# **RESEARCH ARTICLE**

# Association of Tumor Suppressor *TP53* and *TP21* Gene Polymoprhisms with Radiotherapy Induced Adverse Reactions in Head and Neck Cancer Patients

Anand K. Gudur<sup>1</sup>, Rashmi A. Gudur<sup>1</sup>, Suresh J. Bhosale<sup>1</sup>, Kailas D. Datkhile<sup>2\*</sup>

### Abstract

Background: The current study was intended to analyze the genotype distribution of the tumor suppressor genes TP53 and TP21, and to investigate their potential association with acute radiotherapy –induced toxicities, such as skin reactions and mucositis, in normal tissues of head and neck cancer (HNC) patients receiving radiotherapy. Materials & Methods: Two hundred and fifty HNC patients undergoing radiotherapy were enrolled in this study and the acute toxicity reactions and radiotherapy response were recorded. The potential association of two single nucleotide polymorphisms (SNPs) (rs1042522, rs28934571) of TP53 gene and (rs1801270, rs1059234) SNPs of TP21 gene, with the risk of acute skin toxicity reactions was analyzed using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) and direct DNA sequencing methods. Results: The findings revealed that the homozygous recessive (C/C) genotype of the 72G>C polymorphism in the TP53 gene exhibited a significantly increased association with acute skin toxicity in (HNC) patients undergoing radiotherapy (OR = 4.32, 95% CI: 1.87–10.01; p = 0.0006, and the heterozygous (G/C) genotype of the same polymorphism demonstrated a robust correlation with acute skin reactions (OR = 9.23, 95% CI: 4.40–19.23; p < 0.0001). The heterozygous variant (G/C) genotype of TP53 was identified as a significant risk factor for oral mucositis exceeding grade 2 severity in HNC patients receiving radiotherapy, with an odds ratio of 4.15 (95% CI: 2.21–7.76; p < 0.0001). Conclusion: The results of genetic polymorphisms of TP53 and TP21 genes and their association with radiotherapy induced acute toxicities demonstrated significant association of TP53, G72C polymorphism of rs1042522 SNP with both acute radiotherapy induced dermatitis and mucositis.

Keywords: Head and neck cancer- Radiotherapy- TP53- TP21- Genetic polymorphism- acute toxicity

Asian Pac J Cancer Prev, 26 (7), 2499-2509

## Introduction

Head and Neck cancer (HNC) is the most prevalent cancer in many countries of the world, accounting for approximately 900, 000 cases and over 450, 000 deaths annually [1]. The incidence of HNC has increased significantly worldwide in past few decades and represents a leading cause of cancer-related deaths among both men as well as women in Asian countries like China, Pakistan, Thailand and India [2]. In the Indian subcontinent, HNC has emerged as a major public health burden, representing approximately 30% of all cancers and contributing to 245,811 new cases and 130,139 deaths in 2022 [3-4]. Established risk factors for HNC include the use of tobacco in various forms, excessive alcohol consumption, poor oral hygiene, and exposure to environmental carcinogens [5-6]. Along with, , genetic susceptibility plays a critical role in HNC pathogenesis, involving complex, multistage processes mediated by alterations in DNA repair genes, tumor suppressor genes, oxidative stress-related genes, and epigenetic modifications [7-9]. Radiotherapy (RT) remains a cornerstone of HNC treatment, either as monotherapy or in combination with chemotherapy; however, radiation-induced toxicities in surrounding normal tissues significantly impact patients' quality of life [10-11]. The most common acute radio-toxicities occurred in the HNC patients are oral mucositis, dermatitis and dysphagia, while late toxicities include fibrosis, osteoradionecrosis or xerostomia [12-13]. Emerging evidence highlights the pivotal role of genetic factors in determining individual clinical radiosensitivity and susceptibility to radiation-induced adverse effects [14-15].

Several studies have investigated the role of genetic polymorphisms in tumor suppressor genes in predicting radiosensitivity and the development of normal tissue toxicity following radiotherapy. Polymorphisms such

<sup>1</sup>Department of Oncology, Krishna Vishwa Vidyapeeth "Deemed to be University", Taluka-Karad, Dist- Satara, Pin-415 539, (Maharashtra), India. <sup>2</sup>Department of Molecular Biology & Genetics, Krishna Institute of Science and Technology, Krishna Vishwa Vidyapeeth "Deemed to be University", Taluka-Karad, Dist- Satara, Pin-415 539, (Maharashtra) India. \*For Correspondence: hodgeneticslab@kvv.edu.in

as Arg72Pro and Arg249Ser in the TP53 gene have been extensively associated with an increased risk of various cancers; however, their specific contribution to radiotherapy-induced normal tissue reactions has not been thoroughly elucidated. Nevertheless, emerging evidence suggests that the TP53 Pro72Arg polymorphism, in conjunction with the p21 Ser31Ser genotype, may significantly influence the susceptibility to acute radiotherapy-induced toxicities in cancer patients [16-18]. Previous studies have also demonstrated that Arg72Pro SNP (rs1042522) in the TP53 gene influences radiosensitivity in different cancer patients and contributes to develop adverse effects following radiotherapy in patients with prostate [19], breast [20] gastric [21], lung [22] and head and neck cancers [23-25]. However, the relationship between radiotherapy induced toxicities and genetic polymorphisms in tumor suppressor genes remains controversial which showed no independent effect of TP53 polymorphisms with the risk of normal tissue toxicity following radiotherapy in breast [26-27] and prostate cancer patients [28-29]. Nevertheless, the combined genotypic effects of TP53 and TP21 polymorphisms have been implicated in increased susceptibility to acute radiotherapy induced toxicities in breast cancer patients. In contrast, no significant association of p21 polymorphisms and radiation-induced toxicity was observed in patients with non-small cell lung carcinoma [30]. Although numerous studies have investigated the role of genetic polymorphisms in radiation-responsive genes, there remains significant scope to further elucidate the association between specific gene variants and radiotherapy-induced normal tissue toxicity, particularly in HNC patients. Therefore, in the present study, we examined the potential association between two single nucleotide polymorphisms (SNPs) of the TP53 gene Pro72Arg (G>C; rs1042522) and Arg249Ser (G>T; rs28934571) and two SNPs of the TP21 gene C98A (rs1801270) and C70T (rs1059234) with the risk of developing acute skin toxicity following therapeutic radiotherapy in HNC patients.

