelial-to-mesenchymal transition. Oncogene. 2017;36(7):942-55. https://doi.org/10.1038/ onc.2016.260.
- 67. Yang T, He X, Chen A, Tan K, Du X. LncRNA HOTAIR contributes to the malignancy of hepatocellular carcinoma by enhancing epithelial-mesenchymal transition via sponging miR-23b-3p from ZEB1. Gene. 2018;670:114-122. https:// doi.org/10.1016/j.gene.2018.05.061.
- 68. Yang ZG, Awan FM, Du WW, Zeng Y, Lyu J, Wu D, et al. The Circular RNA Interacts with *STAT3*, Increasing Its Nuclear Translocation and Wound Repair by Modulating Dnmt3a and miR-17 Function. Mol Ther. 2017;25(9):2062-74. https:// doi.org/10.1016/j.ymthe.2017.05.022.
- Liu T, Ye P, Ye Y, Lu S, Han B. Circular RNA hsa_ circRNA_002178 silencing retards breast cancer progression via microRNA-328-3p-mediated inhibition of COL1A1. J Cell Mol Med. 2020;24(3):2189-201. https://doi. org/10.1111/jcmm.14875.
- Sullenger BA, Nair S. From the RNA world to the clinic. Science. 2016;352(6292):1417-20. https://doi.org/10.1126/ science.aad8709.
- 71. Yang N, Zhu S, Lv X, Qiao Y, Liu YJ, Chen J. MicroRNAs: Pleiotropic Regulators in the Tumor Microenvironment. Front Immunol. 2018;9:2491. https://doi.org/10.3389/ fimmu.2018.02491.
- 72. Zhou Y, Yamamoto Y, Takeshita F, Yamamoto T, Xiao Z, Ochiya T. Delivery of miR-424-5p via Extracellular Vesicles Promotes the Apoptosis of MDA-MB-231 TNBC Cells in the Tumor Microenvironment. Int J Mol Sci. 2021;22(2):844. https://doi.org/10.3390/ijms22020844.
- Barsoum FS, Awad AS, Hussein NH, Eissa RA, El Tayebi HM. MALAT-1: LncRNA ruling miR-182/PIG-C/ mesothelin triad in triple negative breast cancer. Pathol Res Pract. 2020;216(12):153274. https://doi.org/10.1016/j. prp.2020.153274.
- 74. Xi Q, Zhang J, Yang G, Zhang L, Chen Y, Wang C, et al. Restoration of miR-340 controls pancreatic cancer cell CD47 expression to promote macrophage phagocytosis and enhance antitumor immunity. J Immunother Cancer. 2020;8(1):e000253. https://doi.org/10.1136/jitc-2019-000253.
- 75. Fan X, Wang J, Qin T, Zhang Y, Liu W, Jiang K, Huang D. Exosome miR-27a-3p secreted from adipocytes targets ICOS to promote antitumor immunity in lung adenocarcinoma. Thorac Cancer. 2020;11(6):1453-64. https://doi.org/10.1111/1759-7714.13411.
- 76. Zhang M, Shi Y, Zhang Y, Wang Y, Alotaibi F, Qiu L, et al.

miRNA-5119 regulates immune checkpoints in dendritic cells to enhance breast cancer immunotherapy. Cancer Immunol Immunother. 2020;69(6):951-67. https://doi.org/10.1007/s00262-020-02507-w.

- 77. Liu Z, Wen J, Wu C, Hu C, Wang J, Bao Q, et al. MicroRNA-200a induces immunosuppression by promoting PTENmediated *PD-L1* upregulation in osteosarcoma. Aging (Albany NY). 2020;12(2):1213-36. https://doi.org/10.18632/ aging.102679.
- Wang J, Zhu M, Zhou X, Wang T, Xi Y, Jing Z, Xi W. MiR-140-3p inhibits natural killer cytotoxicity to human ovarian cancer via targeting MAPK1. J Biosci. 2020;45:66.
- 79. Zhou X, Liu S, Liu J, Zhang Z, Mao X, Zhou H. MicroRNA-130a enhances the killing ability of natural killer cells against non-small cell lung cancer cells by targeting signal transducers and activators of transcription 3. Biochem Biophys Res Commun. 2020;523(2):481-6. https://doi. org/10.1016/j.bbrc.2019.11.099.