### **Materials and Methods**

### Patient enrollment and Clinical Information

A total of 250 patients, histopathologically confirmed with HNC and visiting to Medical Oncology Out Patient Department (OPD) for the treatment at the Krishna Institute of Medical Sciences were enrolled in accordance based on predefined inclusion and exclusion criteria. Eligible participants included individuals aged between 25 to 85 years diagnosed with HNC on histopathology and no evidence of metastatic disease at the time of enrollment.

### *Treatment of HNC patients with radiotherapy & chemoradiotherapy*

All HNC patients were treated using threedimensional conformal radiation therapy (3DCRT) or Intensity modulated radiation therapy (IMRT), based on computed tomography (CT)-guided planning, simulation, verification, and rigorous quality assurance protocols. Patients were treated using Linear accelerator (Model: Unique Performance, Make: Varian Medical System, USA) 6-Mega Volt (MV) (X-ray) with the total radiotherapy dose ranging from 60- 66 Gy (2 Gy per fractions for 5 days a week) with volumetric modulated arc therapy (VMAT) technique. Patients after surgical resection having positive margins were given a dose of 66 Gy in 33 fractions. Concurrent chemotherapy was added where clinically indicated, using cisplatin at a dose of 40 mg/m2, administered weekly for up to 6 cycles in combination with radiotherapy.

### Follow up and Toxicity Assessment

To assess acute normal tissue toxicity, patients were prospectively followed for three months post-radiotherapy to evaluate clinical outcomes, including partial, complete, or no response, disease stability or progression, early mortality, and treatment-related toxicity. The facial and cervical skin regions were designated as target areas for monitoring acute radiation-induced skin reactions. Acute adverse effects, specifically oral mucositis and skin reactions, were documented weekly during radiotherapy at 1 and 3 months post-treatment, according to the Radiation Therapy Oncology Group (RTOG) criteria. Acute radiation toxicity was defined as any injury manifesting from the initiation of radiotherapy up to three months post-treatment. Severity grading of radiation-induced dermatitis and oral mucositis was performed by radiation oncologists using the RTOG scale. Patients presenting with grade >2 skin reactions or mucositis were classified as radiosensitive and compared against patients with grade  $\leq$ 2 toxicity to investigate associations with polymorphisms in tumor suppressor genes. All patient information was systematically recorded, and participants were monitored for a minimum of three months following therapy. Written informed consent was obtained from all participants prior to enrollment. The study protocol was reviewed and approved by the Institutional Ethics Committee of Krishna Institute of Medical Sciences.

#### Blood sample collection and genomic DNA isolation

Five milliliter (mL) of whole blood from patients was collected in sterile EDTA containing vacutainer after receiving informed consent. The blood sample from patients was collected before initiation of radiotherapy treatment. The genomic DNA extraction was carried out by salting out method where the whole blood was processed with lysis buffer-1 containing 10mM Tris-HCl pH-7.6, 320mM sucrose, 5mM MgCl<sub>2</sub>, 1% triton X-100, pH 7.6 to lyse RBCs, thereafter the sample was treated with the lysis buffer 2 to lyse out WBCs (10mM Tris-HCl, 11.4mM sodium citrate, 1mM EDTA, 1% SDS, pH-8.0). The sample was further treated with Proteinase K (200µg/µl) to digest the proteins and subsequently RNase A (200µg/ µl). The genomic DNA was precipitated by addition of twice the volume of ice cold ethanol and 1/10th volume of 3M Sodium acetate (pH-5.2). The precipitated DNA was aggregated together by centrifugation. The obtained pellet of DNA was then resuspended in T10E1 buffer and was checked on 1% agarose gel for its quality and quantity. This purified DNA was used for further genotyping assays after quantitative and qualitative analysis.

*Genotyping assays of Tumor suppressor (TP53 & TP21)* genes

The genotyping assay of tumor suppressor genes (p53, p21), was performed by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) using appropriate primer set as shown in Table 1. The PCR conditions for amplification of TP53 (309 bp) codon -72, exon-4, Pro72Arg, G>C) and codon249 of exon-7 with Arg249Ser (G>T) polymorphisms and TP21 with codon 31 of exon-2, C98A (272bp) and TP21 at exon 3 (C70T) polymorphisms with 300 bp are depicted in Table 1. After performing PCR programme for each of the reaction, the PCR products were analyzed by agarose gel electrophoresis in Tris-Acetate-EDTA (TAE) buffer. The agarose gels stained with ethidium bromide (10 mg/mL) and visualized under UV Transilluminator and photographed in gel documentation system (BioRad Laboratories).

#### Restriction Fragment Length Polymorphism

After confirmation of DNA amplification, each PCR product was digested with an appropriate restriction enzyme for genotyping. Ten micro liters of the PCR products digested at 37°C overnight with specific restriction enzymes in 20  $\mu$ L reaction mixtures containing buffer supplied with each restriction enzyme (Table1). After the overnight incubation, digestion products were separated on a 2-3% low EEO agarose (GeNei) gel at 100 V for 30 min stained with ethidium bromide and photographed with Gel Documentation System.

### Statistical Analysis

The genotypic frequencies for tumor suppressor genes in the patient's were determined. The relative risk and odds ratios (OR) with corresponding 95% confidence intervals (CI) were calculated using unconditional multiple logistic regression analysis to assess the association between acute toxicity grades and SNPs. Statistical significance was defined as a p-value less than 0.05. Acute post-radiotherapy adverse events were categorized based on the severity of skin reactions and oral mucositis, with events classified as grade >2 considered clinically significant. The patients with oral mucositis grade >2 are radiosensitive groups (cases) compared with  $\leq 2$  grade (controls) for determining their association with polymorphism of different tumor suppressor genes. All statistical analyses were performed using SPSS version 11.0 software.

### Results

# Demographic and Clinical characteristics of study population

A total of 250 patients, aged 25 to 85 years (median age: 55 years), were enrolled in the study. Of these, 187 were male (74.80%) and 63 were female (25.20%). The majority of patients were tobacco smokers (87.60%) compared to non-smokers (12.40%). When stratified by the primary site of cancer, the distribution was as follows: oropharynx (36.0%), hypopharynx (22.0%), oral cavity (23.60%), nasopharynx (2.80%), larynx (8.0%), and

other sites (4.80%). Of the total cohort, 26% underwent radiotherapy alone, while 74% received a combination of radiation therapy and chemotherapy. Regarding tumor size, 65.60% of patients presented with tumors larger than 2 cm, while 34.40% had tumors  $\leq$  2 cm. Among the 250 patients treated with radiotherapy, 63 (25.20%) exhibited grade >2 (grade 3) severe skin reactions, including intense erythema, moderate edema, patchy moist desquamation, increased pain, yellow exudate secretion, itching, and skin tightening. The remaining 187 (74.80%) patients reported grade  $\leq 2$  skin reactions, such as faint erythema, itching, and skin tightening. Additionally, 132 (52.80%) patients exhibited grade >2 (grade 3 or 4) oral mucositis, characterized by severe pain, fibrinous mucositis, ulceration, hemorrhage, and necrosis. In contrast, 118 (47.20%) patients experienced grade  $\leq 2$  mucositis, with symptoms including irritation, patchy mucositis, serosanguineous discharge, and moderate pain in response to radiation exposure.