- 80. Yu J, Zhao Y, Liu C, Hu B, Zhao M, Ma Y, Jiang J. Synergistic anti-tumor effect of paclitaxel and miR-34a combined with ultrasound microbubbles on cervical cancer in vivo and in vitro. Clin Transl Oncol. 2020;22(1):60-69. https://doi. org/10.1007/s12094-019-02131-w.
- 81. Ji Y, Fioravanti J, Zhu W, Wang H, Wu T, Hu J, et al. miR-155 harnesses Phf19 to potentiate cancer immunotherapy through epigenetic reprogramming of CD8+ T cell fate. Nat Commun. 2019;10(1):2157. https://doi.org/10.1038/ s41467-019-09882-8.
- Puccetti MV, Adams CM, Dan TD, Palagani A, Simone BA, DeAngelis T, et al. MicroRNA-21 is required for hematopoietic cell viability after radiation exposure. Int J Radiat Oncol Biol Phys. 2019;104(5):1165-74. https://doi. org/10.1016/j.ijrobp.2019.04.020.
- 83. Kao SH, Cheng WC, Wang YT, Wu HT, Yeh HY, Chen YJ, et al. Regulation of miRNA biogenesis and histone modification by K63-polyubiquitinated DDX17 controls cancer stem-like features. Cancer Res. 2019;79(10):2549-63. https://doi.org/10.1158/0008-5472.CAN-18-2376.
- 84. Xu Y, Ji K, Wu M, Hao B, Yao KT, Xu Y. A miRNA-HERC4 pathway promotes breast tumorigenesis by inactivating tumor suppressor LATS1. Protein Cell. 2019;10(8):595-605. https://doi.org/10.1007/s13238-019-0607-2.
- Fortunato O, Borzi C, Milione M, Centonze G, Conte D, Boeri M, et al. Circulating miR-320a promotes immunosuppressive macrophages M2 phenotype associated with lung cancer risk. Int J Cancer. 2019;144(11):2746-61. https://doi.org/10.1002/ijc.31988.
- 86. Yu D, Liu X, Han G, Liu Y, Zhao X, Wang D, et al. The let-7 family of microRNAs suppresses immune evasion in head and neck squamous cell carcinoma by promoting *PD-L1* degradation. Cell Commun Signal. 2019;17(1):173. https:// doi.org/10.1186/s12964-019-0490-8.
- Zou P, Zhu M, Lian C, Wang J, Chen Z, Zhang X, et al. miR-192-5p suppresses the progression of lung cancer bone metastasis by targeting TRIM44. Sci Rep. 2019;9(1):19619. https://doi.org/10.1038/s41598-019-56018-5.
- Zhang M, Gao D, Shi Y, Wang Y, Joshi R, Yu Q, et al. miR-149-3p reverses CD8+ T-cell exhaustion by reducing inhibitory receptors and promoting cytokine secretion in breast cancer cells. Open Biol. 2019;9(10):190061. https:// doi.org/10.1098/rsob.190061.
- Neviani P, Wise PM, Murtadha M, Liu CW, Wu CH, Jong AY, et al. Natural Killer-Derived Exosomal miR-186 Inhibits Neuroblastoma Growth and Immune Escape Mechanisms. Cancer Res. 2019;79(6):1151-64. https://doi. org/10.1158/0008-5472.CAN-18-0779.
- 90. Zhou Y, Zheng X, Chen LJ, Xu B, Jiang JT. microRNA-181b

suppresses the metastasis of lung cancer cells by targeting sex determining region Y-related high mobility group-box 6 (Sox6). Pathol Res Pract. 2019;215(2):335-342. https://doi. org/10.1016/j.prp.2018.12.009.

- 91. Yang Y, Huang G, Zhou Z, Fewell JG, Kleinerman ES. miR-20a Regulates FAS Expression in Osteosarcoma Cells by Modulating FAS Promoter Activity and Can be Therapeutically Targeted to Inhibit Lung Metastases. Mol Cancer Ther. 2018;17(1):130-139. https://doi. org/10.1158/1535-7163.MCT-17-0042.