# *Genotype Distribution of TP53 and TP21 genes and radiotherapy toxicity in HNC patients*

The tumor suppressor TP53 and TP21 were analyzed to assess the association of polymorphisms with genes radiotherapy-induced normal tissue toxicity. Univariate logistic regression analysis of the polymorphisms in TP53 and TP21 revealed a significant association between the TP53 G72C polymorphism (rs1042522 SNP) and both acute skin reactions and mucositis (Table 2). The analysis indicated that the homozygous recessive (C/C) genotype of the TP53 72G>C polymorphism was associated with a 4.32-fold increased risk of acute skin reactions (OR=4.32, 95% CI: 1.87-10.01; p=0.0006), while the heterozygous (G/C) genotype was linked to a 9.23-fold increased risk (OR=9.23, 95% CI: 4.40-19.23; p<0.0001) in HNC patients following radiotherapy. Additionally, the univariate analysis indicated that the odds ratio for severe oral mucositis (grade >2) in patients with the recessive allele of the TP53 gene was 3.54 times higher (95% CI: 1.74–7.14, p=0.0005) (Table 2). The heterozygous (G/C) genotype of TP53 also showed a 4.15-fold increased risk of severe oral mucositis (OR=4.15, 95% CI: 2.21-7.76; p<0.0001) in HNC patients exposed to radiotherapy. However, no significant association was observed between the rs28934571 SNP of TP53 and the rs1801270 and rs1059234 SNPs of TP21 with normal tissue toxicity, specifically regarding skin reactions or mucositis, as recessive and heterozygous genotypes did not show significantly higher frequencies in affected patients.

# Association of TP53 and TP21 gene polymorphisms with risk of toxicity effects of radiotherapy in HNC patients

The logistic regression analysis in the current study revealed no significant association between the 249G>T polymorphism of *TP53* and the 98C>A and 70C>T polymorphisms of *TP21* with skin reactions in HNC patients following radiotherapy (Table 3). The multivariate analysis produced odds ratios (ORs) with 95% confidence intervals (CIs) for patients experiencing acute dermatitis with the recessive allele of *TP53* (rs28934571), which showed an OR of 1.10 (95% CI: 0.54–2.23; p=0.783), and

<i>TP21</i> rs1055 <i>exon-3</i>	TP21 rs180) codon-31 exon-2 C98A	TP53 rs2893 Codon-249 Exon-7 Arg249Ser	TP53 rs104; Codon-72 Exon-4 Pro72Arg	Gene rs Genotype numl	SNP, Single nucleotide pc Table 1. The List of (		(rs1059234)	TP21		(rs1801270)	C98A	TD2 1	(rs28934571)	G249T	TP53	(131072322)	G72C	TP53		Gene Name	Table 2. Univariance $r$
9234 (C>T)	1270 (C>A)	4571 (G>T)	2522 (G>C)	Amino acid ber nucleotide change	Olymorphism; OR, Oc Candidate Tumor	C/T +T/T	T/T	C/C	C/A + A/A	A/A	C/A		G/T +T/T	G/T	G/G	G/C+C/C	0/D 35	G/G		Genotype	Analysis or roryn
FP: 5'-CCC RP: 5'-GGG	FP: 5'-GTC RP: 5'-CTC	FP: 5'-GGC RP: 5'-GGG	FP: 5'-TTC. RP: 5'- CTC		lds ratio; CI, Con r Suppressor (	53	6	134	41	1	40 40	116	30 73	43	114	67	23 23	120	reaction ≤2 n=187	Skin	TO SHISHIG TO
AGG GAA GGG T CGG CCA GGG 1	AGA ACC GGC TO CTC CCA ACT C/	GAC AGA GCG A TCA GCG GCA A	ACC CAT CTA CA AGG GCA ACT C	Primer Sequen Forward/Reve	fidence interval; Sig Genes ( <i>TP53</i> and	15	4 1	48	13	1	12	<b>2</b> 0	13 20	7	43	50	1 S	13	>2 n=03	Skin reaction	I UITIOF Suppress
GT CCT-3' CAT GTA-3'	GG GGA TG-3' AT CCC GG-3'	.GATTC CA -3' .GCAGA GG -3'	G TCC -3' }AC CGT-3'	rse	nificance p< 0.05; *, In   <i>TP21</i> ) Genes with	0.79 (0.40-1.53)	0.65 (0.31-1.36) 1.86 (0.50-6.87)	1 (Reference)	0.92 (0.45-1.86)	2.92 (0.17-47.55)	0.87 (0.42-1.80)	$\frac{1}{(\text{Poference})}$	1.14 (0.54-2.40)	0.43 (0.18-1.03)	1 (Reference)	6.88 (3.49-13.59)	9.23 (4.40-19.33)	1 (Reference)	10 %CK	OR	or <i>Troo</i> , and <i>Trai</i>
950C- 5 min, 30 cycles o 950C- 20 sec, 580C- 20 se	950C- 5 min, 30 cycles o 950C- 30 sec, 580C- 30 se 720C-30 sec, 720C- 10 mi	950C- 5 min, 30 cycles o 950C- 20 sec, 600C-20 sec 720C-20 sec, 720C- 10 mi	950C- 5 min, 30 cycles o 950C- 30 sec, 520C-30 sec 720C-30 sec, 720C- 10 mi	PCR Conditions	licates significant Odds Ratio Details of PCR and RF	0.485	0.256		0.829	0.451	0.718	0.201	0.713	0.059		<0.0001*	<0.0001*	0 0 0 -		p value Or	Offics alle Naulauoli II
300 bp	272 bp c, n	286 bp n	n 309 bp	PCR product size	(p<0.05); p val LP Procedui	30	5	88	25	1	24 24	0/ 1	17 44	27	74	35	20 15	83	52 n=118	al mucositis	
1 U of PstI 370C Incubation for 16 bre	1 U of Bpul (Blpl) 370C Incubation for 16 hrs	1 U of HaeIII (BsuRI) 370C Incubation for 16 hrs	1 U of Bstul (Bsh12361) 370C Incu- bation for 16 hrs	Enzyme / Digestion conditions	ue determined based on $\chi^2$ es Including Primers a	38	5	94	29	1	102 28	101 7	26 40	23	83	82 82	20 20	50	>2 n=132	Oral mucositis	NEACTIONS AND IMPROVED
126bp, 174bp	183 bp, 89 bp	92bp, 66bp, Small fragments	175bp, 134bp	Dominant (Wild type)	nd Restricti	1.18 (0	0.93 (0	1 (Re	1.06 (0	0.92 (0.	1.07 (0	0.22 (0	0.83 (0	0.75 (0	1(Ref	3.88 (2	4.15 (2	1(Ref	CK		IS III IIEAU a
300 bp, 174 bp, 126 hn	272 bp, 183 bp, 89 bp		309bp, 175bp, 134bp	Hetero- zygous	on Enzyme	.67-2.07)	.26-3.34)	ference)	.58-1.95)	05-14.94)	.58-1.98)		59-1.77)	.40-1.43)	erence)	.29-6.60)	.21-7.76)	erence)	% CI	OR	THU INCCA C
300 bp	272bp	158bp, 92bp, 66bp, Small fragments	309bр	Recessive (Mutant)	S	0.55	0.486 0.919		0.828	0.954	0.816	0.570	0.647	0.398		< 0.0001*	<0.0001*	0 0 0		p value	alleer Faurins

Anand K. Gudur et al

**2502** Asian Pacific Journal of Cancer Prevention, Vol 26

Gene /SNP	Genotypes	All Patients	Radiosensitive patients	OR 95% CI	p value
TP53	G/G	133	13	1 (Reference)	
(rs1042522)	G/C	70	35	5.11 (2.54-10.29)	< 0.0001*
	C/C	47	15	3.26 (1.44-7.36)	0.004*
	G/C +C/C	117	50	4.37 (2.26-8.44)	< 0.0001*
TP53	G/G	157	43	1 (Reference)	
(rs28934571)	G/T	50	7	0.51 (0.21-1.20)	0.126
	T/T	43	13	1.10 (0.54-2.23)	0.783
	G/T +T/T	93	20	0.78 (0.43-1.41)	0.421
TP21	C/C	196	50	1 (Reference)	
(rs1801270)	C/A	52	12	0.90 (0.44-1.82)	0.779
	A/A	2	1	1.96 (0.17-22.05)	0.585
	C/A+A/A	54	13	0.84 (0.47-1.86)	0.867
TP21	C/C	182	48	1 (Reference)	
(rs1059234)	C/T	58	11	0.71 (0.35-1.47)	0.368
	T/T	10	4	1.51 (0.45-5.04)	0.497
	C/T +T/T	68	15	0.83 (0.43-1.59)	0.586

Table 3. Association of Polymorphisms of Tumor Suppressor *TP53* and *TP21* Genes with Risk of Skin Reaction after Radiotherapy in Head and Neck Cancer Patients

SNP, Single nucleotide polymorphism; OR, Odds ratio; CI, Confidence interval; Significance p < 0.05; \*, Indicates significant Odds Ratio (p < 0.05); p value determined based on  $\chi^2$ .

for the heterozygous G/T genotype, which showed an OR of 0.51 (95% CI: 0.21–1.20; p=0.126). Both associations were non-significant with respect to acute skin reactions in HNC patients. The ORs for the heterozygous variant alleles of *TP21* rs1801270 (OR=1.96, 95% CI: 0.17–22.05; p=0.585) and rs1059234 (OR=1.51, 95% CI: 0.45–5.04; p=0.497) also indicated no significant association with skin toxicity following radiotherapy in HNC patients. However, the odds ratio for patients with grade >2 oral mucositis showed a significant increase associated with

the recessive (C/C) genotype of *TP53* (rs1042522) with an OR of 3.26 (95% CI: 1.44–7.36; p=0.004) and the heterozygous (G/C) genotype with an OR of 5.11 (95% CI: 2.54–10.29; p<0.0001). The multivariate logistic regression analysis also evaluated the association of tumor suppressor genes, particularly radiosensitive genes, with oral mucositis in HNC patients treated with radiotherapy. The OR for patients experiencing grade >2 oral mucositis with the recessive (C/C) genotype of *TP53* (rs1042522) was 1.81 (95% CI: 1.04–3.15; p=0.035), and

Table 4. Association of Polymorphisms of Tumor Suppressor *TP53* and *P21* Genes with Risk of Mucositis after Radiotherapy in Head and Neck Cancer Patients

Gene /SNP	Genotypes	All Patients	Radiosensitive patients	OR 95% CI	p value
TP53	G/G	133	50	1 (Reference)	
(rs1042522)	G/C	70	50	1.90 (1.16-3.09)	0.009*
	C/C	47	32	1.81 (1.04-3.15)	0.035*
	G/C+C/C	117	82	1.86 (1.21-2.86)	0.004*
TP53	G/G	157	83	1 (Reference)	
(rs28934571)	G/T	50	23	0.87 (0.49-1.52)	0.626
	T/T	43	26	1.14 (0.65-1.99)	0.635
	G/T +T/T	93	49	0.99 (0.64-1.54)	0.987
TP21	C/C	196	102	1 (Reference)	
(rs1801270)	C/A	52	28	1.03 (00.61-1.73)	0.897
	A/A	2	1	0.96 (0.08-10.72)	0.974
	C/A+A/A	54	29	1.03 (0.61-1.71)	0.903
TP21	C/C	182	94	1 (Reference)	
(rs1059234)	C/T	58	33	1.10 (0.67-1.80)	0.701
	T/T	10	5	0.96 (0.32-2.91)	0.954
	C/T+T/T	68	38	1.08 (0.67-1.72)	0.741

SNP, Single nucleotide polymorphism; OR, Odds ratio; CI, Confidence interval; Significance p < 0.05; \*, Indicates significant Odds Ratio (p < 0.05); p value determined based on  $\chi^2$ .