- 92. Jia L, Xi Q, Wang H, Zhang Z, Liu H, Cheng Y, et al. miR-142-5p regulates tumor cell *PD-L1* expression and enhances anti-tumor immunity. Biochem Biophys Res Commun. 2017;488(2):425-431. https://doi.org/10.1016/j. bbrc.2017.05.074.
- 93. Xu S, Tao Z, Hai B, Liang H, Shi Y, Wang T, et al. miR-424(322) reverses chemoresistance via T-cell immune response activation by blocking the *PD-L1* immune checkpoint. Nat Commun. 2016;7:11406. https://doi. org/10.1038/ncomms11406.
- 94. Yang L, Cai Y, Zhang D, Sun J, Xu C, Zhao W, et al. miR-195/miR-497 Regulate CD274 Expression of Immune Regulatory Ligands in Triple-Negative Breast Cancer. J Breast Cancer. 2018;21(4):371-381. https://doi.org/10.4048/ jbc.2018.21.e60.
- 95. Shao Y, Song X, Jiang W, Chen Y, Ning Z, Gu W, Jiang J. MicroRNA-621 Acts as a Tumor Radiosensitizer by Directly Targeting SETDB1 in Hepatocellular Carcinoma. Mol Ther. 2019;27(2):355-364. https://doi.org/10.1016/j. ymthe.2018.11.005.
- 96. Yang X, Zhang L, Song X, He W, Zhang D, Lu Q, et al. MicroRNA-613 promotes colon cancer cell proliferation, invasion and migration by targeting ATOH1. Biochem Biophys Res Commun. 2018;504(4):827-833. https://doi. org/10.1016/j.bbrc.2018.09.054.
- 97. Shao Y, Zhang D, Li X, Yang J, Chen L, Ning Z, et al. MicroRNA-203 Increases Cell Radiosensitivity via Directly Targeting Bmi-1 in Hepatocellular Carcinoma. Mol Pharm. 2018;15(8):3205-3215. https://doi.org/10.1021/acs. molpharmaceut.8b00302.
- 98. Huang F, Wang B, Zeng J, Sang S, Lei J, Lu Y. MicroRNA-374b inhibits liver cancer progression via down-regulating programmed cell death-1 expression on cytokine-induced killer cells. Oncol Lett. 2018;15(4):4797-4804. https://doi. org/10.3892/ol.2018.7951.
- 99. He J, Ji Y, Li A, Zhang Q, Song W, Li Y, et al. MiR-122 directly inhibits human papillomavirus E6 gene and enhances interferon signaling through blocking suppressor of cytokine signaling 1 in SiHa cells. PLoS One. 2014;9(9):e108410. https://doi.org/10.1371/journal.pone.0108410.
- 100. van der Deen M, Taipaleenmäki H, Zhang Y, Teplyuk NM, Gupta A, Cinghu S, et al. MicroRNA-34c inversely couples the biological functions of the runt-related transcription factor RUNX2 and the tumor suppressor *p53* in osteosarcoma. J Biol Chem. 2013;288(29):21307-19. https://doi.org/10.1074/jbc.M112.445890.
- 101. Sun X, Zhang J, Hou Z, Han Q, Zhang C, Tian Z. miR-146a is directly regulated by STAT3 in human hepatocellular carcinoma cells and involved in anti-tumor immune suppression. Cell Cycle. 2015;14(2):243-52. https://doi.or g/10.4161/15384101.2014.977112.
- 102. Arts N, Cané S, Hennequart M, Lamy J, Bommer G, Van den Eynde B, De Plaen E. microRNA-155, induced by interleukin-1β, represses the expression of microphthalmiaassociated transcription factor (MITF-M) in melanoma cells. PLoS One. 2015;10(4):e0122517. https://doi.org/10.1371/ journal.pone.0122517.

- 103. Nishida N, Mimori K, Fabbri M, Yokobori T, Sudo T, Tanaka F, et al. MicroRNA-125a-5p is an independent prognostic factor in gastric cancer and inhibits the proliferation of human gastric cancer cells in combination with trastuzumab. Clin Cancer Res. 2011;17(9):2725-33. https://doi.org/10.1158/1078-0432.CCR-10-2132.