THOID DI LUDUCIUMUL	Come of the offer	C C L C L C C C C C C C C C C C C C C C	A T TO COL T		0				
Gene Name	Genotype	Tumor	stage	OR 95% CI	p value	Histolog	ical Grade	OR	p value
(SNP)		T1, T2	T3, T4			Ι, Π	III, IV	95% CI	
		n=134	n=116			n=103	n=147		
TP53	G/G	80	53	1 (Reference)		58	75	1(Reference)	
G72C	G/C	35	35	1.50 (0.84-2.70)	0.166	30	40	1.03 (0.57-1.84)	0.918
(rs1042522)	C/C	19	28	2.22 (1.12-4.38)	0.020*	15	32	1.64 (0.817-3.33)	
	G/C+ C/C	54	63	0.76 (1.06-2.91)	0.027*	45	72	1.23 (0.74-2.05)	0.409
TP53	G/G	85	72	1 (Reference)		66	91	1(Reference)	
G249T	G/T	28	22	0.92 (0.48-1.76)	0.818	20	30	1.08(0.56-2.08)	0.799
(rs28934571)	T/T	21	22	1.23 (0.62-2.42)	0.537	17	26	1.10 (0.55-2.20)	0.767
	G/T+ T/T	49	44	1.06 (0.63-1.77)	0.823	37	56	1.09 (0.65-1.85)	0.726
TP21	C/C	106	90	1 (Reference)		87	109	1(Reference)	
C98A	C/A	28	24	1.00 (0.54-1.86)	0.975	14	38	2.16 (1.10-4.25)	0.024*
(rs1801270)	A/A	0	2	5.88 (0.27-124.15)	0.254	2	0	0.15 (0.007-3.37)	0.238
	C/A+A/A	28	26	1.09 (0.59-1.99)	0.771	16	38	1.89 (0.99-3.62)	0.053
TP21	C/C	95	87	1 (Reference)		73	109	1 (Reference)	
C70T	C/T	33	25	0.82 (0.45-1.50)	0.532	24	34	0.94 (0.52-1.73)	0.863
(rs1059234)	T/T	9	4	0.72 (0.19-2.66)	0.631	6	4	0.44 (0.12-1.63)	0.223
	C/T+T/T	39	29	0.81 (0.46-1.42)	0.467	30	38	0.84(0.48-1.48)	0.566

**2504** Asian Pacific Journal of Cancer Prevention, Vol 26

for the heterozygous (G/C) genotype, it was 1.90 (95% CI: 1.16–3.09; p=0.009), both indicating significant associations with oral mucositis toxicity following radiotherapy in HNC patients (Table 4). Therefore, no significant association was observed between genetic variants of TP53 (rs28934571) and TP21 (rs1801270, rs1059234) and the development of increased acute skin toxicity or oral mucositis reactions after radiotherapy.

### Association of TP53 and TP21 gene polymorphisms with tumor and node response towards radiotherapy in HNC patients

Univariate logistic regression analysis was performed to evaluate the association between TP53 (rs1042522, rs28934571) and TP21 (rs1801270, rs1059234) gene polymorphisms with clinically and histopathologically confirmed tumor grades and tumor response to radiotherapy, as presented in Tables 5 and 6. The univariate analysis revealed that the rs1042522 SNP of the TP53 gene was not associated with histological tumor grades >III, and the rs28934571 SNP showed no association with either clinical or histopathological tumor grades. However, the recessive (C/C) genotype of TP53 (rs1042522) was significantly associated with high tumor stage (stage >3), with an odds ratio (OR) of 2.22 (95% CI: 1.12–4.38; p=0.02). Similarly, the heterozygous C/A genotype of TP53 (rs1801270) was associated with histological tumor grade >3 (OR=2.16, 95% CI: 1.10-4.25; p=0.024), while the rs1059234 SNP of *TP21* did not show any significant association with clinical stage >3 or histopathological grade >III (Table 5). Regarding the relationship between TP53 and TP21 polymorphisms and tumor response to radiotherapy, no association was found between the rs1042522 and rs28934571 SNPs of TP53 and tumor or nodal response to radiotherapy, as shown in Table 6. Furthermore, the C98A polymorphism of TP21 (rs1801270) showed that neither the recessive nor the heterozygous genotypes were associated with tumor or nodal response in HNC patients following radiotherapy, evaluated three months post-treatment. A significant positive association was observed for the heterozygous (C/T) genotype of the TP21 (rs1059234) SNP with complete tumor response to radiotherapy (OR=2.19, 95% CI: 1.06–4.51; p=0.033), whereas the recessive T/T genotype was not associated with tumor or nodal response. Additionally, the C70T polymorphism of TP21, with the recessive T/T genotype (OR=3.47, 95% CI: 0.83–14.50; p=0.088) and the heterozygous C/T genotype (OR=1.68, 95% CI: 0.73-3.84; p=0.213), was not associated with nodal response in HNC patients to radiotherapy. When HNC patients were stratified based on body mass index (BMI), no significant association was found between any TP53 or TP21 gene polymorphisms and the risk of acute radiotherapy toxicity (Table 7).

### Discussion

Radiotherapy is a crucial treatment modality for HNC, typically administered in fractions to enhance treatment efficacy. However, radiotherapy can induce toxic reactions in normal tissues. Prominent toxicities

70

lable b. Association	i between Genotypes o	t Iumor Suppre	essor TP53 and	<i>IP21</i> Genes with Tumor a	nd Node Kespoi	nse in Head an	d Neck Cancer	Patients towards Radioth	ıerapy
Gene Name	Genotype	Tumor Re	sponse	Risk Ratio (RR)	p value	Node R	esponse	Risk Ratio (RR)	p value
(SNP)		CR	PR/NR	95% CI		CR	PR/NR	95% CI	
		n=209	n=41			n=217	n=33		
TP53	G/G	108	25	1 (Reference)		113	20	1(Reference)	
G72C	G/C	61	9	0.63 (0.27-1.45)	0.284	61	9	0.83 (0.35-1.94)	0.673
(TS1042322)	C/C	40	7	0.75 (0.30-1.88)	0.548	43	4	0.52 (0.16-1.62)	0.264
	G/C+ C/C	101	16	0.68 (0.34-1.35)	0.276	104	13	0.70(0.33-1.49)	0.361
TP53	G/G	130	27	1 (Reference)		134	23	1(Reference)	
G249T	G/T	41	9	1.05 (0.45-2.42)	0.896	43	7	0.94 (0.38-2.36)	0.909
(1764669781)	T/T	38	S	0.59(0.18-1.94)	0.394	40	ω	0.43 (0.12-1.53)	0.195
	G/T+ T/T	79	14	0.80 (0.32-2.02)	0.647	83	10	0.70 (0.31-1.54)	0.38
TP21	C/C	166	30	1 (Reference)		174	22	1(Reference)	
C98A	C/A	41	11	1.48 (0.68-3.20)	0.315	42	10	1.88 (0.82-4.27)	0.13
(077100151)	A/A	2	0	1.09 (0.05-23.30)	0.955	1	1	7.90 (0.47-130.99)	0.148
	C/A+A/A	43	11	1.41 (0.65-3.05)	0.375	43	11	2.02 (0.91-4.48)	0.083
TP21	C/C	157	25	1 (Reference)		162	20	1 (Reference)	
C70T	C/T	43	15	2.19 (1.06-4.51)	0.033*	48	10	1.68 (0.732-3.84)	0.213
(+67660181)	T/T	9	1	0.69 (0.08-5.74)	0.738	7	ω	3.47 (0.83-14.50)	0.088
	C/T+ T/T	52	16	1.93(0.95 - 3.89)	0.065	55	13	1.91 (0.89-4.10)	0.094
SNP, Single nucleotide p determined based on $\chi^2$	olymorphism; CR, Complete	e Response; PR, Pa	rtial Response; NR,	No Response; RR, Risk ratio; Cl	l, Confidence interv	al; Significance p<	< 0.05; *, Indicates :	significant Odds Ratio (p<0.05	), p value

Gene Name	Genotype	Normal Weight					Ονι	erweight	
(SNP)			(BMI)	≤20)			(BI	MI ≤20)	
		All Patients	<b>RS</b> patients	OR 95% CI	p value	All Patients	<b>RS</b> patients	OR 95% CI	p value
TP53	G/G	55	22	1 (Reference)		78	36	1 (Reference)	
G72C	G/C	33	10	0.75 (0.31-1.79)	0.528	47	17	0.78 (0.39-1.54)	0.482
(rs1042522)	C/C	23	16	0.71 (0.33-1.50)	0.375	33	14	0.91 (0.43-1.92)	0.823
	G/C+ C/C	56	26	1.16 (0.58-2.28)	0.667	80	31	0.83 (0.47-1.48)	0.549
TP53	G/G	76	32	1 (Reference)		81	36	1 (Reference)	
G249T	G/T	17	9	1.25 (0.50-3.11)	0.62	33	13	0.88 (0.41-1.88)	0.753
(TSZ87343/1)	T/T	18	9	1.18 (0.48-2.92)	0.708	25	18	1.62 (0.78-3.33)	0.19
	G/T+ T/T	35	18	1.22 (0.60-2.46)	0.576	58	31	1.20 (0.66-2.16)	0.537
TP21	C/C	83	36	1 (Reference)		113	54	1 (Reference)	
C98A	C/A	27	12	1.02 (0.46-2.24)	0.951	25	22	1.84 (0.95-3.55)	0.069
(TS1001270)	A/A	1	0	0.76 (0.03-19.16)	0.869	1	1	2.09 (0.12-34.09)	0.604
	C/A + A/A	28	12	0.98 (0.45-2.15)	0.976	26	23	1.85 (0.96-3.53)	0.062
TP21	C/C	80	43	1 (Reference)		102	48	1 (Reference)	
C70T	C/T	28	18	1.19 (0.59-2.40)	0.615	30	15	1.06 (0.52-2.15)	0.866
(TS1U39234)	T/T	З	2	1.24 (0.19-7.71)	0.817	7	ы	0.91 (0.22-3.67)	0.895
	C/T+ T/T	31	20	1.20 (0.61-2.35)	0.595	37	18	1.03 (0.53-1.99)	0.921

Anand K. Gudur et al

resulting from radiotherapy in HNC patients include subcutaneous fibrosis, oral mucositis, and skin reactions such as dermatitis, particularly following adjuvant radiation therapy or concurrent chemoradiotherapy. Host genetic factors play a significant role in determining an individual's susceptibility to radiation-induced adverse effects, and a better understanding of these factors could help mitigate the long-term consequences of radiotherapy. Single nucleotide polymorphisms (SNPs) in genes involved in the DNA repair pathway may influence the ability of adjacent cells to repair radiation-induced DNA damage, potentially leading to more severe toxicity. Additionally, polymorphisms in tumor suppressor genes may offer insights into their role in the radiation response during treatment. Although extensive research has been conducted on the genetic variants of tumor suppressor genes and their involvement in cancer development, limited literature exists regarding their association with radiotherapy outcomes. Therefore, identifying genetic variations in these genes is essential for personalizing therapy, improving treatment safety and outcomes, and addressing the potential reduction in radiotherapy efficacy in patients. Tumor suppressor proteins, such as p53 and p21, are key regulators of cellular responses and play essential roles in controlling cell growth and apoptosis in response to radiation-induced damage. Upon exposure to ionizing radiation, p53 is rapidly activated and facilitates G1 phase cell cycle arrest by transactivating the expression of *p21*, thereby inducing cell cycle checkpoint control [31]. Polymorphisms in the TP53 and TP21 genes have been associated with increased susceptibility to both acute and chronic radiation-induced toxicities across various cancers, although findings have been inconsistent. In this study, we aimed to investigate the polymorphisms of the tumor suppressor genes TP53 and TP21 and their potential association with radiotherapy-induced acute toxicities in HNC patients. Our results demonstrated that the Arg72Pro polymorphism of *p53* was significantly associated with an increased risk of acute radiation-induced toxicities, including dermatitis (OR=4.32, 95% CI: 1.87-10.01; p=0.0006) and mucositis (OR=3.54, 95% CI: 1.74-7.14; p=0.0005) following radiotherapy. These findings are consistent with previous studies that have linked p53 polymorphisms to adverse toxicity outcomes in HNC [32] and breast cancer [17] patients undergoing radiotherapy, either alone or in combination with chemoradiotherapy. Several studies have also reported an association between the p53 Arg72Pro polymorphism and the risk of skin toxicities in patients undergoing radiotherapy for breast [26], lung [22], and prostate cancer [29]. In the present study, no significant correlation was found between the p21 Ser31Arg polymorphism and the risk of acute toxicities, such as dermatitis or mucositis, which is consistent with other studies on *p21* polymorphisms in breast cancer [17, 33]. When examining the clinicopathological features and tumor and nodal response of head and neck cancer (HNC) patients to radiotherapy, as well as their association with p53 and p21 gene polymorphisms, our results confirmed that the recessive (C/C) genotype of TP53 (rs1042522) was associated with a higher tumor stage (p=0.02). Similarly, the heterozygous (C/T) genotype of TP21

(rs1059234) was significantly associated with a complete tumor response to radiotherapy (p=0.033).