- 104. Mao S, Jin J, Li Z, Yang W. Knockdown of long non coding RNA ANRIL inhibits the proliferation and promotes the apoptosis of Burkitt lymphoma cells through the TGF β 1 signaling pathway. Mol Med Rep. 2021;23(2):146. https://doi.org/10.3892/mmr.2020.11785.
- 105. Xu J, Shao T, Song M, Xie Y, Zhou J, Yin J, et al. MIR22HG acts as a tumor suppressor via TGFβ/SMAD signaling and facilitates immunotherapy in colorectal cancer. Mol Cancer. 2020;19(1):51. https://doi.org/10.1186/s12943-020-01174-w.
- 106. Tian X, Zheng Y, Yin K, Ma J, Tian J, Zhang Y, et al. LncRNA AK036396 Inhibits Maturation and Accelerates Immunosuppression of Polymorphonuclear Myeloid-Derived Suppressor Cells by Enhancing the Stability of Ficolin B. Cancer Immunol Res. 2020;8(4):565-577. https:// doi.org/10.1158/2326-6066.CIR-19-0595.
- 107. Qin L, Zhong M, Adah D, Qin L, Chen X, Ma C, et al. A novel tumour suppressor lncRNA F630028O10Rik inhibits lung cancer angiogenesis by regulating miR-223-3p. J Cell Mol Med. 2020;24(6):3549-3559. https://doi.org/10.1111/ jcmm.15044.
- 108. Shi Y, Zhu Y, Zheng X, Zheng Z. LINC00449 regulates the proliferation and invasion of acute monocytic leukemia and predicts favorable prognosis. J Cell Physiol. 2020;235(10):6536-6547. https://doi.org/10.1002/jcp.29487.
- 109. Ma H, Chang H, Yang W, Lu Y, Hu J, Jin S. A novel IFNα-induced long noncoding RNA negatively regulates immunosuppression by interrupting H3K27 acetylation in head and neck squamous cell carcinoma. Mol Cancer. 2020;19(1):4. https://doi.org/10.1186/s12943-019-1123-y.
- 110. Shang A, Wang W, Gu C, Chen C, Zeng B, Yang Y, et al. Long non-coding RNA HOTTIP enhances *IL-6* expression to potentiate immune escape of ovarian cancer cells by upregulating the expression of *PD-L1* in neutrophils. J Exp Clin Cancer Res. 2019;38(1):411. https://doi.org/10.1186/ s13046-019-1394-6.
- 111. Marín-Béjar O, Mas AM, González J, Martinez D, Athie A, Morales X, et al. The human lncRNA LINC-PINT inhibits tumor cell invasion through a highly conserved sequence element. Genome Biol. 2017;18(1):202. https:// doi.org/10.1186/s13059-017-1331-y.
- 112. Mai S, Liu L, Jiang J, Ren P, Diao D, Wang H, Cai K. Oesophageal squamous cell carcinoma-associated IL-33 rewires macrophage polarization towards M2 via activating ornithine decarboxylase. Cell Prolif. 2021;54(2):e12960. https://doi.org/10.1111/cpr.12960.
- 113. Bastaki S, Aravindhan S, Ahmadpour Saheb N, Afsari Kashani M, Evgenievich Dorofeev A, Karoon Kiani F, et al. Codelivery of stat3 and pd-l1 sirna by hyaluronatetat trimethyl/thiolated chitosan nanoparticles suppresses cancer progression in tumor-bearing mice. Life Sci. 2021;266:118847. https://doi.org/10.1016/j.lfs.2020.118847.
- 114. Thiramanas R, Li M, Jiang S, Landfester K, Mailänder V. Cellular Uptake of siRNA-Loaded Nanocarriers to Knockdown *PD-L1*: Strategies to Improve T-cell Functions. Cells. 2020;9(9):2043. https://doi.org/10.3390/cells9092043.
- 115. Ghasemi-Chaleshtari M, Kiaie SH, Irandoust M, Karami H, Nabi Afjadi M, Ghani S, et al. Concomitant blockade of A2AR and CTLA-4 by siRNA-loaded polyethylene glycol-chitosan-alginate nanoparticles synergistically enhances antitumor T-cell responses. J Cell Physiol.