Conclusion: The results obtained from this study evidenced a significant association between Arg72Pro polymorphism of the *TP53* gene with risk of acute radiation-induced toxicities, such as dermatitis and mucositis in HNC patients treated with radiotherapy. Specifically, the results suggest a significant correlation between *TP53* G72C polymorphism (rs1042522 SNP) with both acute dermatitis and mucositis. Further investigation of genetic variants and their association with radiotherapy response and its adverse toxicity effects is required to be out to confirm our findings by using large number of samples and broad range of SNPs for comprehensive genotyping.

### **Author Contribution Statement**

Concept: AKG, RAG, SJB, Design: AKG, KDD, RAG, Experimental Studies: AKG, RAG, KDD Clinical studies: AKG, RAG Data analysis: KDD, AKG, Statistical analysis: AKG, KDD Manuscript preparation: AKG, RAG, KDD, All authors read and approved the final manuscript.

### Acknowledgements

### Funding statement

Authors are thankful to Indian Council of Medical Research (ICMR) for financial assistance to the research project (Grant No. NCD/Ad-hoc/120/2021-22 dated 22/11/2021.

### Approval

The study protocol was approved by protocol committee of Krishna Vishwa Vidyapeeth (Deemed to be University)

### Ethics Committee Approval

The study protocol was approved by Institutional Ethics Committee of Krishna Vishwa Vidyapeeth (Deemed to be University), Karad.

### Declaration of Conflict of interest

The authors declare that they have no competing financial or any other conflict of interests that could have appeared to influence the work reported in this paper.

#### Abbreviations

HNC: Head and neck cancer *TP53*: Tumor suppressor 53 *TP21* Tumor suppressor 21 SNP: Single nucleotide polymorphism PCR-RFLP: Polymerase Chain Reaction-Restriction Fragment Length Polymorphism DNA: Deoxyribose Nucleic Acid OR: Odds ratio CI: Confidence interval RT: Radiotherapy OPD:Out Patient Department MV: Mega Volt

Asian Pacific Journal of Cancer Prevention, Vol 26 2507

VMAT: volumetric modulated arc therapy RTOG: Radiation Therapy Oncology Group mL: milliliter µl: Microliter EDTA: Ethylenediamdie Tetra acetate SDS: Sodium dodecyl Sulphate TAE: Tris-Acetate-EDTA

# References

- Global Cancer Observatory. International Agency for Research on Cancer. World Health Organization. Available from: https://gco.iarc.fr/ (Accessed on January 23, 2023).https://gco.iarc.who.int/media/globocan/factsheets/ populations/900-world-fact-sheet.pdf
- Chaturvedi AK, Anderson WF, Lortet-Tieulent J, Curado MP, Ferlay J, Franceschi S, et al. Worldwide trends in incidence rates for oral cavity and oropharyngeal cancers. J Clin Oncol. 2013;31(36):4550-9. https://doi.org/10.1200/ JCO.2013.50.3870.
- Sathishkumar K, Chaturvedi M, Das P, Stephen S, Mathur P. Cancer incidence estimates for 2022 & projection for 2025: Result from National Cancer Registry Programme, India. Ind J Med Res. 2022;156(4&5):598-607. https://doi. org/10.4103/ijmr.ijmr 1821 22.
- 4. Bagal S, Budukh A, Thakur JS, Dora T, Qayyumi B, KhannaD, et al. Head and neck cancer burden in India: an analysis from published data of 37 population-based cancer registries. Ecancer med sci. 2023;17:1603. https://doi.org/10.3332/ ecancer.2023.1603.
- Marcu LG, Yeoh E. A review of risk factors and genetic alterations in head and neck carcinogenesis and implications for current and future approaches to treatment. J Cancer Res Clin Oncol. 2009;135(10):1303-14. https://doi.org/10.1007/ s00432-009-0648-7.
- Hari Ram, Sarkar J, Kumar H, Konwar R, Bhatt ML, Mohammad S. Oral cancer: Risk factors and molecular pathogenesis. J Maxillofacial Oral Surg. 2011;10(2):132-7. https://doi.org/10.1007/s12663-011-0195-z.
- Fostira F, Koutsodontis G, Vagia E, Economopoulou P, Kotsantis I, Sasaki C, et al. Predisposing germline mutations in young patients with squamous cell cancer of the oral cavity. JCO Precision Oncol. 2018;2:1-8. https:// doi.org/10.1200/PO.18.00022.
- Vukovic V, Stojanovic J, Vecchioni A, Pastorino R, Boccia S. Systematic Review and Meta-analysis of SNPs from Genome-Wide Association Studies of Head and Neck Cancer. Otolaryngol-Head Neck Surg. 2018;159(4):615-24. https://doi.org/10.1177/0194599818792262.
- Campbell BR, Chen Z, Faden DL, Agrawal N, Li RJ, Hanna GJ, et al. The mutational landscape of early- and typicalonset oral tongue squamous cell carcinoma. Cancer. 2021;127(4):544-53. https://doi.org/10.1002/cncr.33309.
- Rodel C, Grabenbauer GG, Kuhn R, Papadopoulos T, Dunst J, Meyer M, et al. Combined-modality treatment and selective organ preservation in invasive bladder cancer: longterm results. J Clin Oncol. 2002;20(14):3061–71. https://doi. org/10.1200/JCO.2002.11.027.
- 11. Suk R, Gurubhagavatula S, Park S, Zhou W, Su L, Lynch TJ, et al. Polymorphisms in ERCC1 and grade 3 or 4 toxicity in non-small cell lung cancer patients. Clin Cancer Res. 2005;11(4):1534-8. https://doi.org/10.1158/1078-0432. CCR-04-1953.
- 12. Trotti A, Bellm LA, Epstein JB, Frame D, Fuchs HJ, Gwede CK, et al. Mucositis incidence, severity and associated outcomes in patients with head and neck cancer receiving

radiotherapy with or without chemotherapy: A systematic siterature seview. Radiother Oncol. 2003;66 (3):253–62. https://doi.org/10.1016/s0167-8140 (02)00404-8.