2020;235(12):10068-80. https://doi.org/10.1002/jcp.29822.

- 116. Li C, Han X. Melanoma cancer immunotherapy using PD-L1 siRNA and imatinib promotes cancer-immunity cycle. Pharm Res. 2020;37(6):109. https://doi.org/10.1007/ s11095-020-02838-4.
- 117. Chen H, Cong X, Wu C, Wu X, Wang J, Mao K, et al. Intratumoral delivery of CCL25 enhances immunotherapy against triple-negative breast cancer by recruiting CCR9+ T cells. Sci Adv. 2020;6(5):eaax4690. https://doi.org/10.1126/ sciadv.aax4690.
- 118. Masjedi A, Ahmadi A, Atyabi F, Farhadi S, Irandoust M, Khazaei-Poul Y, et al. Silencing of *IL-6* and *STAT3* by siRNA-loaded hyaluronate-N,N,N-trimethyl chitosan nanoparticles potently reduces cancer cell progression. Int J Biol Macromol. 2020;149:487-500. https://doi. org/10.1016/j.ijbiomac.2020.01.273.
- 119. Kwak SY, Lee S, Han HD, Chang S, Kim KP, Ahn HJ. PLGA nanoparticles co-delivering siRNAs against programmed cell death protein-1 and its ligand gene for suppression of colon tumor growth. Mol Pharm. 2019;16(12):4940-53. https:// doi.org/10.1021/acs.molpharmaceut.9b00826.
- 120. Wan WJ, Qu CX, Zhou YJ, Zhang L, Chen MT, Liu Y, et al. Doxorubicin and siRNA-PD-L1 co-delivery with T7-modified ROS-sensitive nanoparticles for tumor chemoimmunotherapy. Int J Pharm. 2019;566:731-744. https://doi.org/10.1016/j.ijpharm.2019.06.030.
- 121. Revia RA, Stephen ZR, Zhang M. Theranostic nanoparticles for RNA-based cancer treatment. Acc Chem Res. 2019;52(6):1496-1506. https://doi.org/10.1021/acs. accounts.9b00101.
- 122. Yoo B, Jordan VC, Sheedy P, Billig AM, Ross A, Pantazopoulos P, Medarova Z. RNAi-mediated *PD-L1* inhibition for pancreatic cancer immunotherapy. Sci Rep. 2019;9(1):4712. https://doi.org/10.1038/s41598-019-41251-9.
- 123. Zhao T, Wei T, Guo J, Wang Y, Shi X, Guo S, et al. PD-1-siRNA delivered by attenuated Salmonella enhances the anti-melanoma effect of pimozide. Cell Death Dis. 2019;10(3):164. https://doi.org/10.1038/s41419-019-1418-3.
- 124. Jadidi-Niaragh F, Atyabi F, Rastegari A, Kheshtchin N, Arab S, Hassannia H, et al. CD73-specific siRNA-loaded chitosan lactate nanoparticles potentiate the antitumor effect of a dendritic cell vaccine in 4T1 breast cancer-bearing mice. J Control Release. 2017;246:46-59. https://doi.org/10.1016/j. jconrel.2016.12.012.
- 125. Yang Z, Qi Y, Lu C, Zhang J, Luo R, Kang S. Small interfering RNA (siRNA)-mediated knockdown of Notch1 suppresses tumor growth and enhances the effect of IL-2 immunotherapy in malignant melanoma. J BUON. 2015;20(6):1553-64.
- 126. Ahn YH, Hong SO, Kim JH, Noh KH, Song KH, Lee YH, et al. The siRNA cocktail targeting interleukin 10 receptor and transforming growth factor-β receptor on dendritic cells potentiates tumour antigen-specific CD8(+) T cell immunity. Clin Exp Immunol. 2015;181(1):164-78. https:// doi.org/10.1111/cei.12620.
- 127. Khairuddin N, Gantier MP, Blake SJ, Wu SY, Behlke MA, Williams BR, McMillan NA. siRNA-induced immunostimulation through TLR7 promotes antitumoral activity against HPV-driven tumors in vivo. Immunol Cell Biol. 2012;90(2):187-96. https://doi.org/10.1038/ icb.2011.19.
- 128. Alshamsan A, Haddadi A, Hamdy S, Samuel J, El-Kadi AO, Uludağ H, Lavasanifar A. STAT3 silencing in dendritic cells by siRNA polyplexes encapsulated in PLGA nanoparticles for the modulation of anticancer immune response. Mol

Pharm. 2010;7(5):1643-54. https://doi.org/10.1021/ mp100067u.

- 129. Geng Y, Zheng X, Hu W, Wang Q, Xu Y, He W, et al. Hsa_circ_0009361 acts as the sponge of miR-582 to suppress colorectal cancer progression by regulating APC2 expression. Clin Sci (Lond). 2019;133(10):1197-1213. https://doi.org/10.1042/CS20190286.
- 130. Zhang L, Song X, Chen X, Wang Q, Zheng X, Wu C, Jiang J. Circular RNA CircCACTIN promotes gastric cancer progression by sponging miR-331-3p and regulating TGFBR1 expression. Int J Biol Sci. 2019;15(5):1091-3. https://doi.org/ 10.7150/ijbs.31533.
- 131. Di Martino MT, Riillo C, Scionti F, Grillone K, Polerà N, Caracciolo D, et al. miRNAs and lncRNAs as novel therapeutic targets to improve cancer immunotherapy. Cancers (Basel). 2021;13(7):1587. https://doi.org/10.3390/cancers13071587.
- 132. Wang S, Zhou S, Liu H, Meng Q, Ma X, Liu H, et al. ncRI: a manually curated database for experimentally validated non-coding RNAs in inflammation. BMC Genomics. 2020;21(1):380. https://doi.org/10.1186/s12864-020-06794-6.
- 133. Wei SC, Duffy CR, Allison JP. Fundamental mechanisms of immune checkpoint blockade therapy. Cancer Discov. 2018;8(9):1069-1086. https://doi.org/10.1158/2159-8290. CD-18-0367.
- 134. Smolle MA, Calin HN, Pichler M, Calin GA. Noncoding RNAs and immune checkpoints-clinical implications as cancer therapeutics. FEBS J. 2017;284(13):1952-1966. https://doi.org/10.1111/febs.14030.
- 135. Sadreddini S, Baradaran B, Aghebati-Maleki A, Sadreddini S, Shanehbandi D, Fotouhi A, Aghebati-Maleki L. Immune checkpoint blockade opens a new way to cancer immunotherapy. J Cell Physiol. 2018;234:8541-8549. https:// doi.org/10.1002/jcp.27816.
- 136. Zhang XL, Xu LL, Wang F. Hsa_circ_0020397 regulates colorectal cancer cell viability, apoptosis, and invasion by promoting the expression of the miR-138 targets TERT and *PD-L1*. Cell Biol Int. 2017;41(9):1056-1064. https://doi. org/10.1002/cbin.10826.
- 137. Schietinger A, Greenberg PD. Tolerance and exhaustion: defining mechanisms of T cell dysfunction. Trends Immunol. 2014;35(2):51-60. https://doi.org/10.1016/j.it.2013.10.001.
- 138. Baruch EN, Berg AL, Besser MJ, Schachter J, Markel G. Adoptive T cell therapy: an overview of obstacles and opportunities. Cancer. 2017;123(S11):2154-62. https://doi. org/10.1002/cncr.30491.
- 139. Sadeghzadeh M, Bornehdeli S, Mohammadrezakhani H, Abolghasemi M, Poursaei E, Asadi M, et al. Dendritic cell therapy in cancer treatment: the state-of-the-art. Life Sci. 2020;254:117580. https://doi.org/10.1016/j.lfs.2020.117580.
- 140. Zhang P, Wu W, Chen Q, Chen M. Non-coding RNAs and their integrated networks. J Integr Bioinform. 2019;16(3):20190027. https://doi.org/10.1515/jib-2019-0027.
- 141. Gokita K, Inoue J, Ishihara H, Kojima K, Inazawa J. Therapeutic potential of LNP-mediated delivery of miR-634 for cancer therapy. Mol Ther Nucleic Acids. 2020;19:330-338. https://doi.org/10.1016/j.omtn.2019.10.045.



This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.