- Machtay M, Moughan J, Trotti A, Garden AS, Weber RS, Cooper JS, et al. Factors Associated With Severe Late Toxicity After Concurrent Chemoradiation for Locally Advanced Head and Neck Cancer: An RTOG Analysis. J Clin Oncol. 2008;26(21):3582–9. https://doi.org/10.1200/ JCO.2007.14.8841.
- 14. Andreassen CN, Dikomey E, Parliament M, West CM. Will SNPs be useful predictors of normal tissue radiosensitivity in the future? Radiother Oncol. 2012;105(3):283–8. https:// doi.org/10.1016/j.radonc.2012.11.003.
- Barnett GC, Coles CE, Elliott RM, Baynes C, Luccarini C, Conroy D, et al. Independent validation of genes and polymorphisms reported to be associated with radiation toxicity: a prospective analysis study. Lancet Oncol. 2012;13(1):65–77. https://doi.org/10.1016/S1470-2045(11)70302-3.
- 16. Vannini I, Zoli W, Tesei A, Rosetti M, Sansone P, Storci G, et al. Role of *p53* codon 72 arginine allele in cell survival in vitro and in the clinical outcome of patients with advanced breast cancer. Tumor Biol. 2008;29(3):145–51. https://doi. org/10.1159/000143400.
- Chang-Claude J, Ambrosone CB, Lilla C, Kropp S, Helmbold I, von Fournier D, et al. Genetic polymorphisms in DNA repair and damage response genes and late normal tissue complications of radiotherapy for breast cancer. Br J Cancer. 2009;100(10):1680-6. https://doi.org/10.1038/ sj.bjc.6605036.
- Su M, Yin ZH, Wu W, Li XL, Zhou BS. Meta-analysis of associations between ATM Asp1853Asn and *TP53* Arg72Pro polymorphisms and adverse effects of cancer radiotherapy. Asian Pac J Cancer Res. 2014;15(24):10675-81. https://doi. org/10.7314/apjcp.2014.15.24.10675.
- Cesaretti JA, Stock RG, Lehrer S, Atencio DA, Bernstein JL, Stone NN, et al. ATM sequence variants are predictive of adverse radiotherapy response among patients treated for prostate cancer. Int J Radiat Oncol Biol Phy. 2005;61(1):196-202. https://doi.org/ 10.1016/j.ijrobp.2004.09.031.
- 20. Andreassen CN, Overgaard J, Alsner J, Overgaard M, Herskind C, Cesaretti JA, et al. ATM sequence variants and risk of radiation-induced subcutaneous fibrosis after postmastectomy radiotherapy. Int J Radiat Oncol Biol Phys. 2006;64(3):776-83. https://doi.org/10.1016/j. ijrobp.2005.09.014.
- 21. Kim JG, Sohn SK, Chae YS. *TP53* codon 72 polymorphism associated with prognosis in patients with advanced gastric cancer treated with paclitaxel and cisplatin. Cancer Chemo Pharmacol. 2009;64(2):355–60. https://doi.org/10.1007/ s00280-008-0879-3.
- 22. Yang M, Zhang L, Bi N, Ji W, Tan W, Zhao L, et al. Association of P53 and ATM polymorphisms with risk of radiationinduced pneumonitis in lung cancer patients treated with radiotherapy. Int J Radiat Oncol Biol Phys. 2011;79(5):1402-7. https://doi.org/10.1016/j.ijrobp.2009.12.042.
- 23. Tu HF, Chen HW, Kao SY, Lin SC, Liu CJ, Chang KW. MDM2 SNP 309 and *p53* codon 72 polymorphisms are associated with the outcome of oral carcinoma patients receiving postoperative irradiation. Radiother Oncol. 2008;87(2):243-52. https://doi.org/10.1016/j.radonc.2008.03.018.
- 24. Xie X, Wang H, Jin H, Ouyang S, Zhou J, Hu J, et al. Expression of pAkt affects *p53* codon 72 polymorphism-based prediction of response to radiotherapy in nasopharyngeal carcinoma. Radiat Oncol. 2013;8:117. https://doi.org/10.1186/1748-717X-8-117.
- 25. Borchiellini D, Etienne-Grimaldi MC, Bensadoun RJ,

Benezery K, Dassonville O, Poissonnet G, et al. Candidate apoptotic and DNA repair gene approach confirms involvement of ERCC1, ERCC5, *TP53* and MDM2 in radiation-induced toxicity in head and neck cancer. Oral Oncol. 2017;67:70-6. https://doi.org/10.1016/j. oraloncology.2017.02.003.

- 26. Tan XL, Popanda O, Ambrosone CB, Kropp S, Helmbold I, von Fournier D, et al. Association between *TP53* and *p21* genetic polymorphisms and acute side effects of radiotherapy in breast cancer patients. Br Cancer Res Treat. 2006;97(3):255–62. https://doi.org/10.1007/s10549-005-9119-2.
- 27. Badie C, Dziwura S, Raffy C, Tsigani T, Alsbeih G, Moody J, et al. Aberrant CDKN1A transcriptional response associates with abnormal sensitivity to radiation treatment. Brazilian J Cancer. 2008;98(11):1845–51. https://doi.org/10.1038/ sj.bjc.6604381.
- Popanda O, Marquardt JU, Chang-Claude J, Schmezer P. Genetic variation in normal tissue toxicity induced by ionizing radiation. Mutation Res. 2009;667(1-2):58–69. https://doi.org/10.1016/j.mrfmmm.2008.10.014.
- Cintra HS, Pinezi JC, Machado GD, de Carvalho GM, Carvalho AT, dos Santos TE, et al. Investigation of genetic polymorphisms related to the outcome of radiotherapy for prostate cancer patients. Disease Mark. 2013;35(6):701-10. https://doi.org/10.1155/2013/762685.
- Matsuzoe D, Hideshima T, Kimura A, Inada K, Watanabe K, Akita Y, et al. *p53* mutations predict nonsmall cell lung carcinoma response to radiotherapy. Cancer Lett 1999;135(2):189–94. https://doi.org/10.1016/s0304-3835 (98)00292-4.
- Mazzatti DJ, Lee YJ, Helt CE, O'Reilly MA, Keng PC. p53 modulates radiation sensitivity independent of p21 transcriptional activation. Am J Clin Oncol. 2005;28(1):43– 50. https://doi.org/10.1097/01.coc.0000139484.51715.5a.
- 32. Sullivan A, Syed N, Gasco M, Bergamaschi D, Trigiante G, Attard M, et al. Polymorphism in wild-type *p53* modulates response to chemotherapy in vitro and in vivo. Oncogene. 2004;23(19):3328–37. https://doi.org/ 10.1038/ sj.onc.1207428.
- 33. Azzato EM, Driver KE, Lesueur F, Shah M, Greenberg D, Easton DF, et al. Effects of common germline genetic variation in cell cycle control genes on breast cancer survival: results from a population-based cohort. Br Cancer Res. 2008;10(3):R47. https://doi.org/10.1186/bcr2100.



This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.